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U.S. DISTRICT COURT
NORTHERN DISTRICT OF CALIFORNIA

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UNITED STATES DISTRICT COURT
NORTHERN DISTRICT OF CALIFORNIA

VALENT U.S.A. CORPORATION AND
SUMITOMO CHEMICAL CO. LTD.,

CV 08 0720
Case No.

**COMPLAINT FOR DECLARATORY
JUDGMENT OF PATENT INVALIDITY
AND NON INFRINGEMENT**

Plaintiffs,

v.

SYNGENTA CROP PROTECTION, INC.,

Defendants.

DEMAND FOR JURY TRIAL

Plaintiffs Valent U.S.A. Corporation ("Valent") and Sumitomo Chemical Company, Ltd., ("SCC"), by their undersigned attorneys, complain for declaratory judgment against Syngenta Crop Protection, Inc., ("Syngenta"), as follows.

I. NATURE OF THE ACTION

1. This civil action arises under the patent laws of the United States, 35 U.S.C. §§ 1 *et seq.*, and seeks a declaratory judgment pursuant to the Federal Declaratory Judgments Act, 28 U.S.C. §§ 2201-02. Valent and SCC seek a declaratory judgment of invalidity and non-infringement of U.S. Patent No. 7,105,469 B2, ("the '469 patent"), which is owned by Syngenta. A true and correct copy of the '469 patent is attached hereto as Exhibit A.

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12 UNITED STATES DISTRICT COURT
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14 NORTHERN DISTRICT OF CALIFORNIA

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16 SUMITOMO CHEMICAL CO. LTD.,

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25 *seq.*, and seeks a declaratory judgment pursuant to the Federal Declaratory Judgments Act, 28 U.S.C.
26 §§ 2201-02. Valent and SCC seek a declaratory judgment of invalidity and non-infringement of U.S.
27 Patent No. 7,105,469 B2, ("the '469 patent"), which is owned by Syngenta. A true and correct copy of
28 the '469 patent is attached hereto as Exhibit A.

1 **II. PARTIES**

2 2. Plaintiff Valent is a California corporation having a principal place of business at 1600
3 Riviera Avenue, Walnut Creek, CA 94596. Valent is a wholly-owned subsidiary of SCC.

4 3. Plaintiff SCC is a Japanese corporation with its principal place of business located at
5 Tokyo Sumitomo Twin Building (East), 27-1, Shinkawa 2-chome, Chuo-ku, Tokyo 104-8260, Japan.

6 4. Defendant Syngenta is a Delaware corporation having a principal place of business at
7 410 S. Swing Rd, Greensboro, North Carolina 27409. Syngenta may be served with process in this
8 state by serving its registered agent, CT Corporation System, 818 West Seventh Street, Los Angeles,
9 CA 90017. Defendant Syngenta is a wholly-owned subsidiary of Syngenta AG, a corporation formed
10 under the laws of Switzerland.

11 **III. JURISDICTION AND VENUE**

12 5. This case arises under the patent laws, 35 U.S.C. §§ 1 *et seq.*, and the Federal
13 Declaratory Judgments Act, 28 U.S.C. §§ 2201-02. An actual, substantial, and continuing justiciable
14 controversy exists between Valent and SCC on one hand and Syngenta on the other hand to which
15 Valent and SCC require a declaration of rights by this Court. The controversy relates to the invalidity
16 and non-infringement of the '469 Patent and Syngenta's right to threaten a suit for infringement of that
17 patent against Valent's and SCC's use and sale of clothianidin for controlling insects on transgenic
18 plants within this Judicial District and throughout the United States.

19 6. This Court has jurisdiction over this case under 28 U.S.C. §§ 1331, 1338, 1367, and
20 2201-02.

21 7. Upon information and belief, Syngenta is subject to personal jurisdiction in this Judicial
22 District because it resides within the State of California and is in systematic and continuous contact
23 with this Judicial District. Syngenta is registered to conduct business with the California Secretary of
24 State/Corporations Division, and has a registered agent for service of process within the State of
25 California. Syngenta has registered a number of insecticides with the California Department of
26 Pesticide Regulation, sells and markets pesticides within the State, maintains a website directed, in
27 part, to residents of this Judicial District, and employs representatives to service customers in this
28

1 Judicial District. Upon information and belief, Syngenta regularly conducts business within this
2 Judicial District.

3 8. Venue is proper in this Judicial District under 28 U.S.C. §§ 1391(b) & (c) and 1400 (b).

4 **IV. SUMMARY OF FACTUAL ALLEGATIONS**

5 9. Valent and SCC develop, manufacture, market, and supply proprietary agricultural
6 chemical products.

7 10. SCC owns all rights to U.S. Patent No. 5,034,404 ("the '404 patent), a landmark patent
8 for clothianidin, which is a valuable and successful insecticide. The '404 patent claims the compound
9 for all purposes, and the method of using it as an insecticide for plants in general.

10 11. About 15 years after the '404 patent was issued, the U.S. Patent & Trademark Office
11 ("USPTO") issued another clothianidin use patent, the '469 patent, to Syngenta. The '469 patent
12 claims patent protection for controlling pests on transgenic useful plants with clothianidin, whereas the
13 earlier '404 patent received patent protection for controlling insect pests on plants in general with
14 clothianidin.

15 12. The '469 patent is both anticipated and obvious and therefore invalid in light of the
16 prior art. This prior art includes: (1) earlier patents and publications that describe the use of
17 clothianidin for controlling insects on plants in general; (2) recognition that clothianidin kills certain
18 insects that are common pests for both transgenic and non-transgenic plants, so that it would be
19 obvious to control the same insects with clothianidin on both types of plants; (3) recognition that
20 transgenic plants need to be treated with a broad spectrum insecticide, and clothianidin is a known
21 broad spectrum insecticide; and (4) the prior art use on transgenic plants of imidacloprid, another
22 neonicotinoid that is similar to clothianidin.

23 13. Valent and SCC plan to begin selling clothianidin in the United States for controlling
24 insects on transgenic plants, including, but not limited to, by treatment of seeds thereof. Despite the
25 clear invalidity of the '469 patent, SCC attempted to negotiate an amicable business resolution with
26 Syngenta simply because SCC believed: (1) a reasonable business resolution was preferable to
27 litigation; and (2) patent litigation might unduly delay Valent's and SCC's plans for launching their
28 own clothianidin products. Syngenta refused an amicable business resolution at least in part because

1 Syngenta has already granted a license to Bayer AG or one of its affiliates ("Bayer") that requires
2 Bayer's consent for any additional licenses and Bayer has refused to consent.

3 14. During SCC's meetings with Syngenta, Syngenta threatened SCC with enforcement of
4 its rights under the '469 patent and warned SCC not to market or sell clothianidin for transgenic plants
5 in the United States. A substantial and actual controversy exists between the parties.

6 **V. NEONICOTINOID INSECTICIDES**

7 15. The great productivity of United States agriculture is achieved in part by the use of
8 pesticides to maximize crop yield and quality.

9 16. A pesticide is any substance that is intended to repel, kill, or otherwise control any
10 species designated as a "pest" – which includes insects (insecticides), rodents (rodenticides), weeds
11 (herbicides), fungi (fungicides), bacteria (bactericides), and other organisms. Pesticides control these
12 pests by physically, chemically or biologically interfering with their metabolism or behavior.

13 17. The present dispute involves neonicotinoid insecticides. Neonicotinoid insecticides are
14 increasingly deployed for insect management and have become an important class of insecticides since
15 the commercialization of imidacloprid, which first received patent protection in 1988 through U.S.
16 Patent No. 4,742,060 ("the '060 patent"). The '060 patent is now expired and thus in the public
17 domain. The '060 patent described the enhanced benefits of using imidacloprid and other
18 neonicotinoids, instead of conventional pesticides, because of their powerful insecticidal properties.

19 18. Imidacloprid had been used successfully to control insect pests in both transgenic and
20 non-transgenic plants before the '469 patent was invented.

21 **VI. THE CLOTHIANIDIN INSECTICIDE**

22 19. Clothianidin is a later generation of neonicotinoid insecticide, which first received
23 patent protection in the '404 patent in 1991.

24 20. The '404 patent makes numerous claims relating to clothianidin and to the use of
25 clothianidin as an insecticide. Notably, the '404 patent not only claims the invention of clothianidin as
26 a compound, but also specifically identifies the use of clothianidin to control insects in plants by
27 contacting the insects directly or indirectly to kill them.

21. By the early 1990's, significant prior art described the use of neonicotinoids, including clothianidin, to control a wide array of insects on plants in general.

VII. SCC ACQUIRES THE '404 PATENT AND TAKEDA'S AGRICULTURAL CHEMICAL BUSINESS

22. In 2002, SCC, through its subsidiary, acquired the agricultural chemical business of Takeda Chemical Industries, Ltd. ("Takeda"). Acquisition of the '404 patent was the "crown jewel" of the transaction because it is the landmark patent that claims the invention of the clothianidin compound.

23. Valent and SCC could not sell clothianidin for seed treatment immediately, however, because the '404 patent was subject to a five year exclusive license in favor of Bayer for seed treatment. With one exception, that period of exclusivity expires in November 2008.

24. Upon such expiration, Valent and SCC can compete in the U.S. for clothianidin sales for seed treatment. This right to compete is important for SCC to maximize the use of the '404 patent, and therefore to recover its significant investment in the acquisition of Takeda's agricultural chemical business.

VIII. SCC'S NEGOTIATIONS WITH SYNGENTA AND BAYER

25. Since the issuance of the '469 patent, SCC has traveled globally to meet with both Syngenta and Bayer several times to investigate obtaining a patent license. SCC learned from Syngenta that Syngenta had granted Bayer a co-exclusive license to the '469 patent and cannot grant a license to SCC without Bayer's consent. In these meetings, SCC expressed its view that the '469 patent is invalid, but that it nonetheless wished to pursue a reasonable licensing agreement to protect its product launch plans and to avoid costly patent litigation. SCC's efforts to negotiate with both Bayer and Syngenta have been fruitless and futile.

26. Moreover, during these negotiations Syngenta notified SCC that if SCC attempts to sell clothianidin in the U.S. for controlling insects on transgenic plants, Syngenta will enforce its patent rights against SCC through a patent infringement lawsuit.

27. The most recent meeting between SCC and Syngenta occurred on January 9, 2008 in Basel, Switzerland. The parties were unable to break their deadlock at this meeting.

1 **IX. VALENT AND SCC PLAN TO ENTER THE MARKET AND INITIATE TESTING**

2 28. Valent and SCC intend to sell clothianidin for controlling insects on transgenic plants.
3 This is an important business strategy for SCC to recover its significant investment in acquiring the
4 Takeda agricultural business. According to SCC's long-standing plan to sell clothianidin in the United
5 States for controlling insects on transgenic plants, Valent will act as the exclusive distributor for SCC's
6 clothianidin products in the United States.

7 29. In order to introduce an agricultural chemical for use in the United States, it is standard
8 industry practice to undertake testing programs over several growing seasons prior to product launch.
9 In keeping with this practice and federal regulatory requirements, Valent began formulating and testing
10 small amounts of clothianidin for controlling insects on transgenic plants in this State and in various
11 other jurisdictions throughout the United States.

12 30. Valent and its subcontractors formulated and tested clothianidin for controlling insects
13 on transgenic corn, cotton, canola and soybean plants during the 2005-07 growing seasons.

14 31. On September 4, 2007, Valent submitted a New Product Registration Application to the
15 United States Environmental Protection Agency on canola, corn and sorghum. All proposed uses are
16 the same as those of Bayer's registered seed treatment product Poncho™.

17 32. Valent intends to conduct additional testing during the 2008 growing season that will be
18 similar in nature and timing to the 2005-2007 programs. All of these testing programs were designed
19 and implemented as part of SCC's long-standing plan to sell clothianidin in the United States in 2009
20 for controlling insects on all plants, including transgenic plants.

21 33. Syngenta's assertion of the '469 patent against SCC has created adverse interests
22 between the parties and a reasonable apprehension of an imminent suit against Valent and SCC and
23 threatens Valent's and SCC's ability to test, market, use, and sell clothianidin within the United States
24 upon its approval by the EPA.

COUNT I

(DECLARATORY JUDGMENT OF INVALIDITY)

34. Valent and SCC repeat and reallege, as though fully set forth herein, the allegations contained in paragraphs 1 through 33 above.

35. Valent and SCC contend that the claims of the '469 patent are invalid for failing to comply with the conditions and requirements for patentability as set forth in the United States laws, Title 35 U.S.C. §§ 101, 102, 103, 112, and the rules, regulations, and laws pertaining thereto.

36. A substantial controversy exists between Valent/SCC and Syngenta due to Syngenta's assertion of the '469 patent, and the parties' legal interests are adverse.

37. Valent and SCC seek a judicial determination and declaration that the claims of the '469 patent are invalid for failure to comply with the conditions and requirements for patentability as set forth in the United States laws, Title 35 U.S.C. §§ 101, 102, 103, 112, and the rules, regulations, and laws pertaining thereto. Such a determination and declaration is necessary and appropriate at this time so that the parties may ascertain their respective rights and duties regarding the invalidity of the '469 patent.

COUNT II

(DECLARATORY JUDGMENT OF NON-INFRINGEMENT)

38. Valent and SCC repeat and reallege, as though fully set forth herein, the allegations contained in paragraphs 1 through 33 above.

39. Valent and SCC have not and do not presently directly or indirectly infringe any valid claim of the '469 patent, either literally or under the doctrine of equivalents.

40. A substantial controversy exists between Valent/SCC and Syngenta due to Syngenta's assertion of the '469 patent, and the parties' legal interests are adverse.

41. Valent and SCC seek a judicial determination and declaration that both parties have not infringed and are not now infringing, either directly, indirectly, literally, or equivalently, any valid claim of the '469 patent. Such a determination and declaration is necessary and appropriate at this time so that the parties may ascertain their respective rights and duties regarding the non-infringement of the '469 patent.

PRAYER FOR RELIEF

WHEREFORE Plaintiffs Valent and SCC respectfully request the following relief:


1. A judicial determination and declaration that United States Patent No. 7,105,469 B2 is invalid, in whole or part;
2. A judicial determination and declaration that Valent and SCC have not infringed and are not now infringing, either directly, indirectly, literally, or equivalently, any valid claim of United States Patent No. 7,105,469 B2;
3. A declaration that this case is "exceptional" within the meaning of 35 U.S.C. § 285, entitling Valent and SCC to an award of its reasonable attorneys' fees, expenses, and costs in this action; and
4. Such other and further relief as the Court deems just and proper.

REQUEST FOR JURY TRIAL

A trial by jury is requested for all issues triable to a jury.

Dated: January 31, 2008

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EXHIBIT A

US007105469B2

(12) **United States Patent**
Lee et al.(10) **Patent No.:** **US 7,105,469 B2**
(45) **Date of Patent:** **Sep. 12, 2006**(54) **USE OF NEONICOTINOIDS IN PEST CONTROL**(75) Inventors: **Bruce Lee**, Bad Krozingen (DE);
Marius Sutter, Binningen (CH);
Hubert Buholzer, Binningen (CH)(73) Assignee: **Syngenta Crop Protection, Inc.**,
Greensboro, NC (US)(*) Notice: Subject to any disclaimer, the term of this
patent is extended or adjusted under 35
U.S.C. 154(b) by 168 days.(21) Appl. No.: **11/019,051**(22) Filed: **Dec. 21, 2004**(65) **Prior Publication Data**

US 2005/0120411 A1 Jun. 2, 2005

Related U.S. Application Data(60) Division of application No. 10/125,136, filed on Apr.
18, 2002, now Pat. No. 6,844,339, which is a con-
tinuation of application No. 09/600,384, filed on Sep.
21, 2000, now abandoned.(30) **Foreign Application Priority Data**Jan. 16, 1998 (CH) 80/98
Mar. 25, 1998 (CH) 706/98(51) **Int. Cl.****A01N 25/26** (2006.01)
A01N 43/48 (2006.01)
A01N 43/78 (2006.01)(52) **U.S. Cl.** 504/100; 504/253; 504/266(58) **Field of Classification Search** 514/229.2;
504/266, 100, 253
See application file for complete search history.(56) **References Cited****U.S. PATENT DOCUMENTS**4,798,837 A 1/1989 Drabek et al.
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Ctr. Of green peach aphid on pot., XP002102508, Longtine, et al.

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Primary Examiner—Alton Pryor(74) *Attorney, Agent, or Firm*—Jacqueline Haley(57) **ABSTRACT**There is now described a method of controlling pests with
nitroimino- or nitroguanidino-compounds; more specifically
a method of controlling pests in and on transgenic crops of
useful plants, such as, for example, in crops of maize,
cereals, soya beans, tomatoes, cotton, potatoes, rice and
mustard, with a nitroimino- or nitroguanidino-compound,
especially with thiamethoxam, characterized in that a pes-
ticial composition comprising a nitroimino- or nitroguani-
dino-compound in free form or in agrochemically useful salt
form and at least one auxiliary is applied to the pests or their
environment, in particular to the crop plant itself.**8 Claims, No Drawings**

US 7,105,469 B2

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USE OF NEONICOTINOIDS IN PEST CONTROL

This application is a divisional application of U.S. patent application Ser. No. 10/125,136, filed Apr. 18, 2002 now U.S. Pat. No. 6,844,339, which is a continuation of U.S. patent application Ser. No. 09/600,384, filed Sep. 21, 2000 (now abandoned), the contents of which are incorporated herein by reference.

The present invention relates to a method of controlling pests with a nitroimino- or nitroguanidino-compound, especially thiamethoxam; more specifically to a novel method of controlling pests in and on transgenic crops of useful plants with a nitroimino- or nitroguanidino-compound.

Certain pest control methods are proposed in the literature. However, these methods are not fully satisfactory in the field of pest control, which is why there is a demand for providing further methods for controlling and combating pests, in particular insects and representatives of the order Acarina, or for protecting plants, especially crop plants. This object is achieved according to the invention by providing the present method.

The present invention therefore relates to a method of controlling pests in crops of transgenic useful plants, such as, for example, in crops of maize, cereals, soya beans, tomatoes, cotton, potatoes, rice and mustard, characterized in that a pesticidal composition comprising a nitroimino- or nitroguanidino-compound, especially thiamethoxam, imidacloprid, Ti-435 or thiacloprid in free form or in agrochemically useful salt form and at least one auxiliary is applied to the pests or their environment, in particular to the crop plant itself; to the use of the composition in question and to propagation material of transgenic plants which has been treated with it.

Surprisingly, it has now emerged that the use of a nitroimino- or nitroguanidino-compound for controlling pests on transgenic useful plants which contain—one or more genes expressing a pesticidally, particularly insecticidally, acaricidally, nematocidally or fungicidally active ingredient, or which are tolerant against herbicides or resistant against the attack of fungi, has a synergistic effect. It is highly surprising that the use of a nitroimino- or nitroguanidino-compound in combination with a transgenic plant exceeds the additive effect, to be expected in principle, on the pests to be controlled and thus extends the range of action of the nitroimino- or nitroguanidino-compound and of the active principle expressed by the transgenic plant in particular in two respects:

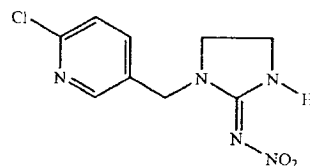
In particular, it has been found, surprisingly, that within the scope of invention the pesticidal activity of a nitroimino- or nitroguanidino-compound in combination with the effect expressed by the transgenic useful plant, is not only additive in comparison with the pesticidal activities of the nitroimino- or nitroguanidino-compound alone and of the transgenic crop plant alone, as can generally be expected, but that a synergistic effect is present. The term "synergistic", however, is in no way to be understood in this connection as being restricted to the pesticidal activity, but the term also refers to other advantageous properties of the method according to the invention compared with the nitroimino- or nitroguanidino-compound and the transgenic useful plant alone. Examples of such advantageous properties which may be mentioned are: extension of the pesticidal spectrum of action to other pests, for example to resistant strains; reduction in the application rate of the nitroimino- or nitroguanidino-compound, or sufficient control of the pests with the aid of the compositions according to the invention

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even at an application rate of the nitroimino- or nitroguanidino-compound alone and the transgenic useful plant alone are entirely ineffective; enhanced crop safety; improved quality of produce such as higher content of nutrient or oil, better fiber quality, enhanced shelf life, reduced content of toxic products such as mycotoxins, reduced content of residues or unfavorable constituents of any kind or better digestability; improved tolerance to unfavorable temperatures, draughts or salt content of water; enhanced assimilation rates such as nutrient uptake, water uptake and photosynthesis; favorable crop properties such as altered leaf area, reduced vegetative growth, increased yields, favorable seed shape/seed thickness or germination properties, altered colonisation by saprophytes or epiphytes, reduction of senescence, improved phytoalexin production, improved accelerated ripening, flower set increase, reduced boll fall and shattering, better attraction to beneficials and predators, increased pollination, reduced attraction to birds; or other advantages known to those skilled in the art.

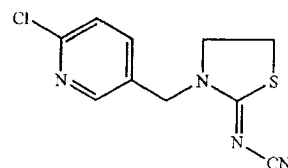
Nitroimino- and nitroguanidino-compounds, such as thiamethoxam (5-(2-Chlorthiazol-5-ylmethyl)-3-methyl-4-nitroimino-perhydro-1,3,5-oxadiazin), are known from EP-A-0'580'553. Within the scope of invention thiamethoxam is preferred.

Also preferred within the scope of invention is imidacloprid of the formula



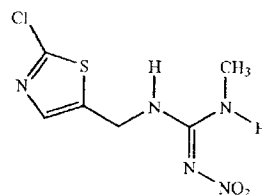
known from The Pesticide Manual, 10th Ed. (1991), The British Crop Protection Council, London, page 591;

also preferred is Thiacloprid of the formula



known from EP-A-235'725;

also preferred is the compound of the formula



known as Ti-435 (clothianidin) from EP-A-376'279

US 7,105,469 B2

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The agrochemically compatible salts of the nitroimino- or nitroguanidino-compounds are, for example, acid addition salts of inorganic and organic acids, in particular of hydrochloric acid, hydrobromic acid, sulfuric acid, nitric acid, perchloric acid, phosphoric acid, formic acid, acetic acid, trifluoroacetic acid, oxalic acid, malonic acid, toluene-sulfonic acid or benzoic acid. Preferred within the scope of the present invention is a composition known per se which comprises, as active ingredient, thiamethoxam and imidacloprid, each in the free form, especially thiamethoxam.

The transgenic plants used according to the invention are plants, or propagation material thereof, which are transformed by means of recombinant DNA technology in such a way that they are—for instance—capable of synthesizing selectively acting toxins as are known, for example, from toxin-producing in vertebrates, especially of the phylum Arthropoda, as can be obtained from *Bacillus thuringiensis* strains; or as are known from plants, such as lectins; or in the alternative capable of expressing a herbicidal or fungicidal resistance. Examples of such toxins, or transgenic plants which are capable of synthesizing such toxins, have been disclosed, for example, in EP-A-0 374 753, WO 93/07278, WO 95/34656, EP-A-0 427 529 and EP-A-451 878 and are incorporated by reference in the present application.

The methods for generating such transgenic plants are widely known to those skilled in the art and described, for example, in the publications mentioned above.

The toxins which can be expressed by such transgenic plants include, for example, toxins, such as proteins which have insecticidal properties and which are expressed by transgenic plants, for example *Bacillus cereus* proteins or

4

Bacillus popilliae proteins; or *Bacillus thuringiensis* endotoxins (B.t.), such as CryIA(a), CryIA(b), CryIA(c), CryIIA, CryIIIA, CryIIIB2 or CytA; VIP1; VIP2; VIP3; or insecticidal proteins of bacteria colonising nematodes like *Photorhabdus* spp or *Xenorhabdus* spp such as *Photorhabdus luminescens*, *Xenorhabdus nematophilus* etc.; proteinase inhibitors, such as trypsin inhibitors, serine protease inhibitors, patatin, cystatin, papain inhibitors; ribosome-inactivating proteins (RIP), such as ricin, maize RIP, abrin, luffin, saporin or bryodin; plant lectins such as pea lectins, barley lectins or snowdrop lectins; or agglutinins; toxins produced by animals, such as scorpion toxins, spider venoms, wasp venoms and other insect-specific neurotoxins; steroid metabolism enzymes, such as 3-hydroxysteroid oxidase, ecdysteroid UDP-glycosyl transferase, cholesterol oxidases, ecdysone inhibitors, HMG-CoA reductase, ion channel blockers such as sodium and calcium, juvenile hormone esterase, diuretic hormone receptors, stilbene synthase, bibenzyl synthase, chitinases and glucanases.

Examples of known transgenic plants which comprise one or more genes which encode insecticidal resistance and express one or more toxins are the following: KnockOut® (maize), YieldGard® (maize); NuCOTN 33B® (cotton), Boligard® (cotton), NewLeaf® (potatoes), NatureGard® and Protecta®.

The following tables comprise further examples of targets and principles and crop phenotypes of transgenic crops which show tolerance against pests mainly insects, mites, nematodes, virus, bacteria and diseases or are tolerant to specific herbicides or classes of herbicides.

TABLE A1

Crop: Maize	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazopyrimidines, Pyrimidylbenzoates, Phthalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkane carboxylic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxachlortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxynil
5-Enolpyruvyl-3-phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1	Xenobiotics and herbicides such as Sulfonylureas
Dimboa biosynthesis (Bx1 gene)	<i>Helminthosporium turcicum</i> , <i>Rhizoglyphus maydis</i> , <i>Diplodia maydis</i> , <i>Ostrinia nubilalis</i> , <i>lepidoptera</i> sp.
CMIII (small basic maize seed peptide)	plant pathogens eg. <i>fusarium</i> , <i>alternaria</i> , <i>sclerotinia</i>
Corn-SAFP (zeamatin)	plant pathogens eg. <i>fusarium</i> , <i>alternaria</i> , <i>sclerotinia</i> , <i>rhizoctonia</i> , <i>chaetomium</i> , <i>phycomyces</i>
Hm1 gene	Cochliobolus
Chitinases	plant pathogens
Glucanases	plant pathogens

US 7,105,469 B2

5

6

TABLE A1-continued

Crop: Maize	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
Coat proteins	viruses such as maize dwarf mosaic virus, maize chlorotic dwarf virus
<i>Bacillus thuringiensis</i> toxins, VIP 3, <i>Bacillus cereus</i> toxins, <i>Photorhabdus</i> and <i>Xenorhabdus</i> toxins	<i>lepidoptera</i> , <i>coleoptera</i> , <i>diptera</i> , nematodes, eg. <i>ostrinia nubilalis</i> , <i>heliothis zea</i> , armyworms eg. <i>spodoptera frugiperda</i> , corn rootworms, <i>sesamia</i> sp., black cutworm, asian corn borer, weevils
3-Hydroxysteroid oxidase	<i>lepidoptera</i> , <i>coleoptera</i> , <i>diptera</i> , nematodes, eg. <i>ostrinia nubilalis</i> , <i>heliothis zea</i> , armyworms eg. <i>spodoptera frugiperda</i> , corn rootworms, <i>sesamia</i> sp., black cutworm, asian corn borer, weevils
Peroxidase	<i>lepidoptera</i> , <i>coleoptera</i> , <i>diptera</i> , nematodes, eg. <i>ostrinia nubilalis</i> , <i>heliothis zea</i> , armyworms eg. <i>spodoptera frugiperda</i> , corn rootworms, <i>sesamia</i> sp., black cutworm, asian corn borer, weevils
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor (LAPI)	<i>lepidoptera</i> , <i>coleoptera</i> , <i>diptera</i> , nematodes, eg. <i>ostrinia nubilalis</i> , <i>heliothis zea</i> , armyworms eg. <i>spodoptera frugiperda</i> , corn rootworms, <i>sesamia</i> sp., black cutworm, asian corn borer, weevils
Limonene synthase	corn rootworms
Lectines	<i>lepidoptera</i> , <i>coleoptera</i> , <i>diptera</i> , nematodes, eg. <i>ostrinia nubilalis</i> , <i>heliothis zea</i> , armyworms eg. <i>spodoptera frugiperda</i> , corn rootworms, <i>sesamia</i> sp., black cutworm, asian corn borer, weevils
Protease Inhibitors eg. cystatin, patatin, virgiferin, CPTI	weevils, corn rootworm
ribosome inactivating protein	<i>lepidoptera</i> , <i>coleoptera</i> , <i>diptera</i> , nematodes, eg. <i>ostrinia nubilalis</i> , <i>heliothis zea</i> , armyworms eg. <i>spodoptera frugiperda</i> , corn rootworms, <i>sesamia</i> sp., black cutworm, asian corn borer, weevils
maize 5C9 polypeptide	<i>lepidoptera</i> , <i>coleoptera</i> , <i>diptera</i> , nematodes, eg. <i>ostrinia nubilalis</i> , <i>heliothis zea</i> , armyworms eg. <i>spodoptera frugiperda</i> , corn rootworms, <i>sesamia</i> sp., black cutworm, asian corn borer, weevils
HMG-CoA reductase	<i>lepidoptera</i> , <i>coleoptera</i> , <i>diptera</i> , nematodes, eg. <i>ostrinia nubilalis</i> , <i>heliothis zea</i> , armyworms eg. <i>spodoptera frugiperda</i> , corn rootworms, <i>sesamia</i> sp., black cutworm, asian corn borer, weevils

TABLE A2

Crop: Wheat	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazolopyrimidines, Pyrimidylxybenzoates, Phthalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkanoic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxaclortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthrnilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxynil
5-Eolpyruvyl-3phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides,

US 7,105,469 B2

7

8

TABLE A2-continued

<u>Crop Wheat</u>	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
Cytochrome P450 eg. P450 SU1	phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Antifungal polypeptide AlyAFP	Xenobiotics and herbicides such as Sulfonyleureas
glucose oxidase	plant pathogenes eg. <i>septoria</i> and <i>fusarium</i>
pyrrolnitrin synthesis genes	plant pathogenes eg. <i>fusarium</i> , <i>septoria</i>
serine/threonine kinases	plant pathogenes eg. <i>fusarium</i> , <i>septoria</i> and other diseases
Hypersensitive response eliciting polypeptide	plant pathogenes eg. <i>fusarium</i> , <i>septoria</i> and other diseases
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Chitinases	plant pathogenes
Glucanases	plant pathogenes
double stranded ribonuclease	viruses such as BYDV and MSMV
Coat proteins	viruses such as BYDV and MSMV
<i>Bacillus thuringiensis</i> toxins, VIP 3, <i>Bacillus cereus</i> toxins, <i>Photorabdus</i> and <i>Xenorhabdus</i> toxins	lepidoptera, coleoptera, diptera, nematodes,
3-Hydroxysteroid oxidase	lepidoptera, coleoptera, diptera, nematodes,
Peroxidase	lepidoptera, coleoptera, diptera, nematodes,
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera, coleoptera, diptera, nematodes,
Lectines	lepidoptera, coleoptera, diptera, nematodes, aphids
Protease Inhibitors eg. cystatin, patatin, virgiferin, CPTI	lepidoptera, coleoptera, diptera, nematodes, aphids
ribosome inactivating protein	lepidoptera, coleoptera, diptera, nematodes, aphids
HMG-CoA reductase	lepidoptera, coleoptera, diptera, nematodes, eg. <i>ostrinia nubilalis</i> , <i>heliothis zea</i> , armyworms eg. <i>spodoptera frugiperda</i> , corn rootworms, <i>sesamia</i> sp., black cutworm, asian corn borer, weevils

TABLE A3

<u>Crop Barley</u>	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
Acetolactate synthase (ALS)	Sulfonyleureas, Imidazolinones, Triazolopyrimidines, Pyrimidylxybenzoates, Phthalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkanoic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxaclortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxynil
5-Enolpyruvyl-3-phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1	Xenobiotics and herbicides such as Sulfonyleureas
Antifungal polypeptide AlyAFP	plant pathogenes eg. <i>septoria</i> and <i>fusarium</i>
glucose oxidase	plant pathogenes eg. <i>fusarium</i> , <i>septoria</i>

US 7,105,469 B2

9

10

TABLE A3-continued

Crop Barley	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
pyrrolnitrin synthesis genes	plant pathogens eg. <i>fusarium</i> , <i>septoria</i>
serine/threonine kinases	plant pathogens eg. <i>fusarium</i> , <i>septoria</i> and other diseases
Hypersensitive response eliciting polypeptide	plant pathogens eg. <i>fusarium</i> , <i>septoria</i> and other diseases
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Chitinases	plant pathogens
Glucanases	plant pathogens
double stranded ribonuclease	viruses such as BYDV and MSMV
Coat proteins	viruses such as BYDV and MSMV
<i>Bacillus thuringiensis</i> toxins, VIP 3, <i>Bacillus cereus</i> toxins, <i>Photorabdus</i> and <i>Xenorhabdus</i> toxins	lepidoptera, coleoptera, diptera, nematodes,
3-Hydroxysteroid oxidase	lepidoptera, coleoptera, diptera, nematodes,
Peroxidase	lepidoptera, coleoptera, diptera, nematodes,
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera, coleoptera, diptera, nematodes,
Lectines	lepidoptera, coleoptera, diptera, nematodes, aphids
Protease Inhibitors eg. cystatin, patatin, virgiferin, CPTI	lepidoptera, coleoptera, diptera, nematodes, aphids
ribosome inactivating protein	lepidoptera, coleoptera, diptera, nematodes, aphids
HMG-CoA reductase	lepidoptera, coleoptera, diptera, nematodes, aphids

TABLE A4

Crop Rice	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazolopyrimidines, Pyrimidylxybenzoates, Phtalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkane carboxylic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxachlortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthraniolate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxynil
5-Enolpyruvyl-3-phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1	Xenobiotics and herbicides such as Sulfonylureas
Antifungal polypeptide AlyAFP	plant pathogens
glucose oxidase	plant pathogens
pyrrolnitrin synthesis genes	plant pathogens
serine/threonine kinases	plant pathogens
Phenylalanine ammonia lyase (PAL)	plant pathogens eg bacterial leaf blight and rice blast, inducible
phytoalexins	plant pathogens eg bacterial leaf blight and rice blast
B-1,3-glucanase antisense	plant pathogens eg bacterial leaf blight and rice blast
receptor kinase	plant pathogens eg bacterial leaf blight and rice blast
Hypersensitive response eliciting	plant pathogens

US 7,105,469 B2

11

12

TABLE A4-continued

Crop Rice	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
polypeptide	
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Chitinases	plant pathogens eg bacterial leaf blight and rice blast
Glucanases	plant pathogens
double stranded ribonuclease	viruses such as BYDV and MSMV
Coat proteins	viruses such as BYDV and MSMV
<i>Bacillus thuringiensis</i> toxins, VIP 3,	lepidoptera eg. stemborer, coleoptera eg
<i>Bacillus cereus</i> toxins, <i>Photobacterium</i> and	rice water weevil, diptera, rice hoppers
<i>Xenorhabdus</i> toxins	eg brown rice hopper
3-Hydroxysteroid oxidase	lepidoptera eg. stemborer, coleoptera eg
	rice water weevil, diptera, rice hoppers
	eg brown rice hopper
Peroxidase	lepidoptera eg. stemborer, coleoptera eg
	rice water weevil, diptera, rice hoppers
	eg brown rice hopper
Amino peptidase inhibitors eg. Leucine	lepidoptera eg. stemborer, coleoptera eg
amino peptidase inhibitor	rice water weevil, diptera, rice hoppers
	eg brown rice hopper
Lectines	lepidoptera eg. stemborer, coleoptera eg
	rice water weevil, diptera, rice hoppers
	eg brown rice hopper
Protease Inhibitors,	lepidoptera eg. stemborer, coleoptera eg
	rice water weevil, diptera, rice hoppers
	eg brown rice hopper
ribosome inactivating protein	lepidoptera eg. stemborer, coleoptera eg
	rice water weevil, diptera, rice hoppers
	eg brown rice hopper
HMG-CoA reductase	lepidoptera eg. stemborer, coleoptera eg
	rice water weevil, diptera, rice hoppers
	eg brown rice hopper

TABLE A5

Crop Soya	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazolopyrimidines, Pyrimidylxybenzoates, Phthalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkane carboxylic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxaflortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxynil
5-Enolpyruvyl-3-phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1 or selection	Xenobiotics and herbicides such as Sulfonylureas
Antifungal polypeptide AlyAFP	bacterial and fungal pathogens such as <i>fusarium</i> , <i>sclerotinia</i> , stemrot
oxalate oxidase	bacterial and fungal pathogens such as <i>fusarium</i> , <i>sclerotinia</i> , stemrot
glucose oxidase	bacterial and fungal pathogens such as <i>fusarium</i> , <i>sclerotinia</i> , stemrot
pyrrolinitrin synthesis genes	bacterial and fungal pathogens such as <i>fusarium</i> , <i>sclerotinia</i> , stemrot

US 7,105,469 B2

13

14

TABLE A5-continued

Effectuated target or expressed principle(s)	Crop phenotype/Tolerance to
serine/threonine kinases	bacterial and fungal pathogens such as <i>fusarium</i> , <i>sclerotinia</i> , stemrot
Phenylalanine ammonia lyase (PAL)	bacterial and fungal pathogens such as <i>fusarium</i> , <i>sclerotinia</i> , stemrot
phytoalexins	plant pathogens eg bacterial leaf blight and rice blast
B-1,3-glucanase antisense	plant pathogens eg bacterial leaf blight and rice blast
receptor kinase	bacterial and fungal pathogens such as <i>fusarium</i> , <i>sclerotinia</i> , stemrot
Hypersensitive response eliciting polypeptide	plant pathogens
Systemic acquired resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Chitinases	bacterial and fungal pathogens such as <i>fusarium</i> , <i>sclerotinia</i> , stemrot
Glucanases	bacterial and fungal pathogens such as <i>fusarium</i> , <i>sclerotinia</i> , stemrot
double stranded ribonuclease	viruses such as BPMV and SbMV
Coat proteins	viruses such as BYDV and MSMV
<i>Bacillus thuringiensis</i> toxins, VIP 3, <i>Bacillus cereus</i> toxins, <i>Photorhabdus</i> and <i>Xenorhabdus</i> toxins	lepidoptera, coleoptera, aphids
3-Hydroxysteroid oxidase	lepidoptera, coleoptera, aphids
Peroxidase	lepidoptera, coleoptera, aphids
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera, coleoptera, aphids
Lectines	lepidoptera, coleoptera, aphids
Protease inhibitors eg virgiferin	lepidoptera, coleoptera, aphids
ribosome inactivating protein	lepidoptera, coleoptera, aphids
HMG-CoA reductase	lepidoptera, coleoptera, aphids
Barnase	nematodes eg root knot nematodes and cyst nematodes
Cyst nematode hatching stimulus	cyst nematodes
Antifeeding principles	nematodes eg root knot nematodes and cyst nematodes

TABLE A6

Effectuated target or expressed principle(s)	Crop phenotype/Tolerance to
Acetolactate synthase (ALS)	Sulfonyleureas, Imidazolinones, Triazolopyrimidines, Pyrimidylloxybenzoates, Phthalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkanoic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxaflortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxynil
5-Enolpyruvyl-3-phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1 or selection	Xenobiotics and herbicides such as Sulfonyleureas
Polyphenol oxidase or Polyphenol oxidase antisense	blackspot bruise
Metallothionein	bacterial and fungal pathogens such as phytophthora
Ribonuclease	<i>Phytophthora</i> , <i>Verticillium</i> , <i>Rhizoctonia</i>

US 7,105,469 B2

15

16

TABLE A6-continued

Effectuated target or expressed principle(s)	Crop phenotype/Tolerance to
Crop Potatoes	
Antifungal polypeptide AlyAFP	bacterial and fungal pathogens such as phytophthora
oxalate oxidase	bacterial and fungal pathogens such as <i>Phytophthora</i> , <i>Verticillium</i> , <i>Rhizoctonia</i>
glucose oxidase	bacterial and fungal pathogens such as <i>Phytophthora</i> , <i>Verticillium</i> , <i>Rhizoctonia</i>
pyrrolnitrin synthesis genes	bacterial and fungal pathogens such as <i>Phytophthora</i> , <i>Verticillium</i> , <i>Rhizoctonia</i>
serine/threonine kinases	bacterial and fungal pathogens such as <i>Phytophthora</i> , <i>Verticillium</i> , <i>Rhizoctonia</i>
Cecropin B	bacteria such as <i>corynebacterium sepedonicum</i> , <i>Erwinia carotovora</i>
Phenylalanine ammonia lyase (PAL)	bacterial and fungal pathogens such as <i>Phytophthora</i> , <i>Verticillium</i> , <i>Rhizoctonia</i>
phytoalexins	bacterial and fungal pathogens such as <i>Phytophthora</i> , <i>Verticillium</i> , <i>Rhizoctonia</i>
B-1,3-glucanase antisense	bacterial and fungal pathogens such as <i>Phytophthora</i> , <i>Verticillium</i> , <i>Rhizoctonia</i>
receptor kinase	bacterial and fungal pathogens such as <i>Phytophthora</i> , <i>Verticillium</i> , <i>Rhizoctonia</i>
Hypersensitive response eliciting polypeptide	bacterial and fungal pathogens such as <i>Phytophthora</i> , <i>Verticillium</i> , <i>Rhizoctonia</i>
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Chitinases	bacterial and fungal pathogens such as <i>Phytophthora</i> , <i>Verticillium</i> , <i>Rhizoctonia</i>
Barnase	bacterial and fungal pathogens such as <i>Phytophthora</i> , <i>Verticillium</i> , <i>Rhizoctonia</i>
Disease resistance response gene 49	bacterial and fungal pathogens such as <i>Phytophthora</i> , <i>Verticillium</i> , <i>Rhizoctonia</i>
trans aldolase antisense	blackspots
Glucanases	bacterial and fungal pathogens such as <i>Phytophthora</i> , <i>Verticillium</i> , <i>Rhizoctonia</i>
double stranded ribonuclease	viruses such as PLRV, PVY and TRV
Coat proteins	viruses such as PLRV, PVY and TRV
17 kDa or 60 kDa protein	viruses such as PLRV, PVY and TRV
Nuclear inclusion proteins eg. a or b	viruses such as PLRV, PVY and TRV
Pseudoubiquitin	viruses such as PLRV, PVY and TRV
Replicase	viruses such as PLRV, PVY and TRV
<i>Bacillus thuringiensis</i> toxins, VIP 3, <i>Bacillus cereus</i> toxins, <i>Photorabdus</i> and <i>Xenorhabdus</i> toxins	<i>coleoptera</i> eg Colorado potato beetle, aphids
3-Hydroxysteroid oxidase	<i>coleoptera</i> eg Colorado potato beetle, aphids
Peroxidase	<i>coleoptera</i> eg Colorado potato beetle, aphids
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	<i>coleoptera</i> eg Colorado potato beetle, aphids
stilbene synthase	<i>coleoptera</i> eg Colorado potato beetle, aphids
Lectines	<i>coleoptera</i> eg Colorado potato beetle, aphids
Protease Inhibitors eg cystatin, psatin	<i>coleoptera</i> eg Colorado potato beetle, aphids
ribosome inactivating protein	<i>coleoptera</i> eg Colorado potato beetle, aphids
HMG-CoA reductase	<i>coleoptera</i> eg Colorado potato beetle, aphids
Cyst nematode hatching stimulus	cyst nematodes
Barnase	nematodes eg root knot nematodes and cyst nematodes
Antifeeding principles	nematodes eg root knot nematodes and cyst nematodes

US 7,105,469 B2

17

18

TABLE A7

Crop Tomatoes	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazolepyrimidines, Pyrimidylxybenzoates, Phthalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkanoic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxachlortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxynil
5-Enolpyruvyl-3-phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1 or selection	Xenobiotics and herbicides such as Sulfonylureas
Polyphenol oxidase or Polyphenol oxidase antisense	blackspot bruise
Metallothionein	bacterial and fungal pathogens such as phytophthora
Ribonuclease	<i>Phytophthora</i> , <i>Verticillium</i> , <i>Rhizoctonia</i>
Antifungal polypeptide AlyAFP	bacterial and fungal pathogens such as bacterial speck, <i>fusarium</i> , soft rot, powdery mildew, crown rot, leaf mould etc.
oxalate oxidase	bacterial and fungal pathogens such as bacterial speck, <i>fusarium</i> , soft rot, powdery mildew, crown rot, leaf mould etc.
glucose oxidase	bacterial and fungal pathogens such as bacterial speck, <i>fusarium</i> , soft rot, powdery mildew, crown rot, leaf mould etc.
pyrrolnitrin synthesis genes	bacterial and fungal pathogens such as bacterial speck, <i>fusarium</i> , soft rot, powdery mildew, crown rot, leaf mould etc.
serine/threonine kinases	bacterial and fungal pathogens such as bacterial speck, <i>fusarium</i> , soft rot, powdery mildew, crown rot, leaf mould etc.
Cecropin B	bacterial and fungal pathogens such as bacterial speck, <i>fusarium</i> , soft rot, powdery mildew, crown rot, leaf mould etc.
Phenylalanine ammonia lyase (PAL)	bacterial and fungal pathogens such as bacterial speck, <i>fusarium</i> , soft rot, powdery mildew, crown rot, leaf mould etc.
Cf genes eg. Cf 9 Cf5 Cf4 Cf2	leaf mould
Osmotin	<i>alternaria solani</i>
Alpha Hordothionin	bacteria
Systemin	bacterial and fungal pathogens such as bacterial speck, <i>fusarium</i> , soft rot, powdery mildew, crown rot, leaf mould etc.
Polygalacturonase inhibitors	bacterial and fungal pathogens such as bacterial speck, <i>fusarium</i> , soft rot, powdery mildew, crown rot, leaf mould etc.
Prf regulatory gene	bacterial and fungal pathogens such as bacterial speck, <i>fusarium</i> , soft rot, powdery mildew, crown rot, leaf mould etc.
12 <i>Fusarium</i> resistance locus	<i>fusarium</i>
phytoalexins	bacterial and fungal pathogens such as bacterial speck, <i>fusarium</i> , soft rot,

US 7,105,469 B2

19

20

TABLE A7-continued

<u>Crop Tomatoes</u>	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
	powdery mildew, crown rot, leaf mould etc.
B-1,3-glucanase antisense	bacterial and fungal pathogens such as bacterial speck, <i>fusarium</i> , soft rot, powdery mildew, crown rot, leaf mould etc.
receptor kinase	bacterial and fungal pathogens such as bacterial speck, <i>fusarium</i> , soft rot, powdery mildew, crown rot, leaf mould etc.
Hypersensitive response eliciting polypeptide	bacterial and fungal pathogens such as bacterial speck, <i>fusarium</i> , soft rot, powdery mildew, crown rot, leaf mould etc.
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Chitinases	bacterial and fungal pathogens such as bacterial speck, <i>fusarium</i> , soft rot, powdery mildew, crown rot, leaf mould etc.
Barnase	bacterial and fungal pathogens such as bacterial speck, <i>fusarium</i> , soft rot, powdery mildew, crown rot, leaf mould etc.
Glucanases	bacterial and fungal pathogens such as bacterial speck, <i>fusarium</i> , soft rot, powdery mildew, crown rot, leaf mould etc.
double stranded ribonuclease	viruses such as PLRV, PVY and ToMoV
Coat proteins	viruses such as PLRV, PVY and ToMoV
17 kDa or 60 kDa protein	viruses such as PLRV, PVY and ToMoV
Nuclear inclusion proteins eg. a or b or Nucleoprotein	viruses such as PLRV, PVY and ToMoV
Pseudoubiquitin	TRV
Replicase	viruses such as PLRV, PVY and ToMoV
<i>Bacillus thuringiensis</i> toxins, VIP 3, <i>Bacillus cereus</i> toxins, <i>Photorhabdus</i> and <i>Xenorhabdus</i> toxins	viruses such as PLRV, PVY and ToMoV
3-Hydroxysteroid oxidase	lepidoptera eg <i>heliiothis</i> , whiteflies aphids
Peroxidase	lepidoptera eg <i>heliiothis</i> , whiteflies aphids
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera eg <i>heliiothis</i> , whiteflies aphids
Lectines	lepidoptera eg <i>heliiothis</i> , whiteflies aphids
Protease Inhibitors eg cystatin, patatin	lepidoptera eg <i>heliiothis</i> , whiteflies aphids
ribosome inactivating protein	lepidoptera eg <i>heliiothis</i> , whiteflies aphids
stilbene synthase	lepidoptera eg <i>heliiothis</i> , whiteflies aphids
HMG-CoA reductase	lepidoptera eg <i>heliiothis</i> , whiteflies aphids
Cyst nematode hatching stimulus	cyst nematodes
Barnase	nematodes eg root knot nematodes and cyst nematodes
Antifeeding principles	nematodes eg root knot nematodes and cyst nematodes

TABLE A8

<u>Crop Peppers</u>	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
Acetolactate synthase (ALS)	Sulfonyleureas, Imidazolinones, Triazolopyrimidines, Pyrimidylxybenzoates, Phthalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkanoic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxaclortol, Trione such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and

US 7,105,469 B2

21

22

TABLE A8-continued

<u>Crop Peppers</u>	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
Nitrilase	catabolism
5-Enolpyruvyl-3-phosphoshikimate Synthase (EPSPS)	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxynil
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Glyphosate or sulfosate
Cytochrome P450 eg. P450 SU1 or selection	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenoxylate, oxadiazoles etc.
Polyphenol oxidase or Polyphenol oxidase antisense	Xenobiotics and herbicides such as Sulfonylureas
Metallothionein	bacterial and fungal pathogens
Ribonuclease	bacterial and fungal pathogens
Antifungal polypeptide AlyAFP	bacterial and fungal pathogens
oxalate oxidase	bacterial and fungal pathogens
glucose oxidase	bacterial and fungal pathogens
pyrrolitrin synthesis genes	bacterial and fungal pathogens
serine/threonine kinases	bacterial and fungal pathogens
Cecropin B	bacterial and fungal pathogens rot, leaf mould etc.
Phenylalanine ammonia lyase (PAL)	bacterial and fungal pathogens
Cf genes eg. Cf 9 Cf5 Cf4 Cf2	bacterial and fungal pathogens
Osmotin	bacterial and fungal pathogens
Alpha Hordothionin	bacterial and fungal pathogens
Systemin	bacterial and fungal pathogens
Polygalacturonase inhibitors	bacterial and fungal pathogens
Prf regulatory gene	bacterial and fungal pathogens
I2 <i>Fusarium</i> resistance locus	<i>fusarium</i>
phytoalexins	bacterial and fungal pathogens
B-1,3-glucanase antisense	bacterial and fungal pathogens
receptor kinase	bacterial and fungal pathogens
Hypersensitive response eliciting polypeptide	bacterial and fungal pathogens
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Chitinases	bacterial and fungal pathogens
Barnase	bacterial and fungal pathogens
Glucanases	bacterial and fungal pathogens
double stranded ribonuclease	viruses such as CMV, TEV
Coat proteins	viruses such as CMV, TEV
17 kDa or 60 kDa protein	viruses such as CMV, TEV
Nuclear inclusion proteins eg. a or b or Nucleoprotein	viruses such as CMV, TEV
Pseudoubiquitin	viruses such as CMV, TEV
Replicase	viruses such as CMV, TEV
<i>Bacillus thuringiensis</i> toxins, VIP 3, <i>Bacillus cereus</i> toxins, <i>Photorhabdus</i> and <i>Xenorhabdus</i> toxins	<i>lepidoptera</i> , whiteflies aphids
3-Hydroxysteroid oxidase	<i>lepidoptera</i> , whiteflies aphids
Peroxidase	<i>lepidoptera</i> , whiteflies aphids
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	<i>lepidoptera</i> , whiteflies aphids
Lectines	<i>lepidoptera</i> , whiteflies aphids
Protease Inhibitors eg cystatin, patatin	<i>lepidoptera</i> , whiteflies aphids
ribosome inactivating protein	<i>lepidoptera</i> , whiteflies aphids
stilbene synthase	<i>lepidoptera</i> , whiteflies aphids
HMG-CoA reductase	<i>lepidoptera</i> , whiteflies aphids
Cyst nematode hatching stimulus	cyst nematodes
Barnase	nematodes eg root knot nematodes and cyst nematodes
Antifeeding principles	nematodes eg root knot nematodes and cyst nematodes

TABLE A9

<u>Crop Grapes</u>	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazopyrimidines, Pyrimidylbenzoates, Phthalides

US 7,105,469 B2

23

24

TABLE A9-continued

Crop Grapes	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkanoic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxachlortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxynil
5-Enolpyruvyl-3-phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1 or selection	Xenobiotics and herbicides such as Sulfonylureas
Polyphenol oxidase or Polyphenol oxidase antisense	bacterial and fungal pathogens like <i>Botrytis</i> and powdery mildew
Metallothionein	bacterial and fungal pathogens like <i>Botrytis</i> and powdery mildew
Ribonuclease	bacterial and fungal pathogens like <i>Botrytis</i> and powdery mildew
Antifungal polypeptide AlyAFP	bacterial and fungal pathogens like <i>Botrytis</i> and powdery mildew
oxalate oxidase	bacterial and fungal pathogens like <i>Botrytis</i> and powdery mildew
glucose oxidase	bacterial and fungal pathogens like <i>Botrytis</i> and powdery mildew
pyrrolinitrin synthesis genes	bacterial and fungal pathogens like <i>Botrytis</i> and powdery mildew
serine/threonine kinases	bacterial and fungal pathogens like <i>Botrytis</i> and powdery mildew
Cecropin B	bacterial and fungal pathogens like <i>Botrytis</i> and powdery mildew
Phenylalanine ammonia lyase (PAL)	bacterial and fungal pathogens like <i>Botrytis</i> and powdery mildew
Cf genes eg. Cf9 Cf5 Cf4 Cf2	bacterial and fungal pathogens like <i>Botrytis</i> and powdery mildew
Osmotin	bacterial and fungal pathogens like <i>Botrytis</i> and powdery mildew
Alpha Hordothionin	bacterial and fungal pathogens like <i>Botrytis</i> and powdery mildew
Systemin	bacterial and fungal pathogens like <i>Botrytis</i> and powdery mildew
Polygalacturonase inhibitors	bacterial and fungal pathogens like <i>Botrytis</i> and powdery mildew
Prf regulatory gene	bacterial and fungal pathogens like <i>Botrytis</i> and powdery mildew
phytoalexins	bacterial and fungal pathogens like <i>Botrytis</i> and powdery mildew
B-1,3-glucanase antisense	bacterial and fungal pathogens like <i>Botrytis</i> and powdery mildew
receptor kinase	bacterial and fungal pathogens like <i>Botrytis</i> and powdery mildew
Hypersensitive response eliciting polypeptide	bacterial and fungal pathogens like <i>Botrytis</i> and powdery mildew
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Chitinases	bacterial and fungal pathogens like <i>Botrytis</i> and powdery mildew
Barnase	bacterial and fungal pathogens like <i>Botrytis</i> and powdery mildew
Glucanases	bacterial and fungal pathogens like <i>Botrytis</i> and powdery mildew
double stranded ribonuclease	viruses
Coat proteins	viruses
17 kDa or 60 kDa protein	viruses
Nuclear inclusion proteins eg. a or b or Nucleoprotein	viruses

US 7,105,469 B2

25

26

TABLE A9-continued

<u>Crop Grapes</u>	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
Pseudoubiquitin	viruses
Replicase	viruses
<i>Bacillus thuringiensis</i> toxins, VIP 3, <i>Bacillus cereus</i> toxins, <i>Photobacterium</i> and <i>Xenorhabdus</i> toxins	lepidoptera, aphids
3-Hydroxysteroid oxidase	lepidoptera, aphids
Peroxidase	lepidoptera, aphids
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera, aphids
Lectines	lepidoptera, aphids
Protease Inhibitors eg cystatin, patatin	lepidoptera, aphids
ribosome inactivating protein	lepidoptera, aphids
stilbene synthase	lepidoptera, aphids, diseases
HMG-CoA reductase	lepidoptera, aphids
Cyst nematode hatching stimulus	cyst nematodes
Barnase	nematodes eg root knot nematodes and cyst nematodes or general diseases
CBI	root knot nematodes
Antifeeding principles	nematodes eg root knot nematodes or root cyst nematodes

TABLE A10

<u>crop Oil Seed rape</u>	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazolopyrimidines, Pyrimidylxybenzoates, Phtalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkancarboxylic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxachlortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxynil
5-Enolpyruvyl-3-phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1 or selection	Xenobiotics and herbicides such as Sulfonylureas
Polyphenol oxidase or Polyphenol oxidase antisense	bacterial and fungal pathogens like <i>Cylindrosporium</i> , <i>Phoma</i> , <i>Sclerotinia</i>
Metallothionein	bacterial and fungal pathogens like <i>Cylindrosporium</i> , <i>Phoma</i> , <i>Sclerotinia</i>
Ribonuclease	bacterial and fungal pathogens like <i>Cylindrosporium</i> , <i>Phoma</i> , <i>Sclerotinia</i>
Antifungal polypeptide AlyAFP	bacterial and fungal pathogens like <i>Cylindrosporium</i> , <i>Phoma</i> , <i>Sclerotinia</i>
oxalate oxidase	bacterial and fungal pathogens like <i>Cylindrosporium</i> , <i>Phoma</i> , <i>Sclerotinia</i>
glucose oxidase	bacterial and fungal pathogens like <i>Cylindrosporium</i> , <i>Phoma</i> , <i>Sclerotinia</i>
pyrrolinitrin synthesis genes	bacterial and fungal pathogens like <i>Cylindrosporium</i> , <i>Phoma</i> , <i>Sclerotinia</i>
serine/threonine kinases	bacterial and fungal pathogens like <i>Cylindrosporium</i> , <i>Phoma</i> , <i>Sclerotinia</i>
Cecropin B	bacterial and fungal pathogens like <i>Cylindrosporium</i> , <i>Phoma</i> , <i>Sclerotinia</i>
Phenylalanine ammonia lyase (PAL)	bacterial and fungal pathogens like <i>Cylindrosporium</i> , <i>Phoma</i> , <i>Sclerotinia</i>

US 7,105,469 B2

27

28

TABLE A10-continued

<u>crop Oil Seed rape</u>	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
Cf genes eg. Cf 9 Cf5 Cf4 Cf2	bacterial and fungal pathogens like <i>Cylindrosporium</i> , <i>Phoma</i> , <i>Sclerotinia</i>
Osmotin	bacterial and fungal pathogens like <i>Cylindrosporium</i> , <i>Phoma</i> , <i>Sclerotinia</i>
Alpha Hordothionin	bacterial and fungal pathogens like <i>Cylindrosporium</i> , <i>Phoma</i> , <i>Sclerotinia</i>
Systemin	bacterial and fungal pathogens like <i>Cylindrosporium</i> , <i>Phoma</i> , <i>Sclerotinia</i>
Polygalacturonase inhibitors	bacterial and fungal pathogens like <i>Cylindrosporium</i> , <i>Phoma</i> , <i>Sclerotinia</i>
Prf regulatory gene	bacterial and fungal pathogens like <i>Cylindrosporium</i> , <i>Phoma</i> , <i>Sclerotinia</i>
phytoalexins	bacterial and fungal pathogens like <i>Cylindrosporium</i> , <i>Phoma</i> , <i>Sclerotinia</i>
B-1,3-glucanase antisense	bacterial and fungal pathogens like <i>Cylindrosporium</i> , <i>Phoma</i> , <i>Sclerotinia</i>
receptor kinase	bacterial and fungal pathogens like <i>Cylindrosporium</i> , <i>Phoma</i> , <i>Sclerotinia</i>
Hypersensitive response eliciting polypeptide	bacterial and fungal pathogens like <i>Cylindrosporium</i> , <i>Phoma</i> , <i>Sclerotinia</i>
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Chitinases	bacterial and fungal pathogens like <i>Cylindrosporium</i> , <i>Phoma</i> , <i>Sclerotinia</i>
Barnase	bacterial and fungal pathogens like <i>Cylindrosporium</i> , <i>Phoma</i> , <i>Sclerotinia</i> , nematodes
Glucanases	bacterial and fungal pathogens like <i>Cylindrosporium</i> , <i>Phoma</i> , <i>Sclerotinia</i>
double stranded ribonuclease	viruses
Coat proteins	viruses
17 kDa or 60 kDa protein	viruses
Nuclear inclusion proteins eg. a or b or Nucleoprotein	viruses
Pseudoubiquitin	viruses
Replicase	viruses
<i>Bacillus thuringiensis</i> toxins, VIP 3, <i>Bacillus cereus</i> toxins, Photorabdus and <i>Xenorhabdus</i> toxins	lepidoptera, aphids
3-Hydroxysteroid oxidase	lepidoptera, aphids
Peroxidase	lepidoptera, aphids
Amino peptidase inhibitors eg. Leucine amino peptidase inhibitor	lepidoptera, aphids
Lectines	lepidoptera, aphids
Protease Inhibitors eg cystatin, patatin, CPTI	lepidoptera, aphids
ribosome inactivating protein	lepidoptera, aphids
stilbene synthase	lepidoptera, aphids, diseases
HMG-CoA reductase	lepidoptera, aphids
Cyst nematode hatching stimulus	cyst nematodes
Barnase	nematodes eg root knot nematodes and cyst nematodes
CBI	root knot nematodes
Antifeeding principles induced at a nematode feeding site	nematodes eg root knot nematodes, root cyst nematodes

TABLE A11

Crop Brassica vegetable (cabbage, brussel sprouts, broccoli etc.)

Effected target or expressed principle(s)	Crop phenotype/Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazolepyrimidines, Pyrimidylxybenzoxates, Phtalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkanecarboxylic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxaclortol, Triones such as mesettrione or sulcotrione

55

TABLE A11-continued

Crop Brassica vegetable (cabbage, brussel sprouts, broccoli etc.)

Effected target or expressed principle(s)	Crop phenotype/Tolerance to
60 Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate lyase (ADSL)	Inhibitors of IMP and AMP synthesis
65 Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis

US 7,105,469 B2

29

TABLE A11-continued

<u>Crop Brassica vegetable (cabbage, brussel sprouts, broccoli etc.)</u>	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxynil
5-Enolpyruvyl-3-phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenoplylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1 or selection	Xenobiotics and herbicides such as Sulfonylureas
Polyphenol oxidase or Polyphenol oxidase antisense	bacterial and fungal pathogens
Metallothionein	bacterial and fungal pathogens
Ribonuclease	bacterial and fungal pathogens
Antifungal polypeptide AlyAFP	bacterial and fungal pathogens
oxalate oxidase	bacterial and fungal pathogens
glucose oxidase	bacterial and fungal pathogens
pyrrolnitrin synthesis genes	bacterial and fungal pathogens
serine/threonine kinases	bacterial and fungal pathogens
Cecropin B	bacterial and fungal pathogens
Phenylalanine ammonia lyase (PAL)	bacterial and fungal pathogens
Cf genes eg. Cf 9 Cf5 Cf4 Cf2	bacterial and fungal pathogens
Osmotin	bacterial and fungal pathogens
Alpha Hordothionin	bacterial and fungal pathogens
Systemin	bacterial and fungal pathogens
Polygalacturonase inhibitors	bacterial and fungal pathogens
Prf regulatory gene	bacterial and fungal pathogens
phytoalexins	bacterial and fungal pathogens
B-1,3-glucanase antisense	bacterial and fungal pathogens
receptor kinase	bacterial and fungal pathogens
Hypersensitive response eliciting polypeptide	bacterial and fungal pathogens
Systemic acquired resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Chitinases	bacterial and fungal pathogens
Barnase	bacterial and fungal pathogens
Glucanases	bacterial and fungal pathogens
double stranded ribonuclease	viruses
Coat proteins	viruses
17 kDa or 60 kDa protein	viruses
Nuclear inclusion proteins eg. a or b or Nucleoprotein	viruses
Pseudobiquitin	viruses
Replicase	viruses
<i>Bacillus thuringiensis</i> toxins, VIP 3, <i>Bacillus cereus</i> toxins, Photorabidus and <i>Xenorhabdus</i> toxins	lepidoptera, aphids
3-Hydroxysteroid oxidase	lepidoptera, aphids
Peroxidase	lepidoptera, aphids
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera, aphids
Lectines	lepidoptera, aphids
Protease Inhibitors eg. cystatin, patatin, CPTI	lepidoptera, aphids
ribosome inactivating protein	lepidoptera, aphids
stilbene synthase	lepidoptera, aphids, diseases
HMG-CoA reductase	lepidoptera, aphids
Cyst nematode hatching stimulus	cyst nematodes
Barnase	nematodes eg. root knot nematodes and cyst nematodes
CBI	root knot nematodes
Antifeeding principles	nematodes eg. root knot nematodes,

30

TABLE A11-continued

<u>Crop Brassica vegetable (cabbage, brussel sprouts, broccoli etc.)</u>	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
induced at a nematode feeding site	root cyst nematodes

TABLE A12

<u>Crop Pome fruits eg. apples, pears</u>	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazopyrimidines, Pyrimidylxybenzoates, Phthalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxycarboxylic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxaclortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxynil
5-Enolpyruvyl-3-phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenoplylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1 or selection	Xenobiotics and herbicides such as Sulfonylureas
Polyphenol oxidase or Polyphenol oxidase antisense	bacterial and fungal pathogens like apple scab or fireblight
Metallothionein	bacterial and fungal pathogens like apple scab or fireblight
Ribonuclease	bacterial and fungal pathogens like apple scab or fireblight
Antifungal polypeptide AlyAFP	bacterial and fungal pathogens like apple scab or fireblight
oxalate oxidase	bacterial and fungal pathogens like apple scab or fireblight
glucose oxidase	bacterial and fungal pathogens like apple scab or fireblight
pyrrolnitrin synthesis genes	bacterial and fungal pathogens like apple scab or fireblight
serine/threonine kinases	bacterial and fungal pathogens like apple scab or fireblight
Cecropin B	bacterial and fungal pathogens like apple scab or fireblight
Phenylalanine ammonia lyase (PAL)	bacterial and fungal pathogens like apple scab or fireblight
Cf genes eg. Cf 9 Cf5 Cf4 Cf2	bacterial and fungal pathogens like apple scab or fireblight
Osmotin	bacterial and fungal pathogens like apple scab or fireblight
Alpha Hordethionin	bacterial and fungal pathogens like apple scab or fireblight
Systemin	bacterial and fungal pathogens like apple scab or fireblight
Polygalacturonase inhibitors	bacterial and fungal pathogens like apple scab or fireblight
Prf regulatory gene	bacterial and fungal pathogens like apple scab or fireblight

US 7,105,469 B2

31

TABLE A12-continued

<u>Crop Pome fruits eg apples, pears</u>	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
phytoalexins	bacterial and fungal pathogens like apple scab or fireblight
B-1,3-glucanase antisense	bacterial and fungal pathogens like apple scab or fireblight
receptor kinase	bacterial and fungal pathogens like apple scab or fireblight
Hypersensitive response eliciting polypeptide	bacterial and fungal pathogens like apple scab or fireblight
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Lytic protein	bacterial and fungal pathogens like apple scab or fireblight
Lysozym	bacterial and fungal pathogens like apple scab or fireblight
Chitinases	bacterial and fungal pathogens like apple scab or fireblight
Barnase	bacterial and fungal pathogens like apple scab or fireblight
Glucanases	bacterial and fungal pathogens like apple scab or fireblight
double stranded ribonuclease	viruses
Coat proteins	viruses
17 kDa or 60 kDa protein	viruses
Nuclear inclusion proteins eg. a or b or Nucleoprotein	viruses
Pseudobiquitin	viruses
Replicase	viruses
<i>Bacillus thuringiensis</i> toxins, VIP 3, <i>Bacillus cereus</i> toxins, Photorabidus and <i>Xenorhabdus</i> toxins	lepidoptera, aphids, mites
3-Hydroxysteroid oxidase	lepidoptera, aphids, mites
Peroxidase	lepidoptera, aphids, mites
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera, aphids, mites
Lectines	lepidoptera, aphids, mites
Protease Inhibitors eg cystatin, patatin, CPTI	lepidoptera, aphids, mites
ribosome inactivating protein	lepidoptera, aphids, mites
stilbene synthase	lepidoptera, aphids, diseases, mites
HMG-CoA reductase	lepidoptera, aphids, mites
Cyst nematode hatching stimulus	cyst nematodes
Barnase	nematodes eg root knot nematodes and cyst nematodes
CBI	root knot nematodes
Antifeeding principles induced at a nematode feeding site	nematodes eg root knot nematodes, root cyst nematodes

TABLE A13

<u>Crop Melons</u>	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazolopyrimidines, Pyrimidylxybenzoates, Phthalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkanecarboxylic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxachlortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis

32

TABLE A13-continued

<u>Crop Melons</u>	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
5 Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxynil
10 5-Enolpyruvyl-3-phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
15 Cytochrome P450 eg. P450 SU1 or selection	Xenobiotics and herbicides such as Sulfonylureas
Polyphenol oxidase or Polyphenol oxidase antisense	bacterial or fungal pathogens like phytophthora
20 Metallothionein	bacterial or fungal pathogens like phytophthora
Ribonuclease	bacterial or fungal pathogens like phytophthora
Antifungal polypeptide AlyAFP	bacterial or fungal pathogens like phytophthora
25 oxalate oxidase	bacterial or fungal pathogens like phytophthora
glucose oxidase	bacterial or fungal pathogens like phytophthora
pyrrolnitrin synthesis genes	bacterial or fungal pathogens like phytophthora
30 serine/threonine kinases	bacterial or fungal pathogens like phytophthora
Cecropin B	bacterial or fungal pathogens like phytophthora
Phenylalanine ammonia lyase (PAL)	bacterial or fungal pathogens like phytophthora
Cf genes eg. Cf 9 Cf5 Cf4 Cf2	bacterial or fungal pathogens like phytophthora
35 Osmotin	bacterial or fungal pathogens like phytophthora
Alpha Hordothionin	bacterial or fungal pathogens like phytophthora
Systemin	bacterial or fungal pathogens like phytophthora
40 Polygalacturonase inhibitors	bacterial or fungal pathogens like phytophthora
Prf regulatory gene	bacterial or fungal pathogens like phytophthora
phytoalexins	bacterial or fungal pathogens like phytophthora
45 B-1,3-glucanase antisense	bacterial or fungal pathogens like phytophthora
receptor kinase	bacterial or fungal pathogens like phytophthora
Hypersensitive response eliciting polypeptide	bacterial or fungal pathogens like phytophthora
50 Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Lytic protein	bacterial or fungal pathogens like phytophthora
Lysozym	bacterial or fungal pathogens like phytophthora
55 Chitinases	bacterial or fungal pathogens like phytophthora
Barnase	bacterial or fungal pathogens like phytophthora
Glucanases	bacterial or fungal pathogens like phytophthora
60 double stranded ribonuclease	viruses as CMV, PRSV, WMV2, SMV, ZYMV
Coat proteins	viruses as CMV, PRSV, WMV2, SMV, ZYMV
17 kDa or 60 kDa protein	viruses as CMV, PRSV, WMV2, SMV, ZYMV
Nuclear inclusion proteins eg. a or b or Nucleoprotein	viruses as CMV, PRSV, WMV2, SMV, ZYMV
65 Pseudobiquitin	viruses as CMV, PRSV, WMV2

US 7,105,469 B2

33

TABLE A13-continued

Crop Melons	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
Replicase	SMV, ZYMV viruses as CMV, PRSV, WMV2, SMV, ZYMV
<i>Bacillus thuringiensis</i> toxins, VIP 3, <i>Bacillus cereus</i> toxins, Photobabidus and <i>Xenorhabdus</i> toxins	lepidoptera, aphids, mites
3-Hydroxysteroid oxidase	lepidoptera, aphids, mites, whitefly
Peroxidase	lepidoptera, aphids, mites, whitefly
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera, aphids, mites, whitefly
Lectines	lepidoptera, aphids, mites, whitefly
Protease Inhibitors eg cystatin, patatin, CPTI, virgiferin	lepidoptera, aphids, mites, whitefly
ribosome inactivating protein	lepidoptera, aphids, mites, whitefly
stilbene synthase	lepidoptera, aphids, mites, whitefly
HMG-CoA reductase	lepidoptera, aphids, mites, whitefly
Cyst nematode hatching stimulus	cyst nematodes
Barnase	nematodes eg root knot nematodes and cyst nematodes
CBI	root knot nematodes
Antifeeding principles induced at a nematode feeding site	nematodes eg root knot nematodes, root cyst nematodes

TABLE A14

Crop Banana	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazolopyrimidines,
AcetylCoA Carboxylase (ACCase)	Pyrimidylloxybenzoates, Phthalides
Hydroxyphenylpyruvate dioxxygenase (HPPD)	Aryloxyphenoxyalkancarboxylic acids, cyclohexanediones
Phosphinothricin acetyl transferase	Isoxazoles such as Isoxaflutol or Isoxaclortol, Triones such as mesotrione or sulcotrione
O-Methyl transferase	Phosphinothricin
Glutamine synthetase	altered lignin levels
Adenylosuccinate Lyase (ADSL)	Glufosinate, Bialaphos
Adenylosuccinate Synthase	Inhibitors of IMP and AMP synthesis
Anthranilate Synthase	Inhibitors of adenylosuccinate synthesis
Nitrilase	Inhibitors of tryptophan syn- thesis and catabolism
5-Enolpyruvyl-3-phosphoshikimate Synthase (EPSPS)	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and loxynil
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Glyphosate or sulfosate
Cytochrome P450 eg. P450 SU1 or selection	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Polyphenol oxidase or Polyphenol oxidase antisense	Xenobiotics and herbicides such as Sulfonylureas
Metallothionein	bacterial or fungal pathogens
Ribonuclease	bacterial or fungal pathogens
Antifungal polypeptide AlyAFP	bacterial or fungal pathogens
oxalate oxidase	bacterial or fungal pathogens

34

TABLE A14-continued

Crop Banana	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
glucose oxidase	bacterial or fungal pathogens
pyrrolnitrin synthesis genes	bacterial or fungal pathogens
serine/threonine kinases	bacterial or fungal pathogens
10 Cecropin B	bacterial or fungal pathogens
Phenylalanine ammonia lyase (PAL)	bacterial or fungal pathogens
Cf genes eg. Cf 9 Cf5 Cf4 Cf2	bacterial or fungal pathogens
Osmotin	bacterial or fungal pathogens
Alpha Hordothionin	bacterial or fungal pathogens
15 Systemin	bacterial or fungal pathogens
Polygalacturonase inhibitors	bacterial or fungal pathogens
Prf regulatory gene	bacterial or fungal pathogens
phytoalexins	bacterial or fungal pathogens
B-1,3-glucanase antisense	bacterial or fungal pathogens
receptor kinase	bacterial or fungal pathogens
20 Hypersensitive response eliciting polypeptide	bacterial or fungal pathogens
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Lytic protein	bacterial or fungal pathogens
Lysozym	bacterial or fungal pathogens
Chitinases	bacterial or fungal pathogens
25 Barnase	bacterial or fungal pathogens
Glucanases	bacterial or fungal pathogens
double stranded ribonuclease	viruses as Banana bunchy top virus (BBTV)
Coat proteins	viruses as Banana bunchy top virus (BBTV)
30 17 kDa or 60 kDa protein	viruses as Banana bunchy top virus (BBTV)
Nuclear inclusion proteins eg. a or b or Nucleoprotein	viruses as Banana bunchy top virus (BBTV)
Pseudoubiquitin	viruses as Banana bunchy top virus (BBTV)
35 Replicase	viruses as Banana bunchy top virus (BBTV)
<i>Bacillus thuringiensis</i> toxins, VIP 3, <i>Bacillus cereus</i> toxins, Photobabidus and <i>Xenorhabdus</i> toxins	lepidoptera, aphids, mites, nematodes
3-Hydroxysteroid oxidase	lepidoptera, aphids, mites, nematodes
40 Peroxidase	lepidoptera, aphids, mites, nematodes
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera, aphids, mites, nematodes
Lectines	lepidoptera, aphids, mites, nematodes
45 Protease Inhibitors eg cystatin, patatin, CPTI, virgiferin	lepidoptera, aphids, mites, nematodes
ribosome inactivating protein	lepidoptera, aphids, mites, nematodes
stilbene synthase	lepidoptera, aphids, mites, nematodes
50 HMG-CoA reductase	lepidoptera, aphids, mites, nematodes
Cyst nematode hatching stimulus	cyst nematodes
Barnase	nematodes eg root knot nematodes and cyst nematodes
55 CBI	root knot nematodes
Antifeeding principles induced at a nematode feeding site	nematodes eg root knot nematodes, root cyst nematodes

TABLE A15

Crop Cotton	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
65 Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazolopyrimidines,

US 7,105,469 B2

35

TABLE A15-continued

Crop Cotton	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
AcetylCoA Carboxylase (ACCase)	Pyrimidylxybenzoates, Phthalides
Hydroxyphenylpyruvate dioxygenase (HPPD)	Aryloxyphenoxyalkancarboxylic acids, cyclohexanediones
Phosphinothricin acetyl transferase	Isoxazoles such as Isoxaflutol or Isoxaclortol, Triones such as mesotrione or sulcotrione
O-Methyl transferase	Phosphinothricin
Glutamine synthetase	altered lignin levels
Adenylosuccinate Lyase (ADSL)	Glufosinate, Bialaphos
Adenylosuccinate Synthase	Inhibitors of IMP and AMP synthesis
Anthranilate Synthase	Inhibitors of adenylosuccinate synthesis
Nitrilase	Inhibitors of tryptophan synthesis and catabolism
5-Enolpyruvyl-3-phosphoshikimate Synthase (EPSPS)	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and loxynil
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1 or selection	Xenobiotics and herbicides such as Sulfonylureas
Polyphenol oxidase or Polyphenol oxidase antisense	bacterial or fungal pathogens
Metallothionein	bacterial or fungal pathogens
Ribonuclease	bacterial or fungal pathogens
Antifungal polypeptide AlyAFP	bacterial or fungal pathogens
oxalate oxidase	bacterial or fungal pathogens
glucose oxidase	bacterial or fungal pathogens
pyrrolnitrin synthesis genes	bacterial or fungal pathogens
serine/threonine kinases	bacterial or fungal pathogens
Cecropin B	bacterial or fungal pathogens
Phenylalanine ammonia lyase (PAL)	bacterial or fungal pathogens
Cf genes eg. Cf 9 Cf5 Cf4 Cf2	bacterial or fungal pathogens
Osmotin	bacterial or fungal pathogens
Alpha Hordothionin	bacterial or fungal pathogens
Systemin	bacterial or fungal pathogens
Polygalacturonase inhibitors	bacterial or fungal pathogens
Prf regulatory gene	bacterial or fungal pathogens
phytoalexins	bacterial or fungal pathogens
B-1,3-glucanase antisense	bacterial or fungal pathogens
receptor kinase	bacterial or fungal pathogens
Hypersensitive response eliciting polypeptide	bacterial or fungal pathogens
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Lytic protein	bacterial or fungal pathogens
Lysozym	bacterial or fungal pathogens
Chitinases	bacterial or fungal pathogens
Barnase	bacterial or fungal pathogens
Glucanases	bacterial or fungal pathogens
double stranded ribonuclease	viruses as wound tumor virus (WTV)
Coat proteins	viruses as wound tumor virus (WTV)
17 kDa or 60 kDa protein	viruses as wound tumor virus (WTV)
Nuclear inclusion proteins eg. a or b or Nucleoprotein	viruses as wound tumor virus (WTV)
Pseudoubiquitin	viruses as wound tumor virus (WTV)
Replicase	viruses as wound tumor virus (WTV)
<i>Bacillus thuringiensis</i> toxins, VIP 3, <i>Bacillus cereus</i> toxins, <i>Photobacterium</i> and <i>Xenorhabdus</i> toxins	lepidoptera, aphids, mites, nematodes, whitefly
3 Hydroxysteroid oxidase	lepidoptera, aphids, mites, nematodes, whitefly

36

TABLE A15-continued

Crop Cotton	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
Peroxidase	lepidoptera, aphids, mites, nematodes, whitefly
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera, aphids, mites, nematodes, whitefly
Lectines	lepidoptera, aphids, mites, nematodes, whitefly
Protease Inhibitors eg cystatin, patatin, CPTI, virgiferin	lepidoptera, aphids, mites, nematodes, whitefly
ribosome inactivating protein	lepidoptera, aphids, mites, nematodes, whitefly
stilbene synthase	lepidoptera, aphids, mites, nematodes, whitefly
HMG-CoA reductase	lepidoptera, aphids, mites, nematodes, whitefly
Cyst nematode hatching stimulus	cyst nematodes
Barnase	nematodes eg root knot nematodes and cyst nematodes
CBI	root knot nematodes
Antifeeding principles induced at a nematode feeding site	nematodes eg root knot nematodes, root cyst nematodes

TABLE A16

Crop Sugarcane	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazolopyrimidines, Pyrimidylxybenzoates, Phthalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkancarboxylic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxaclortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and loxynil
5-Enolpyruvyl-3-phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1 or selection	Xenobiotics and herbicides such as Sulfonylureas
Polyphenol oxidase or Polyphenol oxidase antisense	bacterial or fungal pathogens
Metallothionein	bacterial or fungal pathogens
Ribonuclease	bacterial or fungal pathogens
Antifungal polypeptide AlyAFP	bacterial or fungal pathogens
oxalate oxidase	bacterial or fungal pathogens
glucose oxidase	bacterial or fungal pathogens
pyrrolnitrin synthesis genes	bacterial or fungal pathogens
serine/threonine kinases	bacterial or fungal pathogens
Cecropin B	bacterial or fungal pathogens
Phenylalanine ammonia lyase (PAL)	bacterial or fungal pathogens
Cf genes eg. Cf 9 Cf5 Cf4 Cf2	bacterial or fungal pathogens

US 7,105,469 B2

37

TABLE A16-continued

Effectuated target or expressed principle(s)	Crop phenotype/Tolerance to
Osmotin	bacterial or fungal pathogens
Alpha Hordothionin	bacterial or fungal pathogens
Systemin	bacterial or fungal pathogens
Polygalacturonase inhibitors	bacterial or fungal pathogens
Prf regulatory gene	bacterial or fungal pathogens
phytoalexins	bacterial or fungal pathogens
B-1,3-glucanase antisense	bacterial or fungal pathogens
receptor kinase	bacterial or fungal pathogens
Hypersensitive response eliciting polypeptide	bacterial or fungal pathogens
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Lytic protein	bacterial or fungal pathogens
Lysozym	bacterial or fungal pathogens eg clavibacter
Chitinases	bacterial or fungal pathogens
Barnase	bacterial or fungal pathogens
Glucanases	bacterial or fungal pathogens
double stranded ribonuclease	viruses as SCMV, SrMV
Coat proteins	viruses as SCMV, SrMV
17 kDa or 60 kDa protein	viruses as SCMV, SrMV
Nuclear inclusion proteins eg. a or b or Nucleoprotein	viruses as SCMV, SrMV
Pseudobiquitin	viruses as SCMV, SrMV
Replicase	viruses as SCMV, SrMV
<i>Bacillus thuringiensis</i> toxins, VIP 3, <i>Bacillus cereus</i> toxins, <i>Photorabdus</i> and <i>Xenorhabdus</i> toxins	lepidoptera, aphids, mites, nematodes, whitefly, beetles eg mexican rice borer
3-Hydroxysteroid oxidase	lepidoptera, aphids, mites, nematodes, whitefly, beetles eg mexican rice borer
Peroxidase	lepidoptera, aphids, mites, nematodes, whitefly, beetles eg mexican rice borer
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera, aphids, mites, nematodes, whitefly, beetles eg mexican rice borer
Lectines	lepidoptera, aphids, mites, nematodes, whitefly, beetles eg mexican rice borer
Protease Inhibitors eg cystatin, patatin, CPTI, virgiferin	lepidoptera, aphids, mites, nematodes, whitefly, beetles eg mexican rice borer
ribosome inactivating protein	lepidoptera, aphids, mites, nematodes, whitefly, beetles eg mexican rice borer
stilbene synthase	lepidoptera, aphids, mites, nematodes, whitefly, beetles eg mexican rice borer
HMG-CoA reductase	lepidoptera, aphids, mites, nematodes, whitefly, beetles eg mexican rice borer
Cyst nematode hatching stimulus	cyst nematodes
Barnase	nematodes eg root knot nematodes and cyst nematodes
CBI	root knot nematodes
Antifeeding principles induced at a nematode feeding site	nematodes eg root knot nematodes, root cyst nematodes

TABLE A17

Effectuated target or expressed principle(s)	Crop phenotype/Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazopyrimidines, Pyrimidylxybenzoates, Phthalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkancarboxylic acids, cyclohexanediones

38

TABLE A17-continued

Effectuated target or expressed principle(s)	Crop phenotype/Tolerance to
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxaclortol, Trione such as mesotrione or sulcotrione Phosphinothricin
Phosphinothricin acetyl transferase	altered lignin levels
O-Methyl transferase	Glufosinate, Bialaphos
Glutamine synthetase	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Lyase (ADSL)	Inhibitors of adenylosuccinate synthesis
Adenylosuccinate Synthase	Inhibitors of tryptophan synthesis and catabolism
Anthranilate Synthase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxynil
Nitrilase	Glyphosate or sulfosate
5-Enolpyruvyl-3-phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenoplylate, oxadiazoles etc.
Protoporphyrinogen oxidase (PROTOX)	Xenobiotics and herbicides such as Sulfonylureas
Cytochrome P450 eg. P450 SU1 or selection	bacterial or fungal pathogens
Polyphenol oxidase or Polyphenol oxidase antisense	bacterial or fungal pathogens
Metallothionein	bacterial or fungal pathogens
Ribonuclease	bacterial or fungal pathogens
Antifungal polypeptide AlyAFP	bacterial or fungal pathogens
oxalate oxidase	eg <i>sclerotinia</i>
glucose oxidase	bacterial or fungal pathogens
pyrrolnitrin synthesis genes	bacterial or fungal pathogens
serine/threonine kinases	bacterial or fungal pathogens
Cecropin B	bacterial or fungal pathogens
Phenylalanine ammonia lyase (PAL)	bacterial or fungal pathogens
Cf genes eg. Cf 9 Cf5 Cf4 Cf2	bacterial or fungal pathogens
Osmotin	bacterial or fungal pathogens
Alpha Hordothionin	bacterial or fungal pathogens
Systemin	bacterial or fungal pathogens
Polygalacturonase inhibitors	bacterial or fungal pathogens
Prf regulatory gene	bacterial or fungal pathogens
phytoalexins	bacterial or fungal pathogens
B-1,3-glucanase antisense	bacterial or fungal pathogens
receptor kinase	bacterial or fungal pathogens
Hypersensitive response eliciting polypeptide	bacterial or fungal pathogens
Systemic acquires resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Lytic protein	bacterial or fungal pathogens
Lysozym	bacterial or fungal pathogens
Chitinases	bacterial or fungal pathogens
Barnase	bacterial or fungal pathogens
Glucanases	bacterial or fungal pathogens
double stranded ribonuclease	viruses as CMV, TMV
Coat proteins	viruses as CMV, TMV
17 kDa or 60 kDa protein	viruses as CMV, TMV
Nuclear inclusion proteins eg. a or b or Nucleoprotein	viruses as CMV, TMV
Pseudobiquitin	viruses as CMV, TMV
Replicase	viruses as CMV, TMV
<i>Bacillus thuringiensis</i> toxins, VIP 3, <i>Bacillus cereus</i> toxins, <i>Photorabdus</i> and <i>Xenorhabdus</i> toxins	lepidoptera, aphids, mites, nematodes, whitefly, beetles
3-Hydroxysteroid oxidase	lepidoptera, aphids, mites, nematodes, whitefly, beetles
Peroxidase	lepidoptera, aphids, mites, nematodes, whitefly, beetles
Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera, aphids, mites, nematodes, whitefly, beetles
Lectines	lepidoptera, aphids, mites, nematodes, whitefly, beetles
Protease Inhibitors eg cystatin,	lepidoptera, aphids, mites,

US 7,105,469 B2

39

TABLE A17-continued

Crop Sunflower	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
patatin, CPTI, virgiferin	nematodes, whitefly, beetles
ribosome inactivating protein	lepidoptera, aphids, mites, nematodes, whitefly, beetles
stilbene synthase	lepidoptera, aphids, mites, nematodes, whitefly, beetles
HMG-CoA reductase	lepidoptera, aphids, mites, nematodes, whitefly, beetles
Cyst nematode hatching stimulus	cyst nematodes
Barnase	nematodes eg root knot nematodes and cyst nematodes
CBI	root knot nematodes
Antifeeding principles induced at a nematode feeding site	nematodes eg root knot nematodes, root cyst nematodes

TABLE A18

Crop Sugarbeet, Beet root	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
Acetolactate synthase (ALS)	Sulfonylureas, Imidazolinones, Triazopyrimidines, Pyrimidylxybenzoates, Pthalides
AcetylCoA Carboxylase (ACCase)	Aryloxyphenoxyalkylcarboxylic acids, cyclohexanediones
Hydroxyphenylpyruvate dioxygenase (HPPD)	Isoxazoles such as Isoxaflutol or Isoxaclortol, Triones such as mesotrione or sulcotrione
Phosphinothricin acetyl transferase	Phosphinothricin
O-Methyl transferase	altered lignin levels
Glutamine synthetase	Glufosinate, Bialaphos
Adenylosuccinate Lyase (ADSL)	Inhibitors of IMP and AMP synthesis
Adenylosuccinate Synthase	Inhibitors of adenylosuccinate synthesis
Anthranilate Synthase	Inhibitors of tryptophan synthesis and catabolism
Nitrilase	3,5-dihalo-4-hydroxy-benzonitriles such as Bromoxynil and Ioxynil
5-Enolpyruvyl-3-phosphoshikimate Synthase (EPSPS)	Glyphosate or sulfosate
Glyphosate oxidoreductase	Glyphosate or sulfosate
Protoporphyrinogen oxidase (PROTOX)	Diphenylethers, cyclic imides, phenylpyrazoles, pyridin derivatives, phenopylate, oxadiazoles etc.
Cytochrome P450 eg. P450 SU1 or selection	Xenobiotics and herbicides such as Sulfonylureas
Polyphenol oxidase or Polyphenol oxidase antisense	bacterial or fungal pathogens
Metallothionein	bacterial or fungal pathogens
Ribonuclease	bacterial or fungal pathogens
Antifungal polypeptide AlyAFP	bacterial or fungal pathogens
oxalate oxidase	eg <i>sclerotinia</i>
glucose oxidase	bacterial or fungal pathogens
pyrrolnitrin synthesis genes	bacterial or fungal pathogens
serine/threonine kinases	bacterial or fungal pathogens
Cecropin B	bacterial or fungal pathogens
Phenylalanine ammonia lyase (PAL)	bacterial or fungal pathogens
Cf genes eg. Cf 9 Cf5 Cf4 Cf2	bacterial or fungal pathogens
Osmotin	bacterial or fungal pathogens
Alpha Hordothionin	bacterial or fungal pathogens
Systemin	bacterial or fungal pathogens
Polygalacturonase inhibitors	bacterial or fungal pathogens
Prf regulatory gene	bacterial or fungal pathogens
phytoalexins	bacterial or fungal pathogens
B-1,3 glucanase antisense	bacterial or fungal pathogens

40

TABLE A18-continued

Crop Sugarbeet, Beet root	
Effected target or expressed principle(s)	Crop phenotype/Tolerance to
AX + WIN proteins	bacterial or fungal pathogens like <i>Cercospora beticola</i>
receptor kinase	bacterial or fungal pathogens
10 Hypersensitive response eliciting polypeptide	bacterial or fungal pathogens
Systemic acquired resistance (SAR) genes	viral, bacterial, fungal, nematodal pathogens
Lytic protein	bacterial or fungal pathogens
Lysozym	bacterial or fungal pathogens
15 Chitinases	bacterial or fungal pathogens
Barnase	bacterial or fungal pathogens
Glucanases	bacterial or fungal pathogens
double stranded ribonuclease	viruses as BNYVV
Coat proteins	viruses as BNYVV
17 kDa or 60 kDa protein	viruses as BNYVV
20 Nuclear inclusion proteins eg. a or b or Nucleoprotein	viruses as BNYVV
Pseudoubiquitin	viruses as BNYVV
Replicase	viruses as BNYVV
<i>Bacillus thuringiensis</i> toxins, VIP 3, <i>Bacillus cereus</i> toxins, Photorabdus and <i>Xenorhabdus</i> toxins	lepidoptera, aphids, mites, nematodes, whitefly, beetles, rootflies
25 3-Hydroxysteroid oxidase	lepidoptera, aphids, mites, nematodes, whitefly, beetles, rootflies
Peroxidase	lepidoptera, aphids, mites, nematodes, whitefly, beetles, rootflies
30 Aminopeptidase inhibitors eg. Leucine aminopeptidase inhibitor	lepidoptera, aphids, mites, nematodes, whitefly, beetles, rootflies
Lectines	lepidoptera, aphids, mites, nematodes, whitefly, beetles, rootflies
35 Protease Inhibitors eg cystatin, patatin, CPTI, virgiferin	lepidoptera, aphids, mites, nematodes, whitefly, beetles, rootflies
ribosome inactivating protein	lepidoptera, aphids, mites, nematodes, whitefly, beetles, rootflies
40 stilbene synthase	lepidoptera, aphids, mites, nematodes, whitefly, beetles, rootflies
HMG-CoA reductase	lepidoptera, aphids, mites, nematodes, whitefly, beetles, rootflies
45 Cyst nematode hatching stimulus	cyst nematodes
Barnase	nematodes eg root knot nematodes and cyst nematodes
Beet cyst nematode resistance locus	cyst nematodes
CBI	root knot nematodes
Antifeeding principles induced at a nematode feeding site	nematodes eg root knot nematodes, root cyst nematodes
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The abovementioned animal pests which can be controlled by the method according to the invention include, for example, insects, representatives of the order acarina and representatives of the class nematoda; especially from the order Lepidoptera *Acleris* spp., *Adoxophyes* spp., especially *Adoxophyes reticulana*; *Aegeria* spp., *Agrotis* spp., especially *Agrotis spinifera*; *Alabama argillaceae*, *Amylois* spp., *Anticarsia gemmatilis*, *Archips* spp., *Argyrotaenia* spp., *Autographa* spp., *Busseola fusca*, *Cadra cautella*, *Carposina nipponensis*, *Chilo* spp., *Choristoneura* spp., *Clysia ambiguella*, *Cnaphalocrocis* spp., *Cnephasia* spp., *Cochylis* spp., *Coleophora* spp., *Crocidolomia binotalis*, *Cryptophlebia leucotreta*, *Cydia* spp., especially *Cydia pomonella*; *Diatraea* spp., *Diparopsis castanea*, *Earias* spp., *Ephestia* spp., especially *E. Khü-*

US 7,105,469 B2

41

niella; *Eucosma* spp., *Eupoecilia ambiguella*, *Euproctis* spp., *Euxoa* spp., *Grapholita* spp., *Hedya nubiferana*, *Heliothis* spp., especially *H. virescens* und *H. zea*; *Helicula undalis*, *Hyphantria cunea*, *Keiferia lycopersicella*, *Leucoptra scitella*, *Lithocolletis* spp., *Lobesia* spp., *Lymantria* spp., *Lyonetia* spp., *Malacosoma* spp., *Mamestra brassicae*, *Manduca sexta*, *Operophtera* spp., *Ostrinia nubilalis*, *Pammene* spp., *Pandemis* spp., *Panolis flammea*, *Pectinophora* spp., *Phthorimaea operculella*, *Pieris rapae*, *Pieris* spp., *Plutella xylostella*, *Prays* spp., *Scirpophaga* spp., *Sesamia* spp., *Sparganothis* spp., *Spodopteralittoralis*, *Synanthedon* spp., *Thaumetopoea* spp., *Tortrix* spp., *Trichoplusia ni* and *Yponomeuta* spp.; from the order Coleoptera, for example *Agriotes* spp., *Anthonomus* spp., *Atomaria linearis*, *Chaetocnema tibialis*, *Cosmopolites* spp., *Curculio* spp., *Dermestes* spp., *Diabrotica* spp., *Epilachna* spp., *Eremnus* spp., *Leptinotarsa decemlineata*, *Lissorhoptrus* spp., *Melolontha* spp., *Oryzaephilus* spp., *Otiorynchus* spp., *Phlyctinus* spp., *Popillia* spp., *Psylliodes* spp., *Rhizopertha* spp., *Scarabaeidae*, *Sitophilus* spp., *Sitotroga* spp., *Tenebrio* spp., *Tribolium* spp. and *Trogoderma* spp.; from the order Orthoptera, for example *Blatta* spp., *Blattella* spp., *Gryllotalpa* spp., *Leucophaea maderae*, *Locusta* spp., *Periplaneta* spp. and *Schistocerca* spp.; from the order Isoptera, for example *Reticulitermes* spp.; from the order Psocoptera, for example *Liposcelis* spp.; from the order Anoplura, for example *Haematopinus* spp., *Linognathus* spp., *Pediculus* spp., *Pemphigus* spp. and *Phylloxera* spp.; from the order Mallophaga, for example *Damalinea* spp. and *Trichodectes* spp.; from the order Thysanoptera, for example *Frankliniella* spp., *Hercinothrips* spp., *Taeniothrips* spp., *Thrips palmi*, *Thrips tabaci* and *Scirtothrips aurantii*; from the order Heteroptera, for example *Cimex* spp., *Distantiella theobroma*, *Dysdercus* spp., *Euchistus* spp., *Eurygaster* spp., *Leptocoris* spp., *Nezara* spp., *Piesma* spp., *Rhodnius* spp., *Sahlbergella singularis*, *Scotinophara* spp. and *Triatoma* spp.; from the order Homoptera, for example *Aleurothrixus floccosus*, *Aleyrodes brassicae*, *Aonidiella aurantii*, *Aphididae*, *Aphis craccivora*, *A. fabae*, *A. gossypii*, *Aspidiotus* spp., *Bemisia tabaci*, *Ceroplaster* spp., *Chrysomphalus aonidium*, *Chrysomphalus dictyospermi*, *Coccus hesperidum*, *Empoasca* spp., *Eriosoma lanigerum*, *Erythroneura* spp., *Gascardia* spp., *Laodelphax* spp., *Lecanium corni*, *Lepidosaphes* spp., *Macrosiphus* spp., *Myzus* spp., especially *M. persicae*; *Nephotettix* spp., especially *N. cincticeps*; *Nilaparvata* spp., especially *N. lugens*; *Paratoria* spp., *Pemphigus* spp., *Planococcus* spp., *Pseudaulacaspis* spp., *Pseudococcus* spp., especially *P. fragilis*, *P. citriculus* and *P. comstocki*; *Psylla* spp., especially *P. pyri*; *Pulvinaria aethiopica*, *Quadraspidiotus* spp., *Rhopalosiphum* spp., *Saissetia* spp., *Scaphoideus* spp., *Schizaphis* spp., *Sitobion* spp., *Trialeurodes vaporariorum*, *Trioza erytrae* and *Unaspis citri*; from the order Hymenoptera, for example *Acromyrmex*, *Atta* spp., *Cephus* spp., *Diprion* spp., *Diprionidae*, *Gilpinia polytoma*, *Hoplocampa* spp., *Lasius* spp., *Monomorium pharaonis*, *Neodiprion* spp., *Solenopsis* spp. and *Vespa* spp.; from the order Diptera, for example *Aedes* spp., *Antherigona soccata*, *Bibio hortulanus*, *Calliphora erythrocephala*, *Ceratitis* spp., *Chrysomyia* spp., *Culex* spp., *Cuterebra* spp., *Dacus* spp., *Drosophila melanogaster*, *Fannia* spp., *Gastrophilus* spp., *Glossina* spp., *Hypoderma* spp., *Hyp-*

42

pobosca spp., *Liriomyza* spp., *Lucilia* spp., *Melanogromyza* spp., *Musca* spp., *Oestrus* spp., *Orseolia* spp., *Oscinella frit*, *Pegomyia hyoscyami*, *Phorbia* spp., *Rhagoletis pomonella*, *Sciara* spp., *Stomoxys* spp., *Tabanus* spp., *Tannia* spp. and *Tipula* spp.; from the order Siphonaptera, for example *Ceratophyllus* spp. and *Xenopsylla cheopis*; from the order Thysanura, for example *Lepisma saccharina* and from the order Acarina, for example *Acarus siro*, *Aceria sheldoni*, *Aculus* spp., especially *A. schlechtendali*; *Amblyomma* spp., *Argas* spp., *Boophilus* spp., *Brevipalpus* spp., especially *B. californicus* and *B. phoenicis*; *Bryobia praetiosa*, *Calipitimerus* spp., *Chorioptes* spp., *Dermanyssus gallinae*, *Eotetranychus* spp., especially *E. carpini* and *E. orientalis*; *Eriophyes* spp., especially *E. vitis*; *Hyalomma* spp., *Ixodes* spp., *Oligonychus pratensis*, *Ornithodoros* spp., *Panonychus* spp., especially *P. ulmi* and *P. citri*; *Phyllocoptura* spp., especially *P. oleivora*; *Polyphagotarsonemus* spp., especially *P. latus*; *Psoroptes* spp., *Rhipicephalus* spp., *Rhizoglyphus* spp., *Sarcoptes* spp., *Tarsonemus* spp. and *Tetranychus* spp., in particular *T. urticae*, *T. cinnabarinus* and *T. Kanzawai*; 25 Representatives of the Class Nematoda; (1) nematodes selected from the group consisting of root knot nematodes, cyst-forming nematodes, stem eelworms and foliar nematodes; (2) nematodes selected from the group consisting of 30 *Anguina* spp.; *Aphelenchoides* spp.; *Ditylenchus* spp.; *Globodera* spp., for example *Globodera rostochiensis*; *Heterodera* spp., for example *Heterodera avenae*, *Heterodera glycines*, *Heterodera schachtii* or *Heterodera trifolii*; *Longidorus* spp.; *Meloidogyne* spp., for example *Meloidogyne incognita* or *Meloidogyne javanica*; *Pratylenchus*, for example *Pratylenchus neglectans* or *Pratylenchus penetrans*; *Radopholus* spp., for example *Radopholus similis*; *Trichodorus* spp.; *Tylenchulus*, for example *Tylenchulus semipenetrans*; and *Xiphinema* spp.; or 35 (3) nematodes selected from the group consisting of *Heterodera* spp., for example *Heterodera glycines*; and *Meloidogyne* spp., for example *Meloidogyne incognita*. The method according to the invention allows pests of the abovementioned type to be controlled, i.e. contained or 40 destroyed, which occur, in particular, on transgenic plants, mainly useful plants and ornamentals in agriculture, in horticulture and in forests, or on parts, such as fruits, flowers, foliage, stalks, tubers or roots, of such plants, the protection against these pests in some cases even extending to plant parts which form at a later point in time. The method according to the invention can be employed advantageously for controlling pests in rice, cereals such as 45 maize or sorghum; in fruit, for example stone fruit, pome fruit and soft fruit such as apples, pears, plums, peaches, almonds, cherries or berries, for example strawberries, raspberries and blackberries; in legumes such as beans, lentils, peas or soya beans; in oil crops such as oilseed rape, mustard, poppies, olives, sunflowers, coconuts, castor-oil plants, cacao or peanuts; in the marrow family such as pumpkins, cucumbers or melons; in fibre plants such as cotton, flax, hemp or jute; in citrus fruit such as oranges, lemons, grapefruit or tangerines; in vegetables such as spinach, lettuce, asparagus, cabbage species, carrots, onions, tomatoes, potatoes, beet or *capsicum*; in the laurel family 50 such as avocado, Cinnamomum or camphor; or in tobacco, nuts, coffee, egg plants, sugar cane, tea, pepper, grapevines, hops, the banana family, latex plants or ornamentals, mainly

US 7,105,469 B2

43

in maize, rice, cereals, soya beans, tomatoes, cotton, potatoes, sugar beet, rice and mustard; in particular in cotton, rice, soya beans, potatoes and maize.

It has emerged that the method according to the invention is valuable preventatively and/or curatively in the field of pest control even at low use concentrations of the pesticidal composition and that a very favourable biocidal spectrum is achieved thereby. Combined with a favourable compatibility of the composition employed with warm-blooded species, fish and plants, the method according to the invention can be employed against all or individual developmental stages of normally-sensitive, but also of normally-resistant, animal pests such as insects and representatives of the order Acarina, depending on the species of the transgenic crop plant to be protected from attack by pests. The insecticidal and/or acaricidal effect of the method according to the invention may become apparent directly, i.e. in a destruction of the pests which occurs immediately or only after some time has elapsed, for example, during ecdysis, or indirectly, for example as a reduced oviposition and/or hatching rate, the good action corresponding to a destruction rate (mortality) of at least 40 to 50%.

Depending on the intended aims and the prevailing circumstances, the pesticides within the scope of invention, which are known per se, are emulsifiable concentrates, suspension concentrates, directly sprayable or dilutable solutions, spreadable pastes, dilute emulsions, wettable powders, soluble powders, dispersible powders, wettable powders, dusts, granules or encapsulations in polymeric substances which comprise a nitroimino- or nitroguanidino-compound.

The active ingredients are employed in these compositions together with at least one of the auxiliaries conventionally used in art of formulation, such as extenders, for example solvents or solid carriers, or such as surface-active compounds (surfactants).

Formulation auxiliaries which are used are, for example, solid carriers, solvents, stabilizers, "slow release" auxiliaries, colourants and, if appropriate, surface-active substances (surfactants). Suitable carriers and auxiliaries are all those substances which are conventionally used for crop protection products. Suitable auxiliaries such as solvents, solid carriers, surface-active compounds, non-ionic surfactants, cationic surfactants, anionic surfactants and other auxiliaries in the compositions employed according to the invention are, for example, those which have been described in EP-A-736 252.

These compositions for controlling pests can be formulated, for example, as wettable powders, dusts, granules, solutions, emulsifiable concentrates, emulsions, suspension concentrates or aerosols. For example, the compositions are of the type described in EP-A-736 252.

The action of the compositions within the scope of invention which comprise a nitroimino- or nitroguanidino-compound can be extended substantially and adapted to prevailing circumstances by adding other insecticidally, acaricidally and/or fungicidally active ingredients. Suitable examples of added active ingredients are representatives of the following classes of active ingredients: organophosphorous compounds, nitrophenols and derivatives, formamidines, ureas, carbamates, pyrethroids, chlorinated hydrocarbons; especially preferred components in mixtures are, for example, abamectin, emamectin, spinosad, pymetrozine, fenoxycarb, Ti-435, fipronil, pyriproxyfen, diazinon or diafenthiuron.

As a rule, the compositions within the scope of invention comprise 0.1 to 99%, in particular 0.1 to 95%, of a

44

nitroimino- or nitroguanidino-compound and 1 to 99.9%, in particular 5 to 99.9%, of—at least—one solid or liquid auxiliary, it being possible, as a rule, for 0 to 25%, in particular 0.1 to 20%, of the compositions to be surfactants (% in each case meaning percent by weight). While concentrated compositions are more preferred as commercial products, the end user will, as a rule, use dilute compositions which have considerably lower concentrations of active ingredient.

The compositions according to the invention may also comprise other solid or liquid auxiliaries, such as stabilisers, for example epoxidized or unepoxidized vegetable oils (for example epoxidized coconut oil, rapeseed oil or soya bean oil), antifoams, for example silicone oil, preservatives, viscosity regulators, binders and/or tackifiers, and also fertilizers or other active ingredients for achieving specific effects, for example, bactericides, fungicides, nematocides, molluscicides or herbicides.

The compositions according to the invention are produced in a known manner, for example prior to mixing with the auxiliary/auxiliaries by grinding, screening and/or compressing the active ingredient, for example to give a particular particle size, and by intimately mixing and/or grinding the active ingredient with the auxiliary/auxiliaries.

The method according to the invention for controlling pests of the abovementioned type is carried out in a manner known per se to those skilled in the art, depending on the intended aims and prevailing circumstances, that is to say by spraying, wetting, atomizing, dusting, brushing on, seed dressing, scattering or pouring of the composition. Typical use concentrations are between 0.1 and 1000 ppm, preferably between 0.1 and 500 ppm of active ingredient. The application rate may vary within wide ranges and depends on the soil constitution, the type of application (foliar application; seed dressing; application in the seed furrow), the transgenic crop plant, the pest to be controlled, the climatic circumstances prevailing in each case, and other factors determined by the type of application, timing of application and target crop. The application rates per hectare are generally 1 to 2000 g of nitroimino- or nitroguanidino-compound per hectare, in particular 10 to 1000 g/ha, preferably 10 to 500 g/ha, especially preferably 10 to 200 g/ha.

A preferred type of application in the field of crop protection within the scope of invention is application to the foliage of the plants (foliar application), it being possible to adapt frequency and rate of application to the risk of infestation with the pest in question. However, the active ingredient may also enter into the plants via the root system (systemic action), by drenching the site of the plants with a liquid composition or by incorporating the active ingredient in solid form into the site of the plants, for example into the soil, for example in the form of granules (soil application). In the case of paddy rice crops, such granules may be metered into the flooded paddy field.

The compositions according to invention are also suitable for protecting propagation material of transgenic plants, for example seed, such as fruits, tubers or kernels, or plant cuttings, from animal pests, in particular insects and representatives of the order Acarina.

The propagation material can be treated with the composition prior to application, for example, seed being dressed prior to sowing. The active ingredient may also be applied to seed kernels (coating), either by soaking the kernels in a liquid composition or by coating them with a solid composition. The composition may also be applied to the site of application when applying the propagation material, for example into the seed furrow during sowing. These treat-

US 7,105,469 B2

45

ment methods for plant propagation material and the plant propagation material treated thus are a further subject of the invention.

Examples of formulations of nitroimino- or nitroguani-
dino-compounds which can be used in the method according
to the invention, for instance solutions, granules, dusts,
sprayable powders, emulsion concentrates, coated granules
and suspension concentrates, are of the type as has been
described in, for example, EP-A-580 553, Examples F1 to
F10.

BIOLOGICAL EXAMPLES

TABLE B

AP	Control of
B.1 CryIA(a)	<i>Adoxophyes</i> spp.
B.2 CryIA(a)	<i>Agrotis</i> spp.
B.3 CryIA(a)	<i>Alabama argillaceae</i>
B.4 CryIA(a)	<i>Anticarsia gemmatilis</i>
B.5 CryIA(a)	<i>Chilo</i> spp.
B.6 CryIA(a)	<i>Clysia ambiguella</i>
B.7 CryIA(a)	<i>Crocidolomia binotalis</i>
B.8 CryIA(a)	<i>Cydia</i> spp.
B.9 CryIA(a)	<i>Diparopsis castanea</i>
B.10 CryIA(a)	<i>Earias</i> spp.
B.11 CryIA(a)	<i>Ephestia</i> spp.
B.12 CryIA(a)	<i>Heliothis</i> spp.
B.13 CryIA(a)	<i>Helitula undalis</i>
B.14 CryIA(a)	<i>Keiferia lycopersicella</i>
B.15 CryIA(a)	<i>Leucopiera scitella</i>
B.16 CryIA(a)	<i>Lithocollethis</i> spp.
B.17 CryIA(a)	<i>Lobesia botrana</i>
B.18 CryIA(a)	<i>Ostrinia nubilalis</i>
B.19 CryIA(a)	<i>Pandemis</i> spp.
B.20 CryIA(a)	<i>Pectinophora gossyp.</i>
B.21 CryIA(a)	<i>Phyllocnistis citrella</i>
B.22 CryIA(a)	<i>Pieris</i> spp.
B.23 CryIA(a)	<i>Plutella xylostella</i>
B.24 CryIA(a)	<i>Scirpophaga</i> spp.
B.25 CryIA(a)	<i>Sesamia</i> spp.
B.26 CryIA(a)	<i>Sparganothis</i> spp.
B.27 CryIA(a)	<i>Spodoptera</i> spp.
B.28 CryIA(a)	<i>Tortrix</i> spp.
B.29 CryIA(a)	<i>Trichoplusia ni</i>
B.30 CryIA(a)	<i>Agrotis</i> spp.
B.31 CryIA(a)	<i>Anthonomus grandis</i>
B.32 CryIA(a)	<i>Curculio</i> spp.
B.33 CryIA(a)	<i>Diabrotica balteata</i>
B.34 CryIA(a)	<i>Leptinotarsa</i> spp.
B.35 CryIA(a)	<i>Lissorhoptrus</i> spp.
B.36 CryIA(a)	<i>Otiorynchus</i> spp.
B.37 CryIA(a)	<i>Aleurothrixus</i> spp.
B.38 CryIA(a)	<i>Aleyrodes</i> spp.
B.39 CryIA(a)	<i>Aonidiella</i> spp.
B.40 CryIA(a)	<i>Aphididae</i> spp.
B.41 CryIA(a)	<i>Aphis</i> spp.
B.42 CryIA(a)	<i>Bemisia tabaci</i>
B.43 CryIA(a)	<i>Empoasca</i> spp.
B.44 CryIA(a)	<i>Mycus</i> spp.
B.45 CryIA(a)	<i>Nephotettix</i> spp.
B.46 CryIA(a)	<i>Nilaparvata</i> spp.
B.47 CryIA(a)	<i>Pseudococcus</i> spp.
B.48 CryIA(a)	<i>Psylla</i> spp.
B.49 CryIA(a)	<i>Quadrastipidiotus</i> spp.
B.50 CryIA(a)	<i>Schizaphis</i> spp.
B.51 CryIA(a)	<i>Trialeurodes</i> spp.
B.52 CryIA(a)	<i>Lyriomyza</i> spp.
B.53 CryIA(a)	<i>Oscinella</i> spp.
B.54 CryIA(a)	<i>Phorbia</i> spp.
B.55 CryIA(a)	<i>Frankliniella</i> spp.
B.56 CryIA(a)	<i>Thrips</i> spp.
B.57 CryIA(a)	<i>Scirtothrips aurantii</i>
B.58 CryIA(a)	<i>Aceria</i> spp.
B.59 CryIA(a)	<i>Aculus</i> spp.
B.60 CryIA(a)	<i>Brevipalpus</i> spp.

46

TABLE B-continued

AP	Control of
B.61 CryIA(a)	<i>Panonychus</i> spp.
B.62 CryIA(a)	<i>Phyllocoptruta</i> spp.
B.63 CryIA(a)	<i>Tetranychus</i> spp.
B.64 CryIA(a)	<i>Heterodera</i> spp.
B.65 CryIA(a)	<i>Meloidogyne</i> spp.
B.66 CryIA(b)	<i>Adoxophyes</i> spp.
B.67 CryIA(b)	<i>Agrotis</i> spp.
B.68 CryIA(b)	<i>Alabama argillaceae</i>
B.69 CryIA(b)	<i>Anticarsia gemmatilis</i>
B.70 CryIA(b)	<i>Chilo</i> spp.
B.71 CryIA(b)	<i>Clysia ambiguella</i>
B.72 CryIA(b)	<i>Crocidolomia binotalis</i>
B.73 CryIA(b)	<i>Cydia</i> spp.
B.74 CryIA(b)	<i>Diparopsis castanea</i>
B.75 CryIA(b)	<i>Earias</i> spp.
B.76 CryIA(b)	<i>Ephestia</i> spp.
B.77 CryIA(b)	<i>Heliothis</i> spp.
B.78 CryIA(b)	<i>Helitula undalis</i>
B.79 CryIA(b)	<i>Keiferia lycopersicella</i>
B.80 CryIA(b)	<i>Leucopiera scitella</i>
B.81 CryIA(b)	<i>Lithocollethis</i> spp.
B.82 CryIA(b)	<i>Lobesia botrana</i>
B.83 CryIA(b)	<i>Ostrinia nubilalis</i>
B.84 CryIA(b)	<i>Pandemis</i> spp.
B.85 CryIA(b)	<i>Pectinophora gossyp.</i>
B.86 CryIA(b)	<i>Phyllocnistis citrella</i>
B.87 CryIA(b)	<i>Pieris</i> spp.
B.88 CryIA(b)	<i>Plutella xylostella</i>
B.89 CryIA(b)	<i>Scirpophaga</i> spp.
B.90 CryIA(b)	<i>Sesamia</i> spp.
B.91 CryIA(b)	<i>Sparganothis</i> spp.
B.92 CryIA(b)	<i>Spodoptera</i> spp.
B.93 CryIA(b)	<i>Tortrix</i> spp.
B.94 CryIA(b)	<i>Trichoplusia ni</i>
B.95 CryIA(b)	<i>Agrotis</i> spp.
B.96 CryIA(b)	<i>Anthonomus grandis</i>
B.97 CryIA(b)	<i>Curculio</i> spp.
B.98 CryIA(b)	<i>Diabrotica balteata</i>
B.99 CryIA(b)	<i>Leptinotarsa</i> spp.
B.100 CryIA(b)	<i>Lissorhoptrus</i> spp.
B.101 CryIA(b)	<i>Otiorynchus</i> spp.
B.102 CryIA(b)	<i>Aleurothrixus</i> spp.
B.103 CryIA(b)	<i>Aleyrodes</i> spp.
B.104 CryIA(b)	<i>Aonidiella</i> spp.
B.105 CryIA(b)	<i>Aphididae</i> spp.
B.106 CryIA(b)	<i>Aphis</i> spp.
B.107 CryIA(b)	<i>Bemisia tabaci</i>
B.108 CryIA(b)	<i>Empoasca</i> spp.
B.109 CryIA(b)	<i>Mycus</i> spp.
B.110 CryIA(b)	<i>Nephotettix</i> spp.
B.111 CryIA(b)	<i>Nilaparvata</i> spp.
B.112 CryIA(b)	<i>Pseudococcus</i> spp.
B.113 CryIA(b)	<i>Psylla</i> spp.
B.114 CryIA(b)	<i>Quadrastipidiotus</i> spp.
B.115 CryIA(b)	<i>Schizaphis</i> spp.
B.116 CryIA(b)	<i>Trialeurodes</i> spp.
B.117 CryIA(b)	<i>Lyriomyza</i> spp.
B.118 CryIA(b)	<i>Oscinella</i> spp.
B.119 CryIA(b)	<i>Phorbia</i> spp.
B.120 CryIA(b)	<i>Frankliniella</i> spp.
B.121 CryIA(b)	<i>Thrips</i> spp.
B.122 CryIA(b)	<i>Scirtothrips aurantii</i>
B.123 CryIA(b)	<i>Aceria</i> spp.
B.124 CryIA(b)	<i>Aculus</i> spp.
B.125 CryIA(b)	<i>Brevipalpus</i> spp.
B.126 CryIA(b)	<i>Panonychus</i> spp.
B.127 CryIA(b)	<i>Phyllocoptruta</i> spp.
B.128 CryIA(b)	<i>Tetranychus</i> spp.
B.129 CryIA(b)	<i>Heterodera</i> spp.
B.130 CryIA(b)	<i>Meloidogyne</i> spp.
B.131 CryIA(c)	<i>Adoxophyes</i> spp.
B.132 CryIA(c)	<i>Agrotis</i> spp.
B.133 CryIA(c)	<i>Alabama argillaceae</i>
B.134 CryIA(c)	<i>Anticarsia gemmatilis</i>
B.135 CryIA(c)	<i>Chilo</i> spp.
B.136 CryIA(c)	<i>Clysia ambiguella</i>
B.137 CryIA(c)	<i>Crocidolomia binotalis</i>

US 7,105,469 B2

47

TABLE B-continued

AP	Control of	
B.138	CryIA(c)	<i>Cydia</i> spp.
B.139	CryIA(c)	<i>Diparopsis castanea</i>
B.140	CryIA(c)	<i>Earias</i> spp.
B.141	CryIA(c)	<i>Ephesia</i> spp.
B.142	CryIA(c)	<i>Heliothis</i> spp.
B.143	CryIA(c)	<i>Helula undalis</i>
B.144	CryIA(c)	<i>Keiferia lycopersicella</i>
B.145	CryIA(c)	<i>Leucoptera scitella</i>
B.146	CryIA(c)	<i>Lithocollethis</i> spp.
B.147	CryIA(c)	<i>Lobesia botrana</i>
B.148	CryIA(c)	<i>Ostrinia nubilalis</i>
B.149	CryIA(c)	<i>Pandemis</i> spp.
B.150	CryIA(c)	<i>Pectinophora gossypiella</i> .
B.151	CryIA(c)	<i>Phyllocnistis citrella</i>
B.152	CryIA(c)	<i>Pieris</i> spp.
B.153	CryIA(c)	<i>Plutella xylostella</i>
B.154	CryIA(c)	<i>Scirpophaga</i> spp.
B.155	CryIA(c)	<i>Sesamia</i> spp.
B.156	CryIA(c)	<i>Sparganothis</i> spp.
B.157	CryIA(c)	<i>Spodoptera</i> spp.
B.158	CryIA(c)	<i>Tortrix</i> spp.
B.159	CryIA(c)	<i>Trichoplusia ni</i>
B.160	CryIA(c)	<i>Agriotes</i> spp.
B.161	CryIA(c)	<i>Anthonomus grandis</i>
B.162	CryIA(c)	<i>Curculio</i> spp.
B.163	CryIA(c)	<i>Diabrotica balteata</i>
B.164	CryIA(c)	<i>Leptinotarsa</i> spp.
B.165	CryIA(c)	<i>Lissorhoptrus</i> spp.
B.166	CryIA(c)	<i>Otiorynchus</i> spp.
B.167	CryIA(c)	<i>Aleurothrixus</i> spp.
B.168	CryIA(c)	<i>Aleyrodes</i> spp.
B.169	CryIA(c)	<i>Aonidiella</i> spp.
B.170	CryIA(c)	<i>Aphididae</i> spp.
B.171	CryIA(c)	<i>Aphis</i> spp.
B.172	CryIA(c)	<i>Bemisia tabaci</i>
B.173	CryIA(c)	<i>Empoasca</i> spp.
B.174	CryIA(c)	<i>Mycus</i> spp.
B.175	CryIA(c)	<i>Nephotettix</i> spp.
B.176	CryIA(c)	<i>Nilaparvata</i> spp.
B.177	CryIA(c)	<i>Pseudococcus</i> spp.
B.178	CryIA(c)	<i>Psylla</i> spp.
B.179	CryIA(c)	<i>Quadrastipitotus</i> spp.
B.180	CryIA(c)	<i>Schizaphis</i> spp.
B.181	CryIA(c)	<i>Trialeurodes</i> spp.
B.182	CryIA(c)	<i>Lyriomyza</i> spp.
B.183	CryIA(c)	<i>Oscinella</i> spp.
B.184	CryIA(c)	<i>Phorbia</i> spp.
B.185	CryIA(c)	<i>Frankliniella</i> spp.
B.186	CryIA(c)	<i>Thrips</i> spp.
B.187	CryIA(c)	<i>Scirtothrips aurantii</i>
B.188	CryIA(c)	<i>Aceria</i> spp.
B.189	CryIA(c)	<i>Aculus</i> spp.
B.190	CryIA(c)	<i>Brevipalpus</i> spp.
B.191	CryIA(c)	<i>Panonychus</i> spp.
B.192	CryIA(c)	<i>Phyllocoptruta</i> spp.
B.193	CryIA(c)	<i>Tetranychus</i> spp.
B.194	CryIA(c)	<i>Heterodera</i> spp.
B.195	CryIA(c)	<i>Meloidogyne</i> spp.
B.196	CryIIA	<i>Adoxophyes</i> spp.
B.197	CryIIA	<i>Agronis</i> spp.
B.198	CryIIA	<i>Alabama argillaceae</i>
B.199	CryIIA	<i>Anncarsia gemmatalis</i>
B.200	CryIIA	<i>Chilo</i> spp.
B.201	CryIIA	<i>Clysia ambigua</i>
B.202	CryIIA	<i>Crocidolomia binotalis</i>
B.203	CryIIA	<i>Cydia</i> spp.
B.204	CryIIA	<i>Diparopsis castanea</i>
B.205	CryIIA	<i>Earias</i> spp.
B.206	CryIIA	<i>Ephesia</i> spp.
B.207	CryIIA	<i>Heliothis</i> spp.
B.208	CryIIA	<i>Helula undalis</i>
B.209	CryIIA	<i>Keiferia lycopersicella</i>
B.210	CryIIA	<i>Leucoptera scitella</i>
B.211	CryIIA	<i>Lithocollethis</i> spp.
B.212	CryIIA	<i>Lobesia botrana</i>
B.213	CryIIA	<i>Ostrinia nubilalis</i>
B.214	CryIIA	<i>Pandemis</i> spp.

48

TABLE B-continued

AP	Control of	
B.215	CryIIA	<i>Pectinophora gossyp.</i>
B.216	CryIIA	<i>Phyllocnistis citrella</i>
B.217	CryIIA	<i>Pieris</i> spp.
B.218	CryIIA	<i>Plutella xylostella</i>
B.219	CryIIA	<i>Scirpophaga</i> spp.
B.220	CryIIA	<i>Sesamia</i> spp.
B.221	CryIIA	<i>Sparganothis</i> spp.
B.222	CryIIA	<i>Spodoptera</i> spp.
B.223	CryIIA	<i>Tortrix</i> spp.
B.224	CryIIA	<i>Trichoplusia ni</i>
B.225	CryIIA	<i>Agriotes</i> spp.
B.226	CryIIA	<i>Anthonomus grandis</i>
B.227	CryIIA	<i>Curculio</i> spp.
B.228	CryIIA	<i>Diabrotica balteata</i>
B.229	CryIIA	<i>Leptinotarsa</i> spp.
B.230	CryIIA	<i>Lissorhoptrus</i> spp.
B.231	CryIIA	<i>Otiorynchus</i> spp.
B.232	CryIIA	<i>Aleurothrixus</i> spp.
B.233	CryIIA	<i>Aleyrodes</i> spp.
B.234	CryIIA	<i>Aonidiella</i> spp.
B.235	CryIIA	<i>Aphididae</i> spp.
B.236	CryIIA	<i>Aphis</i> spp.
B.237	CryIIA	<i>Bemisia tabaci</i>
B.238	CryIIA	<i>Empoasca</i> spp.
B.239	CryIIA	<i>Mycus</i> spp.
B.240	CryIIA	<i>Nephotettix</i> spp.
B.241	CryIIA	<i>Nilaparvata</i> spp.
B.242	CryIIA	<i>Pseudococcus</i> spp.
B.243	CryIIA	<i>Psylla</i> spp.
B.244	CryIIA	<i>Quadrastipitotus</i> spp.
B.245	CryIIA	<i>Schizaphis</i> spp.
B.246	CryIIA	<i>Trialeurodes</i> spp.
B.247	CryIIA	<i>Lyriomyza</i> spp.
B.248	CryIIA	<i>Oscinella</i> spp.
B.249	CryIIA	<i>Phorbia</i> spp.
B.250	CryIIA	<i>Frankliniella</i> spp.
B.251	CryIIA	<i>Thrips</i> spp.
B.252	CryIIA	<i>Scirtothrips aurantii</i>
B.253	CryIIA	<i>Aceria</i> spp.
B.254	CryIIA	<i>Aculus</i> spp.
B.255	CryIIA	<i>Brevipalpus</i> spp.
B.256	CryIIA	<i>Panonychus</i> spp.
B.257	CryIIA	<i>Phyllocoptruta</i> spp.
B.258	CryIIA	<i>Tetranychus</i> spp.
B.259	CryIIA	<i>Heterodera</i> spp.
B.260	CryIIA	<i>Meloidogyne</i> spp.
B.261	CryIIIA	<i>Adoxophyes</i> spp.
B.262	CryIIIA	<i>Agrotis</i> spp.
B.263	CryIIIA	<i>Alabama argillaceae</i>
B.264	CryIIIA	<i>Anticarsia gemmatalis</i>
B.265	CryIIIA	<i>Chilo</i> spp.
B.266	CryIIIA	<i>Clysia ambigua</i>
B.267	CryIIIA	<i>Crocidolomia binotalis</i>
B.268	CryIIIA	<i>Cydia</i> spp.
B.269	CryIIIA	<i>Diparopsis castanea</i>
B.270	CryIIIA	<i>Earias</i> spp.
B.271	CryIIIA	<i>Ephesia</i> spp.
B.272	CryIIIA	<i>Heliothis</i> spp.
B.273	CryIIIA	<i>Helula undalis</i>
B.274	CryIIIA	<i>Keiferia lycopersicella</i>
B.275	CryIIIA	<i>Leucoptera scitella</i>
B.276	CryIIIA	<i>Lithocollethis</i> spp.
B.277	CryIIIA	<i>Lobesia botrana</i>
B.278	CryIIIA	<i>Ostrinia nubilalis</i>
B.279	CryIIIA	<i>Pandemis</i> spp.
B.280	CryIIIA	<i>Pectinophora gossyp.</i>
B.281	CryIIIA	<i>Phyllocnistis citrella</i>
B.282	CryIIIA	<i>Pieris</i> spp.
B.283	CryIIIA	<i>Plutella xylostella</i>
B.284	CryIIIA	<i>Scirpophaga</i> spp.
B.285	CryIIIA	<i>Sesamia</i> spp.
B.286	CryIIIA	<i>Sparganothis</i> spp.
B.287	CryIIIA	<i>Spodoptera</i> spp.
B.288	CryIIIA	<i>Tortrix</i> spp.
B.289	CryIIIA	<i>Trichoplusia ni</i>
B.290	CryIIIA	<i>Agriotes</i> spp.
B.291	CryIIIA	<i>Anthonomus grandis</i>

US 7,105,469 B2

49

TABLE B-continued

	AP	Control of	
B.292	CryIIIA	<i>Curculio</i> spp.	5
B.293	CryIIIA	<i>Diabrotica balteata</i>	
B.294	CryIIIA	<i>Leptinotarsa</i> spp.	
B.295	CryIIIA	<i>Lissorhynchus</i> spp.	
B.296	CryIIIA	<i>Otiorynchus</i> spp.	
B.297	CryIIIA	<i>Aleurothrixus</i> spp.	
B.298	CryIIIA	<i>Aleyrodes</i> spp.	10
B.299	CryIIIA	<i>Aonidiella</i> spp.	
B.300	CryIIIA	<i>Aphididae</i> spp.	
B.301	CryIIIA	<i>Aphis</i> spp.	
B.302	CryIIIA	<i>Bemisia tabaci</i>	
B.303	CryIIIA	<i>Empoasca</i> spp.	
B.304	CryIIIA	<i>Mycus</i> spp.	15
B.305	CryIIIA	<i>Nephotettix</i> spp.	
B.306	CryIIIA	<i>Nilaparvata</i> spp.	
B.307	CryIIIA	<i>Pseudococcus</i> spp.	
B.308	CryIIIA	<i>Psylla</i> spp.	
B.309	CryIIIA	<i>Quadraspidiotus</i> spp.	
B.310	CryIIIA	<i>Schizaphis</i> spp.	20
B.311	CryIIIA	<i>Trialeurodes</i> spp.	
B.312	CryIIIA	<i>Lynomyza</i> spp.	
B.313	CryIIIA	<i>Oscinella</i> spp.	
B.314	CryIIIA	<i>Phorbia</i> spp.	
B.315	CryIIIA	<i>Frankliniella</i> spp.	
B.316	CryIIIA	<i>Thrips</i> spp.	
B.317	CryIIIA	<i>Scirtothrips aurantii</i>	25
B.318	CryIIIA	<i>Aceria</i> spp.	
B.319	CryIIIA	<i>Aculus</i> spp.	
B.320	CryIIIA	<i>Brevipalpus</i> spp.	
B.321	CryIIIA	<i>Panonychus</i> spp.	
B.322	CryIIIA	<i>Phyllocoptruta</i> spp.	
B.323	CryIIIA	<i>Tetranychus</i> spp.	30
B.324	CryIIIA	<i>Heterodera</i> spp.	
B.325	CryIIIA	<i>Meloidogyne</i> spp.	
B.326	CryIIIB2	<i>Adoxophyes</i> spp.	
B.327	CryIIIB2	<i>Agrotis</i> spp.	
B.328	CryIIIB2	<i>Alabama argillaceae</i>	
B.329	CryIIIB2	<i>Anticarsia gemmatilis</i>	35
B.330	CryIIIB2	<i>Chilo</i> spp.	
B.331	CryIIIB2	<i>Clysia ambiguella</i>	
B.332	CryIIIB2	<i>Crocidolomia binotalis</i>	
B.333	CryIIIB2	<i>Cydia</i> spp.	
B.334	CryIIIB2	<i>Diparopsis castanea</i>	
B.335	CryIIIB2	<i>Earias</i> spp.	40
B.336	CryIIIB2	<i>Ephestia</i> spp.	
B.337	CryIIIB2	<i>Heliothis</i> spp.	
B.338	CryIIIB2	<i>Hellula undalis</i>	
B.339	CryIIIB2	<i>Keiferia lycopersicella</i>	
B.340	CryIIIB2	<i>Leucophaea scitella</i>	
B.341	CryIIIB2	<i>Lithocolletis</i> spp.	45
B.342	CryIIIB2	<i>Lobesia botrana</i>	
B.343	CryIIIB2	<i>Ostrinia nubilalis</i>	
B.344	CryIIIB2	<i>Pandemis</i> spp.	
B.345	CryIIIB2	<i>Pectinophora gossyp.</i>	
B.346	CryIIIB2	<i>Phyllocnistis citrella</i>	
B.347	CryIIIB2	<i>Pieris</i> spp.	
B.348	CryIIIB2	<i>Plutella xylostella</i>	50
B.349	CryIIIB2	<i>Scirpophaga</i> spp.	
B.350	CryIIIB2	<i>Sesamia</i> spp.	
B.351	CryIIIB2	<i>Sparganothis</i> spp.	
B.352	CryIIIB2	<i>Spodoptera</i> spp.	
B.353	CryIIIB2	<i>Tortrix</i> spp.	
B.354	CryIIIB2	<i>Trichoplusia ni</i>	55
B.355	CryIIIB2	<i>Agrotis</i> spp.	
B.356	CryIIIB2	<i>Anthonomus grandis</i>	
B.357	CryIIIB2	<i>Curculio</i> spp.	
B.358	CryIIIB2	<i>Diabrotica balteata</i>	
B.359	CryIIIB2	<i>Leptinotarsa</i> spp.	
B.360	CryIIIB2	<i>Lissorhynchus</i> spp.	60
B.361	CryIIIB2	<i>Otiorynchus</i> spp.	
B.362	CryIIIB2	<i>Aleurothrixus</i> spp.	
B.363	CryIIIB2	<i>Aleyrodes</i> spp.	
B.364	CryIIIB2	<i>Aonidiella</i> spp.	
B.365	CryIIIB2	<i>Aphididae</i> spp.	
B.366	CryIIIB2	<i>Aphis</i> spp.	
B.367	CryIIIB2	<i>Bemisia tabaci</i>	65
B.368	CryIIIB2	<i>Empoasca</i> spp.	

50

TABLE B-continued

	AP	Control of
B.369	CryIIIB2	<i>Mycus</i> spp.
B.370	CryIIIB2	<i>Nephotettix</i> spp.
B.371	CryIIIB2	<i>Nilaparvata</i> spp.
B.372	CryIIIB2	<i>Pseudococcus</i> spp.
B.373	CryIIIB2	<i>Psylla</i> spp.
B.374	CryIIIB2	<i>Quadraspidiotus</i> spp.
B.375	CryIIIB2	<i>Schizaphis</i> spp.
B.376	CryIIIB2	<i>Trialeurodes</i> spp.
B.377	CryIIIB2	<i>Lynomyza</i> spp.
B.378	CryIIIB2	<i>Oscinella</i> spp.
B.379	CryIIIB2	<i>Phorbia</i> spp.
B.380	CryIIIB2	<i>Frankliniella</i> spp.
B.381	CryIIIB2	<i>Thrips</i> spp.
B.382	CryIIIB2	<i>Scirtothrips aurantii</i>
B.383	CryIIIB2	<i>Aceria</i> spp.
B.384	CryIIIB2	<i>Aculus</i> spp.
B.385	CryIIIB2	<i>Brevipalpus</i> spp.
B.386	CryIIIB2	<i>Panonychus</i> spp.
B.387	CryIIIB2	<i>Phyllocoptruta</i> spp.
B.388	CryIIIB2	<i>Tetranychus</i> spp.
B.389	CryIIIB2	<i>Heterodera</i> spp.
B.390	CryIIIB2	<i>Meloidogyne</i> spp.
B.391	CyTA	<i>Adoxophyes</i> spp.
B.392	CyTA	<i>Agrotis</i> spp.
B.393	CyTA	<i>Alabama argillaceae</i>
B.394	CyTA	<i>Anticarsia gemmatilis</i>
B.395	CyTA	<i>Chilo</i> spp.
B.396	CyTA	<i>Clysia ambiguella</i>
B.397	CyTA	<i>Crocidolomia binotalis</i>
B.398	CyTA	<i>Cydia</i> spp.
B.399	CyTA	<i>Diparopsis castanea</i>
B.400	CyTA	<i>Earias</i> spp.
B.401	CyTA	<i>Ephestia</i> spp.
B.402	CyTA	<i>Heliothis</i> spp.
B.403	CyTA	<i>Hellula undalis</i>
B.404	CyTA	<i>Keiferia lycopersicella</i>
B.405	CyTA	<i>Leucophaea scitella</i>
B.406	CyTA	<i>Lithocolletis</i> spp.
B.407	CyTA	<i>Lobesia botrana</i>
B.408	CyTA	<i>Ostrinia nubilalis</i>
B.409	CyTA	<i>Pandemis</i> spp.
B.410	CyTA	<i>Pectinophora gossyp.</i>
B.411	CyTA	<i>Phyllocnistis citrella</i>
B.412	CyTA	<i>Pieris</i> spp.
B.413	CyTA	<i>Plutella xylostella</i>
B.414	CyTA	<i>Scirpophaga</i> spp.
B.415	CyTA	<i>Sesamia</i> spp.
B.416	CyTA	<i>Sparganothis</i> spp.
B.417	CyTA	<i>Spodoptera</i> spp.
B.418	CyTA	<i>Tortrix</i> spp.
B.419	CyTA	<i>Trichoplusia ni</i>
B.420	CyTA	<i>Agrotis</i> spp.
B.421	CyTA	<i>Anthonomus grandis</i>
B.422	CyTA	<i>Curculio</i> spp.
B.423	CyTA	<i>Diabrotica balteata</i>
B.424	CyTA	<i>Leptinotarsa</i> spp.
B.425	CyTA	<i>Lissorhynchus</i> spp.
B.426	CyTA	<i>Otiorynchus</i> spp.
B.427	CyTA	<i>Aleurothrixus</i> spp.
B.428	CyTA	<i>Aleyrodes</i> spp.
B.429	CyTA	<i>Aonidiella</i> spp.
B.430	CyTA	<i>Aphididae</i> spp.
B.431	CyTA	<i>Aphis</i> spp.
B.432	CyTA	<i>Bemisia tabaci</i>
B.433	CyTA	<i>Empoasca</i> spp.
B.434	CyTA	<i>Mycus</i> spp.
B.435	CyTA	<i>Nephotettix</i> spp.
B.436	CyTA	<i>Nilaparvata</i> spp.
B.437	CyTA	<i>Pseudococcus</i> spp.
B.438	CyTA	<i>Psylla</i> spp.
B.439	CyTA	<i>Quadraspidiotus</i> spp.
B.440	CyTA	<i>Schizaphis</i> spp.
B.441	CyTA	<i>Trialeurodes</i> spp.
B.442	CyTA	<i>Lynomyza</i> spp.
B.443	CyTA	<i>Oscinella</i> spp.
B.444	CyTA	<i>Phorbia</i> spp.
B.445	CyTA	<i>Frankliniella</i> spp.

US 7,105,469 B2

51

TABLE B-continued

	AP	Control of	
B.446	CyA	<i>Thrips</i> spp.	5
B.447	CyA	<i>Scirtothrips aurantii</i>	
B.448	CyA	<i>Aceria</i> spp.	
B.449	CyA	<i>Aculus</i> spp.	
B.450	CyA	<i>Brevipalpus</i> spp.	
B.451	CyA	<i>Panonychus</i> spp.	
B.452	CyA	<i>Phyllocoptruta</i> spp.	10
B.453	CyA	<i>Tetranychus</i> spp.	
B.454	CyA	<i>Heterodera</i> spp.	
B.455	CyA	<i>Meloidogyne</i> spp.	
B.456	VIP3	<i>Adoxophyes</i> spp.	
B.457	VIP3	<i>Agrotis</i> spp.	
B.458	VIP3	<i>Alabama argillaceae</i>	15
B.459	VIP3	<i>Anticarsia gemmatilis</i>	
B.460	VIP3	<i>Chilo</i> spp.	
B.461	VIP3	<i>Clysia ambiguella</i>	
B.462	VIP3	<i>Crocidolomia binotalis</i>	
B.463	VIP3	<i>Cydia</i> spp.	
B.464	VIP3	<i>Diparopsis castanea</i>	20
B.465	VIP3	<i>Earias</i> spp.	
B.466	VIP3	<i>Ephestia</i> spp.	
B.467	VIP3	<i>Heliothis</i> spp.	
B.468	VIP3	<i>Hellula undalis</i>	
B.469	VIP3	<i>Keiferia lycopersicella</i>	
B.470	VIP3	<i>Leucophaea scitella</i>	25
B.471	VIP3	<i>Lithocollethis</i> spp.	
B.472	VIP3	<i>Lobesia botrana</i>	
B.473	VIP3	<i>Ostrinia nubilalis</i>	
B.474	VIP3	<i>Pandemis</i> spp.	
B.475	VIP3	<i>Pectinophora gossypi</i>	
B.476	VIP3	<i>Phyllocnistis citrella</i>	
B.477	VIP3	<i>Pieris</i> spp.	30
B.478	VIP3	<i>Plutella xylostella</i>	
B.479	VIP3	<i>Scirpophaga</i> spp.	
B.480	VIP3	<i>Sesamia</i> spp.	
B.481	VIP3	<i>Sparganothis</i> spp.	
B.482	VIP3	<i>Spodoptera</i> spp.	
B.483	VIP3	<i>Tortrix</i> spp.	35
B.484	VIP3	<i>Trichoplusia ni</i>	
B.485	VIP3	<i>Agrotis</i> spp.	
B.486	VIP3	<i>Anthonomus grandis</i>	
B.487	VIP3	<i>Curculio</i> spp.	
B.488	VIP3	<i>Diabrotica balteata</i>	
B.489	VIP3	<i>Leptinotarsa</i> spp.	40
B.490	VIP3	<i>Lissorhoptrus</i> spp.	
B.491	VIP3	<i>Otiorynchus</i> spp.	
B.492	VIP3	<i>Aleurothrixus</i> spp.	
B.493	VIP3	<i>Aleyrodes</i> spp.	
B.494	VIP3	<i>Aonidiella</i> spp.	
B.495	VIP3	<i>Aphididae</i> spp.	45
B.496	VIP3	<i>Aphis</i> spp.	
B.497	VIP3	<i>Bemisia tabaci</i>	
B.498	VIP3	<i>Empoasca</i> spp.	
B.499	VIP3	<i>Mycus</i> spp.	
B.500	VIP3	<i>Nephotettix</i> spp.	
B.501	VIP3	<i>Nilaparvata</i> spp.	
B.502	VIP3	<i>Pseudococcus</i> spp.	50
B.503	VIP3	<i>Psylla</i> spp.	
B.504	VIP3	<i>Quadrastipidiotus</i> spp.	
B.505	VIP3	<i>Schizaphis</i> spp.	
B.506	VIP3	<i>Trialeurodes</i> spp.	
B.507	VIP3	<i>Lyriomyza</i> spp.	
B.508	VIP3	<i>Oscinella</i> spp.	55
B.509	VIP3	<i>Phorbia</i> spp.	
B.510	VIP3	<i>Frankliniella</i> spp.	
B.511	VIP3	<i>Thrips</i> spp.	
B.512	VIP3	<i>Scirtothrips aurantii</i>	
B.513	VIP3	<i>Aceria</i> spp.	
B.514	VIP3	<i>Aculus</i> spp.	60
B.515	VIP3	<i>Brevipalpus</i> spp.	
B.516	VIP3	<i>Panonychus</i> spp.	
B.517	VIP3	<i>Phyllocoptruta</i> spp.	
B.518	VIP3	<i>Tetranychus</i> spp.	
B.519	VIP3	<i>Heterodera</i> spp.	
B.520	VIP3	<i>Meloidogyne</i> spp.	
B.521	GL	<i>Adoxophyes</i> spp.	65
B.522	GL	<i>Agrotis</i> spp.	

52

TABLE B-continued

	AP	Control of
B.523	GL	<i>Alabama argillaceae</i>
B.524	GL	<i>Anticarsia gemmatilis</i>
B.525	GL	<i>Chilo</i> spp.
B.526	GL	<i>Clysia ambiguella</i>
B.527	GL	<i>Crocidolomia binotalis</i>
B.528	GL	<i>Cydia</i> spp.
B.529	GL	<i>Diparopsis castanea</i>
B.530	GL	<i>Earias</i> spp.
B.531	GL	<i>Ephestia</i> spp.
B.532	GL	<i>Heliothis</i> spp.
B.533	GL	<i>Hellula undalis</i>
B.534	GL	<i>Keiferia lycopersicella</i>
B.535	GL	<i>Leucophaea scitella</i>
B.536	GL	<i>Lithocollethis</i> spp.
B.537	GL	<i>Lobesia botrana</i>
B.538	GL	<i>Ostrinia nubilalis</i>
B.539	GL	<i>Pandemis</i> spp.
B.540	GL	<i>Pectinophora gossypi</i>
B.541	GL	<i>Phyllocnistis citrella</i>
B.542	GL	<i>Pieris</i> spp.
B.543	GL	<i>Plutella xylostella</i>
B.544	GL	<i>Scirpophaga</i> spp.
B.545	GL	<i>Sesamia</i> spp.
B.546	GL	<i>Sparganothis</i> spp.
B.547	GL	<i>Spodoptera</i> spp.
B.548	GL	<i>Tortrix</i> spp.
B.549	GL	<i>Trichoplusia ni</i>
B.550	GL	<i>Agrotis</i> spp.
B.551	GL	<i>Anthonomus grandis</i>
B.552	GL	<i>Curculio</i> spp.
B.553	GL	<i>Diabrotica balteata</i>
B.554	GL	<i>Leptinotarsa</i> spp.
B.555	GL	<i>Lissorhoptrus</i> spp.
B.556	GL	<i>Otiorynchus</i> spp.
B.557	GL	<i>Aleurothrixus</i> spp.
B.558	GL	<i>Aleyrodes</i> spp.
B.559	GL	<i>Aonidiella</i> spp.
B.560	GL	<i>Aphididae</i> spp.
B.561	GL	<i>Aphis</i> spp.
B.562	GL	<i>Bemisia tabaci</i>
B.563	GL	<i>Empoasca</i> spp.
B.564	GL	<i>Mycus</i> spp.
B.565	GL	<i>Nephotettix</i> spp.
B.566	GL	<i>Nilaparvata</i> spp.
B.567	GL	<i>Pseudococcus</i> spp.
B.568	GL	<i>Psylla</i> spp.
B.569	GL	<i>Quadrastipidiotus</i> spp.
B.570	GL	<i>Schizaphis</i> spp.
B.571	GL	<i>Trialeurodes</i> spp.
B.572	GL	<i>Lyriomyza</i> spp.
B.573	GL	<i>Oscinella</i> spp.
B.574	GL	<i>Phorbia</i> spp.
B.575	GL	<i>Frankliniella</i> spp.
B.576	GL	<i>Thrips</i> spp.
B.577	GL	<i>Scirtothrips aurantii</i>
B.578	GL	<i>Aceria</i> spp.
B.579	GL	<i>Aculus</i> spp.
B.580	GL	<i>Brevipalpus</i> spp.
B.581	GL	<i>Panonychus</i> spp.
B.582	GL	<i>Phyllocoptruta</i> spp.
B.583	GL	<i>Tetranychus</i> spp.
B.584	GL	<i>Heterodera</i> spp.
B.585	GL	<i>Meloidogyne</i> spp.
B.586	PL	<i>Adoxophyes</i> spp.
B.587	PL	<i>Agrotis</i> spp.
B.588	PL	<i>Alabama argillaceae</i>
B.589	PL	<i>Anticarsia gemmatilis</i>
B.590	PL	<i>Chilo</i> spp.
B.591	PL	<i>Clysia ambiguella</i>
B.592	PL	<i>Crocidolomia binotalis</i>
B.593	PL	<i>Cydia</i> spp.
B.594	PL	<i>Diparopsis castanea</i>
B.595	PL	<i>Earias</i> spp.
B.596	PL	<i>Ephestia</i> spp.
B.597	PL	<i>Heliothis</i> spp.
B.598	PL	<i>Hellula undalis</i>
B.599	PL	<i>Keiferia lycopersicella</i>

US 7,105,469 B2

53

TABLE B-continued

	AP	Control of
B.600	PL	<i>Leucoptera scitella</i>
B.601	PL	<i>Lithocolletis</i> spp.
B.602	PL	<i>Lobesia botrana</i>
B.603	PL	<i>Ostrinia nubilalis</i>
B.604	PL	<i>Pandemis</i> spp.
B.605	PL	<i>Pectinophora gossyp.</i>
B.606	PL	<i>Phyllocnistis citrella</i>
B.607	PL	<i>Pieris</i> spp.
B.608	PL	<i>Plutella xylostella</i>
B.609	PL	<i>Scirpophaga</i> spp.
B.610	PL	<i>Sesamia</i> spp.
B.611	PL	<i>Sparganothis</i> spp.
B.612	PL	<i>Spodoptera</i> spp.
B.613	PL	<i>Tortrix</i> spp.
B.614	PL	<i>Trichoplusia ni</i>
B.615	PL	<i>Agrotis</i> spp.
B.616	PL	<i>Anthonomus grandis</i>
B.617	PL	<i>Curculio</i> spp.
B.618	PL	<i>Diabrotica balteata</i>
B.619	PL	<i>Leptinotarsa</i> spp.
B.620	PL	<i>Lissorhoptrus</i> spp.
B.621	PL	<i>Otiorthynchus</i> spp.
B.622	PL	<i>Aleurothrixus</i> spp.
B.623	PL	<i>Aleyrodes</i> spp.
B.624	PL	<i>Aonidiella</i> spp.
B.625	PL	<i>Aphididae</i> spp.
B.626	PL	<i>Aphis</i> spp.
B.627	PL	<i>Bemisia tabaci</i>
B.628	PL	<i>Empoasca</i> spp.
B.629	PL	<i>Mycus</i> spp.
B.630	PL	<i>Nephotettix</i> spp.
B.631	PL	<i>Nilaparvata</i> spp.
B.632	PL	<i>Pseudococcus</i> spp.
B.633	PL	<i>Psylla</i> spp.
B.634	PL	<i>Quadrastipidiotus</i> spp.
B.635	PL	<i>Schizaphis</i> spp.
B.636	PL	<i>Trialeurodes</i> spp.
B.637	PL	<i>Lyriomyza</i> spp.
B.638	PL	<i>Oscinella</i> spp.
B.639	PL	<i>Phorbia</i> spp.
B.640	PL	<i>Frankliniella</i> spp.
B.641	PL	<i>Thrips</i> spp.
B.642	PL	<i>Scirtothrips aurantii</i>
B.643	PL	<i>Aceria</i> spp.
B.644	PL	<i>Aculus</i> spp.
B.645	PL	<i>Brevipalpus</i> spp.
B.646	PL	<i>Panonychus</i> spp.
B.647	PL	<i>Phyllocoptura</i> spp.
B.648	PL	<i>Tetranychus</i> spp.
B.649	PL	<i>Heterodera</i> spp.
B.650	PL	<i>Meloidogyne</i> spp.
B.651	XN	<i>Adoxophyes</i> spp.
B.652	XN	<i>Agrotis</i> spp.
B.653	XN	<i>Alabama argillaceae</i>
B.654	XN	<i>Anticarsia gemmatilis</i>
B.655	XN	<i>Chilo</i> spp.
B.656	XN	<i>Clysia ambiguella</i>
B.657	XN	<i>Crocidolomia binotalis</i>
B.658	XN	<i>Cydia</i> spp.
B.659	XN	<i>Diparopsis castanea</i>
B.660	XN	<i>Earias</i> spp.
B.661	XN	<i>Ephesia</i> spp.
B.662	XN	<i>Heliothis</i> spp.
B.663	XN	<i>Helicula undalis</i>
B.664	XN	<i>Keiferia lycopersicella</i>
B.665	XN	<i>Leucoptera scitella</i>
B.666	XN	<i>Lithocolletis</i> spp.
B.667	XN	<i>Lobesia botrana</i>
B.668	XN	<i>Ostrinia nubilalis</i>
B.669	XN	<i>Pandemis</i> spp.
B.670	XN	<i>Pectinophora gossyp.</i>
B.671	XN	<i>Phyllocnistis citrella</i>
B.672	XN	<i>Pieris</i> spp.
B.673	XN	<i>Plutella xylostella</i>
B.674	XN	<i>Scirpophaga</i> spp.
B.675	XN	<i>Sesamia</i> spp.
B.676	XN	<i>Sparganothis</i> spp.

54

TABLE B-continued

	AP	Control of
B.677	XN	<i>Spodoptera</i> spp.
B.678	XN	<i>Tortrix</i> spp.
B.679	XN	<i>Trichoplusia ni</i>
B.680	XN	<i>Agrotis</i> spp.
B.681	XN	<i>Anthonomus grandis</i>
B.682	XN	<i>Curculio</i> spp.
B.683	XN	<i>Diabrotica balteata</i>
B.684	XN	<i>Leptinotarsa</i> spp.
B.685	XN	<i>Lissorhoptrus</i> spp.
B.686	XN	<i>Otiorthynchus</i> spp.
B.687	XN	<i>Aleurothrixus</i> spp.
B.688	XN	<i>Aleyrodes</i> spp.
B.689	XN	<i>Aonidiella</i> spp.
B.690	XN	<i>Aphididae</i> spp.
B.691	XN	<i>Aphis</i> spp.
B.692	XN	<i>Bemisia tabaci</i>
B.693	XN	<i>Empoasca</i> spp.
B.694	XN	<i>Mycus</i> spp.
B.695	XN	<i>Nephotettix</i> spp.
B.696	XN	<i>Nilaparvata</i> spp.
B.697	XN	<i>Pseudococcus</i> spp.
B.698	XN	<i>Psylla</i> spp.
B.699	XN	<i>Quadrastipidiotus</i> spp.
B.700	XN	<i>Schizaphis</i> spp.
B.701	XN	<i>Trialeurodes</i> spp.
B.702	XN	<i>Lyriomyza</i> spp.
B.703	XN	<i>Oscinella</i> spp.
B.704	XN	<i>Phorbia</i> spp.
B.705	XN	<i>Frankliniella</i> spp.
B.706	XN	<i>Thrips</i> spp.
B.707	XN	<i>Scirtothrips aurantii</i>
B.708	XN	<i>Aceria</i> spp.
B.709	XN	<i>Aculus</i> spp.
B.710	XN	<i>Brevipalpus</i> spp.
B.711	XN	<i>Panonychus</i> spp.
B.712	XN	<i>Phyllocoptura</i> spp.
B.713	XN	<i>Tetranychus</i> spp.
B.714	XN	<i>Heterodera</i> spp.
B.715	XN	<i>Meloidogyne</i> spp.
B.716	Plnh.	<i>Adoxophyes</i> spp.
B.717	Plnh.	<i>Agrotis</i> spp.
B.718	Plnh.	<i>Alabama argillaceae</i>
B.719	Plnh.	<i>Anticarsia gemmatilis</i>
B.720	Plnh.	<i>Chilo</i> spp.
B.721	Plnh.	<i>Clysia ambiguella</i>
B.722	Plnh.	<i>Crocidolomia binotalis</i>
B.723	Plnh.	<i>Cydia</i> spp.
B.724	Plnh.	<i>Diparopsis castanea</i>
B.725	Plnh.	<i>Earias</i> spp.
B.726	Plnh.	<i>Ephesia</i> spp.
B.727	Plnh.	<i>Heliothis</i> spp.
B.728	Plnh.	<i>Helicula undalis</i>
B.729	Plnh.	<i>Keiferia lycopersicella</i>
B.730	Plnh.	<i>Leucoptera scitella</i>
B.731	Plnh.	<i>Lithocolletis</i> spp.
B.732	Plnh.	<i>Lobesia botrana</i>
B.733	Plnh.	<i>Ostrinia nubilalis</i>
B.734	Plnh.	<i>Pandemis</i> spp.
B.735	Plnh.	<i>Pectinophora gossyp.</i>
B.736	Plnh.	<i>Phyllocnistis citrella</i>
B.737	Plnh.	<i>Pieris</i> spp.
B.738	Plnh.	<i>Plutella xylostella</i>
B.739	Plnh.	<i>Scirpophaga</i> spp.
B.740	Plnh.	<i>Sesamia</i> spp.
B.741	Plnh.	<i>Sparganothis</i> spp.
B.742	Plnh.	<i>Spodoptera</i> spp.
B.743	Plnh.	<i>Tortrix</i> spp.
B.744	Plnh.	<i>Trichoplusia ni</i>
B.745	Plnh.	<i>Agrotis</i> spp.
B.746	Plnh.	<i>Anthonomus grandis</i>
B.747	Plnh.	<i>Curculio</i> spp.
B.748	Plnh.	<i>Diabrotica balteata</i>
B.749	Plnh.	<i>Leptinotarsa</i> spp.
B.750	Plnh.	<i>Lissorhoptrus</i> spp.
B.751	Plnh.	<i>Otiorthynchus</i> spp.
B.752	Plnh.	<i>Aleurothrixus</i> spp.
B.753	Plnh.	<i>Aleyrodes</i> spp.

US 7,105,469 B2

55

TABLE B-continued

	AP	Control of	
B.754	Plnh.	<i>Aonidiella</i> spp.	5
B.755	Plnh.	<i>Aphididae</i> spp.	
B.756	Plnh.	<i>Aphis</i> spp.	
B.757	Plnh.	<i>Bemisia tabaci</i>	
B.758	Plnh.	<i>Empoasca</i> spp.	
B.759	Plnh.	<i>Mycus</i> spp.	
B.760	Plnh.	<i>Nephotettix</i> spp.	10
B.761	Plnh.	<i>Nilaparvata</i> spp.	
B.762	Plnh.	<i>Pseudococcus</i> spp.	
B.763	Plnh.	<i>Psylla</i> spp.	
B.764	Plnh.	<i>Quadraspidiotus</i> spp.	
B.765	Plnh.	<i>Schizaphis</i> spp.	
B.766	Plnh.	<i>Trialeurodes</i> spp.	15
B.767	Plnh.	<i>Lyriomyza</i> spp.	
B.768	Plnh.	<i>Oscinella</i> spp.	
B.769	Plnh.	<i>Phorbia</i> spp.	
B.770	Plnh.	<i>Frankliniella</i> spp.	
B.771	Plnh.	<i>Thrips</i> spp.	
B.772	Plnh.	<i>Scirtothrips aurantii</i>	20
B.773	Plnh.	<i>Aceria</i> spp.	
B.774	Plnh.	<i>Aculus</i> spp.	
B.775	Plnh.	<i>Brevipalpus</i> spp.	
B.776	Plnh.	<i>Panonychus</i> spp.	
B.777	Plnh.	<i>Phyllocoptruta</i> spp.	
B.778	Plnh.	<i>Tetranychus</i> spp.	
B.779	Plnh.	<i>Heterodera</i> spp.	25
B.780	Plnh.	<i>Meloidogyne</i> spp.	
B.781	PLec.	<i>Adoxophyes</i> spp.	
B.782	PLec.	<i>Agrotis</i> spp.	
B.783	PLec.	<i>Alabama argillaceae</i>	
B.784	PLec.	<i>Anticarsia gemmatilis</i>	
B.785	PLec.	<i>Chilo</i> spp.	30
B.786	PLec.	<i>Clysia ambiguella</i>	
B.787	PLec.	<i>Crocidolomia binotalis</i>	
B.788	PLec.	<i>Cydia</i> spp.	
B.789	PLec.	<i>Diparopsis castanea</i>	
B.790	PLec.	<i>Earias</i> spp.	
B.791	PLec.	<i>Ephestia</i> spp.	35
B.792	PLec.	<i>Heliothis</i> spp.	
B.793	PLec.	<i>Hellula undalis</i>	
B.794	PLec.	<i>Keiferia lycopersicella</i>	
B.795	PLec.	<i>Leucopetera scitella</i>	
B.796	PLec.	<i>Lithocolletis</i> spp.	
B.797	PLec.	<i>Lobesia botrana</i>	40
B.798	PLec.	<i>Ostrinia nubilalis</i>	
B.799	PLec.	<i>Pandemis</i> spp.	
B.800	PLec.	<i>Pectinophora gossyp.</i>	
B.801	PLec.	<i>Phyllocnistis citrella</i>	
B.802	PLec.	<i>Pieris</i> spp.	
B.803	PLec.	<i>Plutella xylostella</i>	45
B.804	PLec.	<i>Scirpophaga</i> spp.	
B.805	PLec.	<i>Sesamia</i> spp.	
B.806	PLec.	<i>Sparganothis</i> spp.	
B.807	PLec.	<i>Spodoptera</i> spp.	
B.808	PLec.	<i>Tortrix</i> spp.	
B.809	PLec.	<i>Trichoplusia ni</i>	50
B.810	PLec.	<i>Agrotis</i> spp.	
B.811	PLec.	<i>Anthonomus grandis</i>	
B.812	PLec.	<i>Curculio</i> spp.	
B.813	PLec.	<i>Diabrotica balteata</i>	
B.814	PLec.	<i>Leptinotarsa</i> spp.	
B.815	PLec.	<i>Lissorhynchus</i> spp.	
B.816	PLec.	<i>Otiorynchus</i> spp.	55
B.817	PLec.	<i>Aleurothrixus</i> spp.	
B.818	PLec.	<i>Aleyrodes</i> spp.	
B.819	PLec.	<i>Aonidiella</i> spp.	
B.820	PLec.	<i>Aphididae</i> spp.	
B.821	PLec.	<i>Aphis</i> spp.	
B.822	PLec.	<i>Bemisia tabaci</i>	60
B.823	PLec.	<i>Empoasca</i> spp.	
B.824	PLec.	<i>Mycus</i> spp.	
B.825	PLec.	<i>Nephotettix</i> spp.	
B.826	PLec.	<i>Nilaparvata</i> spp.	
B.827	PLec.	<i>Pseudococcus</i> spp.	
B.828	PLec.	<i>Psylla</i> spp.	
B.829	PLec.	<i>Quadraspidiotus</i> spp.	65
B.830	PLec.	<i>Schizaphis</i> spp.	

56

TABLE B-continued

	AP	Control of
B.831	PLec.	<i>Trialeurodes</i> spp.
B.832	PLec.	<i>Lyriomyza</i> spp.
B.833	PLec.	<i>Oscinella</i> spp.
B.834	PLec.	<i>Phorbia</i> spp.
B.835	PLec.	<i>Frankliniella</i> spp.
B.836	PLec.	<i>Thrips</i> spp.
B.837	PLec.	<i>Scirtothrips aurantii</i>
B.838	PLec.	<i>Aceria</i> spp.
B.839	PLec.	<i>Aculus</i> spp.
B.840	PLec.	<i>Brevipalpus</i> spp.
B.841	PLec.	<i>Panonychus</i> spp.
B.842	PLec.	<i>Phyllocoptruta</i> spp.
B.843	PLec.	<i>Tetranychus</i> spp.
B.844	PLec.	<i>Heterodera</i> spp.
B.845	PLec.	<i>Meloidogyne</i> spp.
B.846	Aggl.	<i>Adoxophyes</i> spp.
B.847	Aggl.	<i>Agrotis</i> spp.
B.848	Aggl.	<i>Alabama argillaceae</i>
B.849	Aggl.	<i>Anticarsia gemmatilis</i>
B.850	Aggl.	<i>Chilo</i> spp.
B.851	Aggl.	<i>Clysia ambiguella</i>
B.852	Aggl.	<i>Crocidolomia binotalis</i>
B.853	Aggl.	<i>Cydia</i> spp.
B.854	Aggl.	<i>Diparopsis castanea</i>
B.855	Aggl.	<i>Earias</i> spp.
B.856	Aggl.	<i>Ephestia</i> spp.
B.857	Aggl.	<i>Heliothis</i> spp.
B.858	Aggl.	<i>Hellula undalis</i>
B.859	Aggl.	<i>Keiferia lycopersicella</i>
B.860	Aggl.	<i>Leucopetera scitella</i>
B.861	Aggl.	<i>Lithocolletis</i> spp.
B.862	Aggl.	<i>Lobesia botrana</i>
B.863	Aggl.	<i>Ostrinia nubilalis</i>
B.864	Aggl.	<i>Pandemis</i> spp.
B.865	Aggl.	<i>Pectinophora gossyp.</i>
B.866	Aggl.	<i>Phyllocnistis citrella</i>
B.867	Aggl.	<i>Pieris</i> spp.
B.868	Aggl.	<i>Plutella xylostella</i>
B.869	Aggl.	<i>Scirpophaga</i> spp.
B.870	Aggl.	<i>Sesamia</i> spp.
B.871	Aggl.	<i>Sparganothis</i> spp.
B.872	Aggl.	<i>Spodoptera</i> spp.
B.873	Aggl.	<i>Tortrix</i> spp.
B.874	Aggl.	<i>Trichoplusia ni</i>
B.875	Aggl.	<i>Agrotis</i> spp.
B.876	Aggl.	<i>Anthonomus grandis</i>
B.877	Aggl.	<i>Curculio</i> spp.
B.878	Aggl.	<i>Diabrotica balteata</i>
B.879	Aggl.	<i>Leptinotarsa</i> spp.
B.880	Aggl.	<i>Lissorhynchus</i> spp.
B.881	Aggl.	<i>Otiorynchus</i> spp.
B.882	Aggl.	<i>Aleurothrixus</i> spp.
B.883	Aggl.	<i>Aleyrodes</i> spp.
B.884	Aggl.	<i>Aonidiella</i> spp.
B.885	Aggl.	<i>Aphididae</i> spp.
B.886	Aggl.	<i>Aphis</i> spp.
B.887	Aggl.	<i>Bemisia tabaci</i>
B.888	Aggl.	<i>Empoasca</i> spp.
B.889	Aggl.	<i>Mycus</i> spp.
B.890	Aggl.	<i>Nephotettix</i> spp.
B.891	Aggl.	<i>Nilaparvata</i> spp.
B.892	Aggl.	<i>Pseudococcus</i> spp.
B.893	Aggl.	<i>Psylla</i> spp.
B.894	Aggl.	<i>Quadraspidiotus</i> spp.
B.895	Aggl.	<i>Schizaphis</i> spp.
B.896	Aggl.	<i>Trialeurodes</i> spp.
B.897	Aggl.	<i>Lynomyza</i> spp.
B.898	Aggl.	<i>Oscinella</i> spp.
B.899	Aggl.	<i>Phorbia</i> spp.
B.900	Aggl.	<i>Frankliniella</i> spp.
B.901	Aggl.	<i>Thrips</i> spp.
B.902	Aggl.	<i>Scirtothrips aurantii</i>
B.903	Aggl.	<i>Aceria</i> spp.
B.904	Aggl.	<i>Aculus</i> spp.
B.905	Aggl.	<i>Brevipalpus</i> spp.
B.906	Aggl.	<i>Panonychus</i> spp.
B.907	Aggl.	<i>Phyllocoptruta</i> spp.

US 7,105,469 B2

57

TABLE B-continued

	AP	Control of	
B.908	Aggl.	<i>Tetranychus</i> spp.	5
B.909	Aggl.	<i>Heterodera</i> spp.	
B.910	Aggl.	<i>Meloidogyne</i> spp.	
B.911	CO	<i>Adoxophyes</i> spp.	
B.912	CO	<i>Agrotis</i> spp.	
B.913	CO	<i>Alabama argillaceae</i>	
B.914	CO	<i>Anticarsia gemmatilis</i>	10
B.915	CO	<i>Chilo</i> spp.	
B.916	CO	<i>Clysia ambiguella</i>	
B.917	CO	<i>Crocidolomia binotalis</i>	
B.918	CO	<i>Cydia</i> spp.	
B.919	CO	<i>Diparopsis castanea</i>	
B.920	CO	<i>Earias</i> spp.	15
B.921	CO	<i>Ephesia</i> spp.	
B.922	CO	<i>Heliothis</i> spp.	
B.923	CO	<i>Hellula undalis</i>	
B.924	CO	<i>Keiferia lycopersicella</i>	
B.925	CO	<i>Leucoptera scitella</i>	
B.926	CO	<i>Lithocollethis</i> spp.	20
B.927	CO	<i>Lobesia botrana</i>	
B.928	CO	<i>Ostrinia nubilalis</i>	
B.929	CO	<i>Pandemis</i> spp.	
B.930	CO	<i>Pectinophora gossyp.</i>	
B.931	CO	<i>Phyllocnistis citrella</i>	
B.932	CO	<i>Pieris</i> spp.	
B.933	CO	<i>Plutella xylostella</i>	25
B.934	CO	<i>Scirpophaga</i> spp.	
B.935	CO	<i>Sesamia</i> spp.	
B.936	CO	<i>Sparganothis</i> spp.	
B.937	CO	<i>Spodoptera</i> spp.	
B.938	CO	<i>Tortrix</i> spp.	
B.939	CO	<i>Trichoplusia ni</i>	30
B.940	CO	<i>Agrotis</i> spp.	
B.941	CO	<i>Anthonomus grandis</i>	
B.942	CO	<i>Curculio</i> spp.	
B.943	CO	<i>Diabrotica balteata</i>	
B.944	CO	<i>Leptinotarsa</i> spp.	
B.945	CO	<i>Lissorhoptrus</i> spp.	35
B.946	CO	<i>Otiorynchus</i> spp.	
B.947	CO	<i>Aleurothrixus</i> spp.	
B.948	CO	<i>Aleyrodes</i> spp.	
B.949	CO	<i>Aonidiella</i> spp.	
B.950	CO	<i>Aphididae</i> spp.	
B.951	CO	<i>Aphis</i> spp.	40
B.952	CO	<i>Bemisia tabaci</i>	
B.953	CO	<i>Empoasca</i> spp.	
B.954	CO	<i>Mycus</i> spp.	
B.955	CO	<i>Nephotettix</i> spp.	
B.956	CO	<i>Nilaparvata</i> spp.	
B.957	CO	<i>Pseudococcus</i> spp.	
B.958	CO	<i>Psylla</i> spp.	45
B.959	CO	<i>Quadrastipidiotus</i> spp.	
B.960	CO	<i>Schizaphis</i> spp.	
B.961	CO	<i>Trialeurodes</i> spp.	
B.962	CO	<i>Lynomyza</i> spp.	
B.963	CO	<i>Oscinella</i> spp.	
B.964	CO	<i>Phorbia</i> spp.	50
B.965	CO	<i>Frankliniella</i> spp.	
B.966	CO	<i>Thrips</i> spp.	
B.967	CO	<i>Scirtothrips aurantii</i>	
B.968	CO	<i>Aceria</i> spp.	
B.969	CO	<i>Aculus</i> spp.	
B.970	CO	<i>Brevipalpus</i> spp.	55
B.971	CO	<i>Panonychus</i> spp.	
B.972	CO	<i>Phyllocoptura</i> spp.	
B.973	CO	<i>Tetranychus</i> spp.	
B.974	CO	<i>Heterodera</i> spp.	
B.975	CO	<i>Meloidogyne</i> spp.	
B.976	CH	<i>Adoxophyes</i> spp.	60
B.977	CH	<i>Agrotis</i> spp.	
B.978	CH	<i>Alabama argillaceae</i>	
B.979	CH	<i>Anticarsia gemmatilis</i>	
B.980	CH	<i>Chilo</i> spp.	
B.981	CH	<i>Clysia ambiguella</i>	
B.982	CH	<i>Crocidolomia binotalis</i>	
B.983	CH	<i>Cydia</i> spp.	65
B.984	CH	<i>Diparopsis castanea</i>	

58

TABLE B-continued

	AP	Control of	
B.985	CH	<i>Earias</i> spp.	
B.986	CH	<i>Ephesia</i> spp.	
B.987	CH	<i>Heliothis</i> spp.	
B.988	CH	<i>Hellula undalis</i>	
B.989	CH	<i>Keiferia lycopersicella</i>	
B.990	CH	<i>Leucoptera scitella</i>	
B.991	CH	<i>Lithocollethis</i> spp.	
B.992	CH	<i>Lobesia botrana</i>	
B.993	CH	<i>Ostrinia nubilalis</i>	
B.994	CH	<i>Pandemis</i> spp.	
B.995	CH	<i>Pectinophora gossyp.</i>	
B.996	CH	<i>Phyllocnistis citrella</i>	
B.997	CH	<i>Pieris</i> spp.	
B.998	CH	<i>Plutella xylostella</i>	
B.999	CH	<i>Scirpophaga</i> spp.	
B.1000	CH	<i>Sesamia</i> spp.	
B.1001	CH	<i>Sparganothis</i> spp.	
B.1002	CH	<i>Spodoptera</i> spp.	
B.1003	CH	<i>Tortrix</i> spp.	
B.1004	CH	<i>Trichoplusia ni</i>	
B.1005	CH	<i>Agrotis</i> spp.	
B.1006	CH	<i>Anthonomus grandis</i>	
B.1007	CH	<i>Curculio</i> spp.	
B.1008	CH	<i>Diabrotica balteata</i>	
B.1009	CH	<i>Leptinotarsa</i> spp.	
B.1010	CH	<i>Lissorhoptrus</i> spp.	
B.1011	CH	<i>Otiorynchus</i> spp.	
B.1012	CH	<i>Aleurothrixus</i> spp.	
B.1013	CH	<i>Aleyrodes</i> spp.	
B.1014	CH	<i>Aonidiella</i> spp.	
B.1015	CH	<i>Aphididae</i> spp.	
B.1016	CH	<i>Aphis</i> spp.	
B.1017	CH	<i>Bemisia tabaci</i>	
B.1018	CH	<i>Empoasca</i> spp.	
B.1019	CH	<i>Mycus</i> spp.	
B.1020	CH	<i>Nephotettix</i> spp.	
B.1021	CH	<i>Nilaparvata</i> spp.	
B.1022	CH	<i>Pseudococcus</i> spp.	
B.1023	CH	<i>Psylla</i> spp.	
B.1024	CH	<i>Quadrastipidiotus</i> spp.	
B.1025	CH	<i>Schizaphis</i> spp.	
B.1026	CH	<i>Trialeurodes</i> spp.	
B.1027	CH	<i>Lynomyza</i> spp.	
B.1028	CH	<i>Oscinella</i> spp.	
B.1029	CH	<i>Phorbia</i> spp.	
B.1030	CH	<i>Frankliniella</i> spp.	
B.1031	CH	<i>Thrips</i> spp.	
B.1032	CH	<i>Scirtothrips aurantii</i>	
B.1033	CH	<i>Aceria</i> spp.	
B.1034	CH	<i>Aculus</i> spp.	
B.1035	CH	<i>Brevipalpus</i> spp.	
B.1036	CH	<i>Panonychus</i> spp.	
B.1037	CH	<i>Phyllocoptura</i> spp.	
B.1038	CH	<i>Tetranychus</i> spp.	
B.1039	CH	<i>Heterodera</i> spp.	
B.1040	CH	<i>Meloidogyne</i> spp.	
B.1041	SS	<i>Adoxophyes</i> spp.	
B.1042	SS	<i>Agrotis</i> spp.	
B.1043	SS	<i>Alabama argillaceae</i>	
B.1044	SS	<i>Anticarsia gemmatilis</i>	
B.1045	SS	<i>Chilo</i> spp.	
B.1046	SS	<i>Clysia ambiguella</i>	
B.1047	SS	<i>Crocidolomia binotalis</i>	
B.1048	SS	<i>Cydia</i> spp.	
B.1049	SS	<i>Diparopsis castanea</i>	
B.1050	SS	<i>Earias</i> spp.	
B.1051	SS	<i>Ephesia</i> spp.	
B.1052	SS	<i>Heliothis</i> spp.	
B.1053	SS	<i>Hellula undalis</i>	
B.1054	SS	<i>Keiferia lycopersicella</i>	
B.1055	SS	<i>Leucoptera scitella</i>	
B.1056	SS	<i>Lithocollethis</i> spp.	
B.1057	SS	<i>Lobesia botrana</i>	
B.1058	SS	<i>Ostrinia nubilalis</i>	
B.1059	SS	<i>Pandemis</i> spp.	
B.1060	SS	<i>Pectinophora gossyp.</i>	
B.1061	SS	<i>Phyllocnistis citrella</i>	

US 7,105,469 B2

59

TABLE B-continued

AP	Control of
B.1062	SS <i>Pieris</i> spp.
B.1063	SS <i>Plutella xylostella</i>
B.1064	SS <i>Scirpophaga</i> spp.
B.1065	SS <i>Sesamia</i> spp.
B.1066	SS <i>Sparganothis</i> spp.
B.1067	SS <i>Spodoptera</i> spp.
B.1068	SS <i>Tortrix</i> spp.
B.1069	SS <i>Trichoplusia ni</i>
B.1070	SS <i>Agrotis</i> spp.
B.1071	SS <i>Anthonomus grandis</i>
B.1072	SS <i>Curculio</i> spp.
B.1073	SS <i>Diabrotica balteata</i>
B.1074	SS <i>Leptinotarsa</i> spp.
B.1075	SS <i>Lissorhoptrus</i> spp.
B.1076	SS <i>Otiorynchus</i> spp.
B.1077	SS <i>Aleurothrixus</i> spp.
B.1078	SS <i>Aleyrodes</i> spp.
B.1079	SS <i>Aonidiella</i> spp.
B.1080	SS <i>Aphididae</i> spp.
B.1081	SS <i>Aphis</i> spp.
B.1082	SS <i>Bemisia tabaci</i>
B.1083	SS <i>Empoasca</i> spp.
B.1084	SS <i>Mycus</i> spp.
B.1085	SS <i>Nephotettix</i> spp.
B.1086	SS <i>Nilaparvata</i> spp.
B.1087	SS <i>Pseudococcus</i> spp.
B.1088	SS <i>Psylla</i> spp.
B.1089	SS <i>Quadrastipidiotus</i> spp.
B.1090	SS <i>Schizaphis</i> spp.
B.1091	SS <i>Trialeurodes</i> spp.
B.1092	SS <i>Lyriomyza</i> spp.
B.1093	SS <i>Oscinella</i> spp.
B.1094	SS <i>Phorbia</i> spp.
B.1095	SS <i>Frankliniella</i> spp.
B.1096	SS <i>Thrips</i> spp.
B.1097	SS <i>Scirtothrips aurantii</i>
B.1098	SS <i>Aceria</i> spp.
B.1099	SS <i>Aculus</i> spp.
B.1100	SS <i>Brevipalpus</i> spp.
B.1101	SS <i>Panonychus</i> spp.
B.1102	SS <i>Phyllocoptruta</i> spp.
B.1103	SS <i>Tetranychus</i> spp.
B.1104	SS <i>Heterodera</i> spp.
B.1105	SS <i>Meloidogyne</i> spp.
B.1106	HO <i>Adoxophyes</i> spp.
B.1107	HO <i>Agrotis</i> spp.
B.1108	HO <i>Alabama argillaceae</i>
B.1109	HO <i>Anticarsia gemmatilis</i>
B.1110	HO <i>Chilo</i> spp.
B.1111	HO <i>Clysia ambiguella</i>
B.1112	HO <i>Crocidolomia binotalis</i>
B.1113	HO <i>Cydia</i> spp.
B.1114	HO <i>Diparopsis castanea</i>
B.1115	HO <i>Earias</i> spp.
B.1116	HO <i>Ephestia</i> spp.
B.1117	HO <i>Heliothis</i> spp.
B.1118	HO <i>Hellula undalis</i>
B.1119	HO <i>Keiferia lycopersticella</i>
B.1120	HO <i>Leucoptera scitella</i>
B.1121	HO <i>Lithocolletis</i> spp.
B.1122	HO <i>Lobesia botrana</i>
B.1123	HO <i>Ostrinia nubilalis</i>
B.1124	HO <i>Pandemis</i> spp.
B.1125	HO <i>Pectinophora gossypiella</i>
B.1126	HO <i>Phyllocnistis citrella</i>
B.1127	HO <i>Pieris</i> spp.
B.1128	HO <i>Plutella xylostella</i>
B.1129	HO <i>Scirpophaga</i> spp.
B.1130	HO <i>Sesamia</i> spp.
B.1131	HO <i>Sparganothis</i> spp.
B.1132	HO <i>Spodoptera</i> spp.
B.1133	HO <i>Tortrix</i> spp.
B.1134	HO <i>Trichoplusia ni</i>
B.1135	HO <i>Agrotis</i> spp.
B.1136	HO <i>Anthonomus grandis</i>
B.1137	HO <i>Curculio</i> spp.
B.1138	HO <i>Diabrotica balteata</i>

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TABLE B-continued

AP	Control of
5 B.1139	HO <i>Leptinotarsa</i> spp.
B.1140	HO <i>Lissorhoptrus</i> spp.
B.1141	HO <i>Otiorynchus</i> spp.
B.1142	HO <i>Aleurothrixus</i> spp.
B.1143	HO <i>Aleyrodes</i> spp.
B.1144	HO <i>Aonidiella</i> spp.
10 B.1145	HO <i>Aphididae</i> spp.
B.1146	HO <i>Aphis</i> spp.
B.1147	HO <i>Bemisia tabaci</i>
B.1148	HO <i>Empoasca</i> spp.
B.1149	HO <i>Mycus</i> spp.
B.1150	HO <i>Nephotettix</i> spp.
15 B.1151	HO <i>Nilaparvata</i> spp.
B.1152	HO <i>Pseudococcus</i> spp.
B.1153	HO <i>Psylla</i> spp.
B.1154	HO <i>Quadrastipidiotus</i> spp.
B.1155	HO <i>Schizaphis</i> spp.
B.1156	HO <i>Trialeurodes</i> spp.
20 B.1157	HO <i>Lyriomyza</i> spp.
B.1158	HO <i>Oscinella</i> spp.
B.1159	HO <i>Phorbia</i> spp.
B.1160	HO <i>Frankliniella</i> spp.
B.1161	HO <i>Thrips</i> spp.
B.1162	HO <i>Scirtothrips aurantii</i>
25 B.1163	HO <i>Aceria</i> spp.
B.1164	HO <i>Aculus</i> spp.
B.1165	HO <i>Brevipalpus</i> spp.
B.1166	HO <i>Panonychus</i> spp.
B.1167	HO <i>Phyllocoptruta</i> spp.
B.1168	HO <i>Tetranychus</i> spp.
B.1169	HO <i>Heterodera</i> spp.
30 B.1170	HO <i>Meloidogyne</i> spp.

The following abbreviations are used in the table:

Active Principle of transgenic plant: AP

Photographus luminescens: PL*Xenorhabdus nematophilus*: XN

35 Proteinase Inhibitors: Plnh.

Plant lectins PLec.

Agglutinins: Aggl.

3-Hydroxysteroid oxidase: HO

Cholesterol oxidase: CO

Chitinase: CH

40 Glucanase: GL

Stilbensynthase SS

Biological Examples

45 Table 1: A method of controlling pests comprising the application of thiamethoxam to transgenic cotton, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

50 Table 2: A method of controlling pests comprising the application of thiamethoxam to transgenic rice, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

55 Table 3: A method of controlling pests comprising the application of thiamethoxam to transgenic potatoes, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

60 Table 4: A method of controlling pests comprising the application of thiamethoxam to transgenic *brassica*, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

61

Table 5: A method of controlling pests comprising the application of thiamethoxam to transgenic tomatoes, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 6: A method of controlling pests comprising the application of thiamethoxam to transgenic cucurbits, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 7: A method of controlling pests comprising the application of thiamethoxam to transgenic soybeans, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 8: A method of controlling pests comprising the application of thiamethoxam to transgenic maize, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 9: A method of controlling pests comprising the application of thiamethoxam to transgenic wheat, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 10: A method of controlling pests comprising the application of thiamethoxam to transgenic bananas, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 11: A method of controlling pests comprising the application of thiamethoxam to transgenic citrus trees, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 12: A method of controlling pests comprising the application of thiamethoxam to transgenic pome fruit trees, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 13: A method of controlling pests comprising the application of thiamethoxam to transgenic peppers, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 14: A method of controlling pests comprising the application of imidacloprid to transgenic cotton, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B

Table 15: A method of controlling pests comprising the application of imidacloprid to transgenic rice, wherein the combination of the active principle expressed by the transgenic plant and the pest to be controlled correspond to anyone of the individualised combinations B.1 to B.1170 of table B.

Table 16: A method of controlling pests comprising the application of imidacloprid to transgenic potatoes, wherein the combination of the active principle expressed by the

US 7,105,469 B2

65

TABLE C-continued

	Principle	Tolerant to	Crop
C.10	ALS	Sulfonylureas etc. ***	pome fruit
C.11	ALS	Sulfonylureas etc. ***	stone fruit
C.12	ALS	Sulfonylureas etc. ***	<i>citrus</i>
C.13	ACCCase	+++	Cotton
C.14	ACCCase	+++	Rice
C.15	ACCCase	+++	<i>Brassica</i>
C.16	ACCCase	+++	Potatoes
C.17	ACCCase	+++	Tomatoes
C.18	ACCCase	+++	Cucurbits
C.19	ACCCase	+++	Soybeans
C.20	ACCCase	+++	Maize
C.21	ACCCase	+++	Wheat
C.22	ACCCase	+++	pome fruit
C.23	ACCCase	+++	stone fruit
C.24	ACCCase	+++	<i>citrus</i>
C.25	HPPD	Isoxaflutol, Isoxachlotol, Sulcotrion, Mesotrion	Cotton
C.26	HPPD	Isoxaflutol, Isoxachlotol, Sulcotrion, Mesotrion	Rice
C.27	HPPD	Isoxaflutol, Isoxachlotol, Sulcotrion, Mesotrion	<i>Brassica</i>
C.28	HPPD	Isoxaflutol, Isoxachlotol, Sulcotrion, Mesotrion	Potatoes
C.29	HPPD	Isoxaflutol, Isoxachlotol, Sulcotrion, Mesotrion	Tomatoes
C.30	HPPD	Isoxaflutol, Isoxachlotol, Sulcotrion, Mesotrion	Cucurbits
C.31	HPPD	Isoxaflutol, Isoxachlotol, Sulcotrion, Mesotrion	Soybeans
C.32	HPPD	Isoxaflutol, Isoxachlotol, Sulcotrion, Mesotrion	Maize
C.33	HPPD	Isoxaflutol, Isoxachlotol, Sulcotrion, Mesotrion	Wheat
C.34	HPPD	Isoxaflutol, Isoxachlotol, Sulcotrion, Mesotrion	pome fruit
C.35	HPPD	Isoxaflutol, Isoxachlotol, Sulcotrion, Mesotrion	stone fruit
C.36	HPPD	Isoxaflutol, Isoxachlotol, Sulcotrion, Mesotrion	<i>citrus</i>
C.37	Nitrilase	Bromoxynil, Ioxynil	Cotton
C.38	Nitrilase	Bromoxynil, Ioxynil	Rice
C.39	Nitrilase	Bromoxynil, Ioxynil	<i>Brassica</i>
C.40	Nitrilase	Bromoxynil, Ioxynil	Potatoes
C.41	Nitrilase	Bromoxynil, Ioxynil	Tomatoes
C.42	Nitrilase	Bromoxynil, Ioxynil	Cucurbits
C.43	Nitrilase	Bromoxynil, Ioxynil	Soybeans
C.44	Nitrilase	Bromoxynil, Ioxynil	Maize
C.45	Nitrilase	Bromoxynil, Ioxynil	Wheat
C.46	Nitrilase	Bromoxynil, Ioxynil	pome fruit
C.47	Nitrilase	Bromoxynil, Ioxynil	stone fruit
C.48	Nitrilase	Bromoxynil, Ioxynil	<i>citrus</i>
C.49	IPS	Chloroactanilides &&&	Cotton
C.50	IPS	Chloroactanilides &&&	Rice
C.51	IPS	Chloroactanilide &&&s	<i>Brassica</i>
C.52	IPS	Chloroactanilides &&&	Potatoes
C.53	IPS	Chloroactanilides &&&	Tomatoes
C.54	IPS	Chloroactanilides &&&	Cucurbits
C.55	IPS	Chloroactanilides &&&	Soybeans
C.56	IPS	Chloroactanilides &&&	Maize
C.57	IPS	Chloroactanilides &&&	Wheat
C.58	IPS	Chloroactanilides &&&	pome fruit
C.59	IPS	Chloroactanilides &&&	stone fruit
C.60	IPS	Chloroactanilides &&&	<i>citrus</i>
C.61	HOM	2,4-D, Mecoprop-P	Cotton
C.62	HOM	2,4-D, Mecoprop-P	Rice
C.63	HOM	2,4-D, Mecoprop-P	<i>Brassica</i>
C.64	HOM	2,4-D, Mecoprop-P	Potatoes
C.65	HOM	2,4-D, Mecoprop-P	Tomatoes
C.66	HOM	2,4-D, Mecoprop-P	Cucurbits
C.67	HOM	2,4-D, Mecoprop-P	Soybeans
C.68	HOM	2,4-D, Mecoprop-P	Maize
C.69	HOM	2,4-D, Mecoprop-P	Wheat
C.70	HOM	2,4-D, Mecoprop-P	pome fruit
C.71	HOM	2,4-D, Mecoprop-P	stone fruit
C.72	HOM	2,4-D, Mecoprop-P	<i>citrus</i>
C.73	PROTOX	Protex inhibitors ///	Cotton
C.74	PROTOX	Protex inhibitors ///	Rice

66

TABLE C-continued

	Principle	Tolerant to	Crop
C.75	PROTOX	Protex inhibitors ///	<i>Brassica</i>
C.76	PROTOX	Protex inhibitors ///	Potatoes
C.77	PROTOX	Protex inhibitors ///	Tomatoes
C.78	PROTOX	Protex inhibitors ///	Cucurbits
C.79	PROTOX	Protex inhibitors ///	Soybeans
C.80	PROTOX	Protex inhibitors ///	Maize
C.81	PROTOX	Protex inhibitors ///	Wheat
C.82	PROTOX	Protex inhibitors ///	pome fruit
C.83	PROTOX	Protex inhibitors ///	stone fruit
C.84	PROTOX	Protex inhibitors ///	<i>citrus</i>
C.85	EPSPS	Glyphosate and/or Sulphosate	Cotton
C.86	EPSPS	Glyphosate and/or Sulphosate	Rice
C.87	EPSPS	Glyphosate and/or Sulphosate	<i>Brassica</i>
C.88	EPSPS	Glyphosate and/or Sulphosate	Potatoes
C.89	EPSPS	Glyphosate and/or Sulphosate	Tomatoes
C.90	EPSPS	Glyphosate and/or Sulphosate	Cucurbits
C.91	EPSPS	Glyphosate and/or Sulphosate	Soybeans
C.92	EPSPS	Glyphosate and/or Sulphosate	Maize
C.93	EPSPS	Glyphosate and/or Sulphosate	Wheat
C.94	EPSPS	Glyphosate and/or Sulphosate	pome fruit
C.95	EPSPS	Glyphosate and/or Sulphosate	stone fruit
C.96	EPSPS	Glyphosate and/or Sulphosate	<i>citrus</i>
C.97	GS	Gluphosinate and/or Bialaphos	Cotton
C.98	GS	Gluphosinate and/or Bialaphos	Rice
C.99	GS	Gluphosinate and/or Bialaphos	<i>Brassica</i>
C.100	GS	Gluphosinate and/or Bialaphos	Potatoes
C.101	GS	Gluphosinate and/or Bialaphos	Tomatoes
C.102	GS	Gluphosinate and/or Bialaphos	Cucurbits
C.103	GS	Gluphosinate and/or Bialaphos	Soybeans
C.104	GS	Gluphosinate and/or Bialaphos	Maize
C.105	GS	Gluphosinate and/or Bialaphos	Wheat
C.106	GS	Gluphosinate and/or Bialaphos	pome fruit
C.107	GS	Gluphosinate and/or Bialaphos	stone fruit
C.108	GS	Gluphosinate and/or Bialaphos	<i>citrus</i>

Abbreviations:

Acetyl-CoA Carboxylase: ACCase

Acetolactate Synthase: ALS

Hydroxyphenylpyruvate dioxygenase: HPPD

Inhibition of protein synthesis: IPS

Hormone mimic: HO

Glutamine Synthetase: GS

Protoporphyrinogen oxidase: PROTOX

5-Enolpyruvyl-3-Phosphoshikimate Synthase: EPSPS

*** Included are Sulfonylureas, Imidazolinones, Triazopyrimidines,

Dimethoxypyrimidines and N-Acylsulfonamides:

Sulfonylureas such as Chlorsulfuron, Chlorimuron, Ethamethsulfuron, Met-

sulfuron, Primisulfuron, Prosulfuron, Triasulfuron, Cinosulfuron, Triflusufu-

ruron, Oxasulfuron, Bensulfuron, Tribenuron, ACC 322140, Fluzasulfuron,

Ethoxysulfuron, Fluzasulfuron, Nicosulfuron, Rimsulfuron, Thifensulfu-

ruron, Pyrazosulfuron, Clopyrasulfuron, NC 330, Azimsulfuron, Imazosulfu-

ruron, Sulfosulfuron, Amidosulfuron, Flupyrasulfuron, CGA 362622

Imidazolinones such as Imazamethabenz, Imazaquin, Imazamethypyr,

Imazethapyr, Imazapyr and Imazamox;

Triazopyrimidines such as DE 511, Flumetsulam and Chloransulam;

Dimethoxypyrimidines such as Pyriothobac, Pyriminobac, Bispyribac and

Pyrbenzoxim.

+++ Tolerant to Diclofop-methyl, Fluazifop-P-butyl, Haloxyfop-P-methyl,

Haloxyfop-P-ethyl, Quizalafop-P-ethyl, clodinafop propargyl, fenoxaprop -

ethyl, - Tepraloxym, Alloxym, Sethoxydim, Cycloxydim, Cloproxy-

dim, Tralkoxydim, Butoxydim, Caloxydim, Clefoxydim, Clethodim,

&&& Chloroactanilides such as Alachlor Acetochlor, Dimethenamid

// Protex inhibitors: For instance diphenylethers such as Acifluorfen,

Aclonifen, Bifenox, Chlomitrofen, Ethoxyfen, Fluoroglycofen, Fomesafen,

Lactofen, Oxyfluorfen; Imides such as Azafenidin, Carfentrazone-ethyl,

Cinidon-ethyl, Flumiclorac-pentyl, Flumioxazin, Fluthiacetmethyl, Oxadi-

argyl, Oxadiazon, Pentoxazone, Sulfentrazone, Imides and others, such as

Flunipropyn, Flupropacil, Nipyraclofen and Thidiazimin; and further Flu-

zazolate and Pyraflufen-ethyl

Biological Examples

Table 49: A method of controlling representatives of the genus *Adoxophyes* comprising the application of thia-methoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by

67

the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 50: A method of controlling representatives of the genus *Agrotis* comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 51: A method of controlling *Alabama argillaceae* comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant, and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 52: A method of controlling *Anticarsia gemmatilis* comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 53: A method of controlling representatives of the genus *Chilo* comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 54: A method of controlling *Clysia ambigua* comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 55: A method of controlling representatives of the genus *Cnephlocrocis* comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 56: A method of controlling *Crocidolomia binotalis* comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 57: A method of controlling representatives of the genus *Cydia* comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 58: A method of controlling *Diparopsis castanea* comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 59: A method of controlling representatives of the genus *Earias* comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 60: A method of controlling representatives of the genus *Ephestia* comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the

73

the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 113: A method of controlling representatives of the genus *Heterodera* comprising the application of thia-methoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 115: A method of controlling *Mamestra brassica* comprising the application of thiamethoxam to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 117: A method of controlling representatives of the genus *Agrotis* comprising the application of imidacloprid to a herbitically resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 118: A method of controlling *Alabama argillaceae* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 119: A method of controlling *Anticarsia gemmatilis* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 120: A method of controlling representatives of the genus *Chilo* comprising the application of imidacloprid to a herbiticidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 121: A method of controlling *Clysia ambiguella* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 122: A method of controlling representatives of the genus *Cnephlocrocis* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 124: A method of controlling representatives of the genus *Cydia* comprising the application of imidacloprid to a hericidially resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 125: A method of controlling *Diparopsis castanea* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 126: A method of controlling representatives of the genus *Earias* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 127: A method of controlling representatives of the genus *Ephestia* comprising the application of imidacloprid to a hereditarily resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 128: A method of controlling representatives of the genus *Heliothis* of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 129: A method of controlling *Helhula undalis* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 130: A method of controlling *Keiferia lycopersicella* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 131: A method of controlling *Leucoptera scitella* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 132: A method of controlling representatives of the genus *Lithocolletis* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

75

Table 133: A method of controlling *Lobesia botrana* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 134: A method of controlling *Ostrinia nubilalis* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 135: A method of controlling representatives of the genus *Pandemis* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 136: A method of controlling *Pectinophora gossypiella* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 137: A method of controlling *Phyllocnistis citrella* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 138: A method of controlling representatives of the genus *Pieris* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 139: A method of controlling *Plutella xylostella* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 140: A method of controlling representatives of the genus *Scirpophaga* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 141: A method of controlling representatives of the genus *Sesamia* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 142: A method of controlling representatives of the genus *Sparganothis* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 143: A method of controlling representatives of the genus *Spodoptera* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

77

Table 155: A method of controlling representatives of the genus *Aonidiella* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 156: A method of controlling representatives of the family *Aphididae* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 157: A method of controlling representatives of the genus *Aphis* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to any one of the lines C.1 to C.108 of table C.

Table 158: A method of controlling *Bemisia tabaci* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 159: A method of controlling representatives of the genus *Empoasca* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 160: A method of controlling representatives of the genus *Mycus* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 161: A method of controlling representatives of the genus *Nephotettix* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 162: A method of controlling representatives of the genus *Nilaparvata* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 163: A method of controlling representatives of the genus *Pseudococcus* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 164: A method of controlling representatives of the genus *Psylla* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 165: A method of controlling representatives of the genus *Quadraspidiotus* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

79

Table 177: A method of controlling representatives of the genus *Panonychus* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 178: A method of controlling representatives of the genus *Phyllocoptruta* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 179: A method of controlling representatives of the genus *Tetranychus* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 180: A method of controlling representatives of the genus *Heterodera* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 181: A method of controlling representatives of the genus *Meloidogyne* comprising the application of imidacloprid to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 182: A method of controlling representatives of the genus *Adoxophyes* comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to any one of the lines C.1 to C.108 of table C.

Table 183: A method of controlling representatives of the genus *Agrotis* comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 184: A method of controlling *Alabama argillaceae* comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 185: A method of controlling *Anticarsia gemmatilis* comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 186: A method of controlling representatives of the genus *Chilo* comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to any one of the lines C.1 to C.108 of table C.

Table 187: A method of controlling *Clysia ambigua* comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

81

Table 199: A method of controlling *Ostrinia nubilalis* comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 200: A method of controlling representatives of the genus *Pandemis* comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

83

Table 221: A method of controlling representatives of the family *Aphididae* comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 222: A method of controlling representatives of the genus *Aphis* comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 223: A method of controlling *Bemisia tabaci* comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 224: A method of controlling representatives of the genus *Empoasca* comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 225: A method of controlling representatives of the genus *Mycus* comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 226: A method of controlling representatives of the genus *Nephotettix* comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 227: A method of controlling representatives of the genus *Nilaparvata* comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 228: A method of controlling representatives of the genus *Pseudococcus* comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 229: A method of controlling representatives of the genus *Psylla* comprising the application of Ti-435 to a hericidially resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 230: A method of controlling representatives of the genus *Quadraspidiotus* comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 231: A method of controlling representatives of the genus *Schizaphis* comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

US 7,105,469 B2

85

Table 243: A method of controlling representatives of the genus *Phyllocoptruta* comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 244: A method of controlling representatives of the genus *Tetranychus* comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 245: A method of controlling representatives of the genus *Heterodera* comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 246: A method of controlling representatives of the genus *Meloidogyne* comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Table 247: A method of controlling *Mamestra brassica* comprising the application of Ti-435 to a herbicidally resistant transgenic crop, wherein the combination of the active principle expressed by the transgenic plant and the crop to be protected against the pest correspond to anyone of the lines C.1 to C.108 of table C.

Example B1

Action Against *Anthonomus grandis* adults,
Spodoptera littoralis or *Heliothis virescens*

Young transgenic cotton plants which express the δ -endotoxin CryIIIA are sprayed with an aqueous emulsion spray mixture comprising 100, 50, 10, 5, 1 ppm of imidacloprid respectively. After the spray coating has dried on, the cotton plants are populated with 10 adult *Anthonomus grandis*, 10 *Spodoptera littoralis* larvae or 10 *Heliothis virescens* larvae respectively and introduced into a plastic container. Evaluation takes place 3 to 10 days later. The percentage reduction in population, or the percentage reduction in feeding damage (% action), is determined by comparing the number of dead beetles and the feeding damage on the transgenic cotton plants with that of non-transgenic cotton plants which have been treated with an emulsion spray mixture comprising imidacloprid and conventional CryIIIA-toxin at a concentration of in each case 100, 50, 10, 5, 1 ppm respectively.

In this test, the control of the tested insects in the transgenic plant is superior to the control on the non-transgenic plant.

Example B2

Action Against *anthonomus grandis* adults,
spodoptera littoralis or *heliothis virescens*

Young transgenic cotton plants which express the δ -endotoxin CryIIIA are sprayed with an aqueous emulsion spray mixture comprising 100, 50, 10, 5, 1 ppm of thiamethoxam respectively. After the spray coating has dried on, the cotton plants are populated with 10 adult *Anthonomus grandis*, 10 *Spodoptera littoralis* larvae or 10 *Heliothis virescens* larvae respectively and introduced into a plastic container. Evaluation

86

takes place 3 to 10 days later. The percentage reduction in population, or the percentage reduction in feeding damage (% action), is determined by comparing the number of dead beetles and the feeding damage on the transgenic cotton plants with that of non-transgenic cotton plants which have been treated with an emulsion spray mixture comprising thiamethoxam and conventional CryIIIA-toxin at a concentration of in each case 100, 50, 10, 5, 1 ppm respectively.

In this test, the control of the tested insects in the transgenic plant is superior, while it is insufficient in the non-transgenic plant.

Example B3

Action Against *Anthonomus grandis* adults,
Spodoptera littoralis or *Heliothis virescens*

Young transgenic cotton plants which express the δ -endotoxin CryIIIA are sprayed with an aqueous emulsion spray mixture comprising 100, 50, 10, 5, 1 ppm of Ti-435 respectively. After the spray coating has dried on, the cotton plants are populated with 10 adult *Anthonomus grandis*, 10 *Spodoptera littoralis* larvae or 10 *Heliothis virescens* larvae respectively and introduced into a plastic container. Evaluation takes place 3 to 10 days later. The percentage reduction in population, or the percentage reduction in feeding damage (% action), is determined by comparing the number of dead beetles and the feeding damage on the transgenic cotton plants with that of non-transgenic cotton plants which have been treated with an emulsion spray mixture comprising Ti-435 and conventional CryIIIA-toxin at a concentration of in each case 100, 50, 10, 5, 1 ppm respectively.

In this test, the control of the tested insects in the transgenic plant is superior, while it is insufficient in the non-transgenic plant.

Example B4

Action Against *Anthonomus grandis* adults,
Spodoptera littoralis or *Heliothis virescens*

Young transgenic cotton plants which express the δ -endotoxin CryIIA(c) are sprayed with an aqueous emulsion spray mixture comprising 100, 50, 10, 5, 1 ppm of Ti-435 respectively. After the spray coating has dried on, the cotton plants are populated with 10 adult *Anthonomus grandis*, 10 *Spodoptera littoralis* larvae or 10 *Heliothis virescens* larvae respectively and introduced into a plastic container. Evaluation takes place 3 to 10 days later. The percentage reduction in population, or the percentage reduction in feeding damage (% action), is determined by comparing the number of dead beetles and the feeding damage on the transgenic cotton plants with that of non-transgenic cotton plants which have been treated with an emulsion spray mixture comprising Ti-435 and conventional CryIIIA-toxin at a concentration of in each case 100, 50, 10, 5, 1 ppm respectively.

In this test, the control of the tested insects in the transgenic plant is superior, while it is insufficient in the non-transgenic plant.

Example B5

Action Against *Anthonomus grandis* adults,
Spodoptera littoralis or *Heliothis virescens*

Young transgenic cotton plants which express the δ -endotoxin CryIIA(c) are sprayed with an aqueous emulsion

US 7,105,469 B2

87

spray mixture comprising 100, 50, 10, 5, 1 ppm of thiamethoxam respectively. After the spray coating has dried on, the cotton plants are populated with 10 adult *Anthonomus grandis*, 10 *Spodoptera littoralis* larvae or 10 *Heliothis virescens* larvae respectively and introduced into a plastic container. Evaluation takes place 3 to 10 days later. The percentage reduction in population, or the percentage reduction in feeding damage (% action), is determined by comparing the number of dead beetles and the feeding damage on the transgenic cotton plants with that of non-transgenic cotton plants which have been treated with an emulsion spray mixture comprising thiamethoxam and conventional CryIIIA-toxin at a concentration of in each case 100, 50, 10, 5, 1 ppm respectively.

In this test, the control of the tested insects in the transgenic plant is superior, while it is insufficient in the non-transgenic plant.

Example B6

Action Against *Anthonomus grandis* adults,
Spodoptera littoralis or *Heliothis virescens*

Young transgenic cotton plants which express the δ -endotoxin CryIa(c) are sprayed with an aqueous emulsion spray mixture comprising 100, 50, 10, 5, 1 ppm of imidacloprid respectively. After the spray coating has dried on, the cotton plants are populated with 10 adult *Anthonomus grandis*, 10 *Spodoptera littoralis* larvae or 10 *Heliothis virescens* larvae respectively and introduced into a plastic container. Evaluation takes place 3 to 10 days later. The percentage reduction in population, or the percentage reduction in feeding damage (% action), is determined by comparing the number of dead beetles and the feeding damage on the transgenic cotton plants with that of non-transgenic cotton plants which have been treated with an emulsion spray mixture comprising imidacloprid conventional CryIIIA-toxin at a concentration of in each case 100, 50, 10, 5, 1 ppm respectively.

In this test, the control of the tested insects in the transgenic plant is superior, while it is insufficient in the non-transgenic plant.

Example B7

Action Against *Ostrinia nubilalis*, *Spodoptera* spp.
or *Heliothis* spp.

A plot (a) planted with maize cv. KnockOut® and an adjacent plot (b) of the same size which is planted with conventional maize, both showing natural infestation with *Ostrinia nubilalis*, *Spodoptera* spp. or *Heliothis*, are sprayed with an aqueous emulsion spray mixture comprising 200, 100, 50, 10, 5, 1 ppm of Ti-435. Immediately afterwards, plot (b) is treated with an emulsion spray mixture comprising 200, 100, 50, 10, 5, 1 ppm of the endotoxin expressed by KnockOut®. Evaluation takes place 6 days later. The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of plot (a) with that on the plants of plot (b).

Improved control of *Ostrinia nubilalis*, *Spodoptera* spp. or *Heliothis* is observed on the plants of plot (a), while plot (b) shows a control level of not over 60%.

88

Example B8

Action Against *Ostrinia nubilalis*, *Spodoptera* spp.
or *Heliothis* spp.

A plot (a) planted with maize cv. KnockOut® and an adjacent plot (b) of the same size which is planted with conventional maize, both showing natural infestation with *Ostrinia nubilalis*, *Spodoptera* spp. or *Heliothis*, are sprayed with an aqueous emulsion spray mixture comprising 200, 100, 50, 10, 5, 1 ppm of thiamethoxam. Immediately afterwards, plot (b) is treated with an emulsion spray mixture comprising 200, 100, 50, 10, 5, 1 ppm of the endotoxin expressed by KnockOut®. Evaluation takes place 6 days later. The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of plot (a) with that on the plants of plot (b).

Improved control of *Ostrinia nubilalis*, *Spodoptera* spp. or *Heliothis* is observed on the plants of plot (a), while plot (b) shows a control level of not over 60%.

Example B9

Action Against *Ostrinia nubilalis*, *Spodoptera* spp.
or *Heliothis* spp.

A plot (a) planted with maize cv. KnockOut® and an adjacent plot (b) of the same size which is planted with conventional maize, both showing natural infestation with *Ostrinia nubilalis*, *Spodoptera* spp. or *Heliothis*, are sprayed with an aqueous emulsion spray mixture comprising 200, 100, 50, 10, 5, 1 ppm of imidacloprid. Immediately afterwards, plot (b) is treated with an emulsion spray mixture comprising 200, 100, 50, 10, 5, 1 ppm of the endotoxin expressed by KnockOut®. Evaluation takes place 6 days later. The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of plot (a) with that on the plants of plot (b).

Improved control of *Ostrinia nubilalis*, *Spodoptera* spp. or *Heliothis* spp. is observed on the plants of plot (a), while plot (b) shows a control level of not over 60%.

Example B10

Action Against *Diabrotica balteata*

A plot (a) planted with maize seedlings cv. KnockOut® and an adjacent plot (b) of the same size which is planted with conventional maize are sprayed with an aqueous emulsion of a spray mixture comprising 400 ppm thiamethoxam. Immediately afterwards, plot (b) is treated with an emulsion spray mixture comprising 400 ppm of the endotoxin expressed by KnockOut®. After the spray coating has dried on, the seedlings are populated with 10 *Diabrotica balteata* larvae in the second stage and transferred to a plastic container. The test is evaluated 6 days later. The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of plot (a) with that on the plants of plot (b).

Improved control of *Diabrotica balteata* is observed on the plants of plot (a), while plot (b) shows a control level of not over 60%.

US 7,105,469 B2

89

Example B11

Action Against *Aphis gossypii*

Cotton seedlings on a plot (a) expressing the δ -endotoxin CryIIa on a plot (a) and conventional cotton seedlings on a plot (b) are infected with *Aphis gossypii* and subsequently sprayed with a spray mixture comprising 400 ppm thiamethoxam. Immediately afterwards, plot (b) is treated with an emulsion spray mixture comprising 400 ppm of the δ -endotoxin CryIIa. The seedlings of plot (a) and (b) are then incubated at 20° C. The test is evaluated after 3 and 6 days.

The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of plot (a) with that on the plants of plot (b). Improved control of *Aphis gossypii* is observed on the plants of plot (a), while plot (b) shows a control level of not over 60%.

Example B12

Action Against *Frankliniella occidentalis*

Cotton seedlings expressing the δ -endotoxin CryIIa on a plot (a) and conventional cotton seedlings on a plot (b) are infected with *Frankliniella occidentalis* and subsequently sprayed with a spray mixture comprising 400 ppm thiamethoxam. Immediately afterwards, plot (b) is treated with an emulsion spray mixture comprising 400 ppm of the δ -endotoxin CryIIa. The seedlings of plot (a) and (b) are then incubated at 20° C. The test is evaluated after 3 and 6 days.

The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of plot (a) with that on the plants of plot (b). Improved control of *Frankliniella occidentalis* is observed on the plants of plot (a), while plot (b) shows a control level of not over 60%.

Example B13

Action Against *Aphis gossypii*

Cotton seedlings expressing the δ -endotoxin CryIA(c) on a plot (a) and conventional cotton seedlings on a plot (b) are infected with *Aphis gossypii* and subsequently sprayed with a spray mixture comprising 400 ppm thiamethoxam. Immediately afterwards, plot (b) is treated with an emulsion spray mixture comprising 400 ppm of the δ -endotoxin CryIIa. The seedlings of plot (a) and (b) are then incubated at 20° C. The test is evaluated after 3 and 6 days.

The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of plot (a) with that on the plants of plot (b). Improved control of *Aphis gossypii* is observed on the plants of plot (a), while plot (b) shows a control level of not over 60%.

Example B14

Action Against *Frankliniella occidentalis*

Cotton seedlings expressing the δ -endotoxin CryIA(c) on a plot (a) and conventional cotton seedlings on a plot (b) are infected with *Frankliniella occidentalis* and subsequently sprayed with a spray mixture comprising 400 ppm thiamethoxam. Immediately afterwards, plot (b) is treated with an emulsion spray mixture comprising 400 ppm of the

90

δ -endotoxin CryIA(c). The seedlings of plot (a) and (b) are then incubated at 20° C. The test is evaluated after 3 and 6 days.

The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of plot (a) with that on the plants of plot (b). Improved control of *Frankliniella occidentalis* is observed on the plants of plot (a), while plot (b) shows a control level of not over 60%.

Example B15

Action Against *Nephotettix cincticeps*

Rice plants on a plot (a) expressing the δ -endotoxin CryIA(b) and conventional rice plants on a plot (b) are sprayed with a spray mixture comprising 400 ppm thiamethoxam. Immediately afterwards, plot (b) is treated with an emulsion spray mixture comprising 400 ppm of the δ -endotoxin CryIA(b). After the spray coating has dried on, the plants are infected with *Nephotettix cincticeps* of the 2nd and 3rd stages. The seedlings of plot (a) and (b) are then incubated at 20° C. The test is evaluated after 21 days.

The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of plot (a) with that on the plants of plot (b). Improved control of *Nephotettix cincticeps* is observed on the plants of plot (a), while plot (b) shows a control level of not over 60%.

Example B16

Action Against *Nephotettix cincticeps* (systemic)

Rice plants expressing the δ -endotoxin CryIA(b) are planted in a in pot (A) and conventional ice plants are planted in a pot (B). Pot (A) is placed in an aqueous emulsion containing 400 ppm thiamethoxam, whereas plot (B) is placed in a pot containing 400 ppm thiamethoxam and 400 ppm of the δ -endotoxin CryI(b). The plants are subsequently infected with *Nephotettix cincticeps* larvae of the second and third stage. The test is evaluated after 6 days.

The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of pot (A) with that on the plants of pot (B). Improved control of *Nephotettix cincticeps* is observed on the plants of pot (A), while pot (B) shows a control level of not over 60%.

Example B17

Action Against *Nilaparvata lugens*

Rice plants on a plot (a) expressing the δ -endotoxin CryIA(b) and conventional rice plants on a plot (b) are infected with *Nilaparvata lugens*, subsequently sprayed with a spray mixture comprising 400 ppm thiamethoxam. Immediately afterwards, plot (b) is treated with an emulsion spray mixture comprising 400 ppm of the δ -endotoxin CryIA(b). The seedlings of plot (a) and (b) are then incubated at 20° C. The test is evaluated after 21 days.

The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of plot (a) with that on the plants of plot (b). Improved control of *Nilaparvata lugens* is observed on the plants of plot (a), while plot (b) shows a control level of not over 60%.

US 7,105,469 B2

91

Example B18

Action Against *Nilaparvata lugens* (Systemic)

Rice plants expressing the 6-endotoxin CryIA(b) are planted in a in pot (A) and conventional rice plants are planted in a pot (B). Pot (A) is placed in an aqueous emulsion containing 400 ppm thiamethoxam, whereas plot (B) is place in a pot copntaining 400 ppm thiamethoxam and 400 ppm of the 6-endotoxin CryIA(b). The plants are subsequently infected with *Nilaparvata lugens* larvae of the second and third stage. The test is evaluated after 6 days.

The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of pot (A) with that on the plants of pot (B). Improved control of *Nephotettix cincticeps* is observed on the plants of pot (A), while pot (B) shows a control level of not over 60%.

Example B19

Action Against *Nephotettix cincticeps*

Rice plants on a plot (a) expressing the 6-endotoxin CryIA(c) and conventional rice plants on a plot (b) are sprayed with a spray mixture comprising 400 ppm thiamethoxam. Immediately afterwards, plot (b) is treated with an emulsion spray mixture comprising 400 ppm of the δ -endotoxin CryIA(c). After the spray coating has dried on, the plants are infected with *Nephotettix cincticeps* of the 2nd and 3rd stages. The seedlings of plot (a) and (b) are then incubated at 20° C. The test is evaluated after 21 days.

The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of plot (a) with that on the plants of plot (b). Improved control of *Nephotettix cincticeps* is observed on the plants of plot (a), while plot (b) shows a control level of not over 60%.

Example B20

Action Against *Nephotettix cincticeps* (Systemic)

Rice plants expressing the 6-endotoxin CryIA(c) are planted in a in pot (A) and conventional ice plants are planted in a pot (B). Pot (A) is placed in an aqueous emulsion containing 400 ppm thiamethoxam, whereas plot (B) is placed in a pot containing 400 ppm thiamethoxam and 400 ppm of the 6-endotoxin CryI(c). The plants are subsequently infected with *Nephotettix cincticeps* larvae of the second and third stage. The test is evaluated after 6 days.

The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of pot (A) with that on the plants of pot (B). Improved control of *Nephotettix cincticeps* is observed on the plants of pot (A), while pot (B) shows a control level of not over 60%.

Example B21

Action Against *Nilaparvata lugens*

Rice plants on a plot (a) expressing the 6-endotoxin CryIA(c) and conventional rice plants on a plot (b) are infected with *Nilaparvata lugens*, subsequently sprayed with

92

a spray mixture comprising 400 ppm thiamethoxam. Immediately afterwards, plot (b) is treated with an emulsion spray mixture comprising 400 ppm of the 6-endotoxin CryIA(c). The seedlings of plot (a) and (b) are then incubated at 20° C. The test is evaluated after 21 days.

The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of plot (a) with that on the plants of plot (b). Improved control of *Nilaparvata lugens* is observed on the plants of plot (a), while plot (b) shows a control level of not over 0%.

Example B22

Action Against *Nilaparvata lugens* (Systemic)

Rice plants expressing the δ -endotoxin CryIA(c) are planted in a in pot (A) and conventional rice plants are planted in a pot (B). Pot (A) is placed in an aqueous emulsion containing 400 ppm thiamethoxam, whereas plot (B) is place in a pot copntaining 400 ppm thiamethoxam and 400 ppm of the 6-endotoxin CryIA(c). The plants are subsequently infected with *Nilaparvata lugens* larvae of the second and third stage. The test is evaluated after 6 days.

The percentage reduction in population (% action) is determined by comparing the number of dead pests on the plants of pot (A) with that on the plants of pot (B). Improved control of *Nephotettix cincticeps* is observed on the plants of pot (A), while pot (B) shows a control level of not over 60%.

The invention claimed is:

1. A method of controlling pests in crops of transgenic useful plants comprising the application of a composition comprising clothioanidin, in free form or in agrochemically useful salt form as active ingredient and at least one auxiliary to the pests, or the transgenic plant or propagation material thereof.

2. The method of claim 1 where the transgenic useful plant contains one or more genes which encode insecticidal resistance and express one or more active toxins.

3. The method of claim 2 wherein the active toxin expressed by the transgenic plant is selected from *Bacillus cereus* proteins, *Bacillus poplia* proteins, *Bacillus thuringiensis* endotoxins (B.t.), insecticidal proteins of bacteria colonising nematodes, proteinase inhibitors, ribosome inactivating proteins, plant lectins, animal toxins, and steroid metabolism enzymes.

4. The method of claim 2 wherein the active toxin expressed by the transgenic plant is selected from CryIA(a), CryIA(b), CryIA(c), Cry IIA, CryIIIA, CryIIIB2, CytA, VIP3, GL, PL, XN, Plnh., Plec., Aggl., CO, CH, SS, and HO.

5. The method of claim 1 where the crops of transgenic useful plants are selected from cotton, rice, potatoes, *brassica*, tomatoes, cucurbits, soybeans, maize, wheat, bananas, citrus trees, pome fruit trees and peppers.

6. The method of claim 1 wherein the composition is applied to the transgenic useful plant.

7. The method of claim 1 wherein clothioanidin is applied to the propagation material of the transgenic useful plant.

8. The method of claim 7 wherein the propagation material is seed.

* * * * *